

Loss-of-Function of *Constitutive Expresser of Pathogenesis Related Genes5* Affects Potassium Homeostasis in *Arabidopsis thaliana*

Monica Borghi^{1a}, Ana Rus^{1b}, David E. Salt^{1*2c}

Department of Horticulture and Landscape Architecture, Purdue University, West Lafayette, Indiana, United States of America

Abstract

Here, we demonstrate that the reduction in leaf K^+ observed in a mutant previously identified in an ionic screen of fast neutron mutagenized *Arabidopsis thaliana* is caused by a loss-of-function allele of *CPR5*, which we name *cpr5-3*. This observation establishes low leaf K^+ as a new phenotype for loss-of-function alleles of *CPR5*. We investigate the factors affecting this low leaf K^+ in *cpr5* using double mutants defective in salicylic acid (SA) and jasmonic acid (JA) signalling, and by gene expression analysis of various channels and transporters. Reciprocal grafting between *cpr5* and Col-0 was used to determine the relative importance of the shoot and root in causing the low leaf K^+ phenotype of *cpr5*. Our data show that loss-of-function of *CPR5* in shoots primarily determines the low leaf K^+ phenotype of *cpr5*, though the roots also contribute to a lesser degree. The low leaf K^+ phenotype of *cpr5* is independent of the elevated SA and JA known to occur in *cpr5*. In *cpr5* expression of genes encoding various *Cyclic Nucleotide Gated Channels* (CNGCs) are uniquely elevated in leaves. Further, expression of *HAK5*, encoding the high affinity K^+ uptake transporter, is reduced in roots of *cpr5* grown with high or low K^+ supply. We suggest a model in which low leaf K^+ in *cpr5* is driven primarily by enhanced shoot-to-root K^+ export caused by a constitutive activation of the expression of various CNGCs. This activation may enhance K^+ efflux, either indirectly via enhanced cytosolic Ca^{2+} and/or directly by increased K^+ transport activity. Enhanced shoot-to-root K^+ export may also cause the reduced expression of *HAK5* observed in roots of *cpr5*, leading to a reduction in uptake of K^+ . All ionic data presented is publically available at www.ionomicshub.org.

Citation: Borghi M, Rus A, Salt DE (2011) Loss-of-Function of *Constitutive Expresser of Pathogenesis Related Genes5* Affects Potassium Homeostasis in *Arabidopsis thaliana*. PLoS ONE 6(10): e26360. doi:10.1371/journal.pone.0026360

Editor: David Edward Somers, Ohio State University, United States of America

Received: June 30, 2011; **Accepted:** September 25, 2011; **Published:** October 27, 2011

Copyright: © 2011 Borghi et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by grants to D.E.S. from the US National Science Foundation Arabidopsis 2010 and Plant Genome Research programs (IOS 0419695 and DBI 0701119). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: david.salt@abdn.ac.uk

^{1a} Current address: Department of Plant Pathology, North Carolina State University, Raleigh, North Carolina, United States of America

^{1b} Current address: Department of Biological Sciences, California State University Long Beach, Long Beach, California, United States of America

^{1c} Current address: Institute of Biological and Environmental Sciences, University of Aberdeen, Aberdeen, United Kingdom

Introduction

Potassium (K^+) is a macronutrient essential for normal plant growth and development. It participates in numerous physiological processes including regulation of enzyme activity, cell expansion, stomata movement and defence towards pathogens [1,2]. Numerous mechanisms are known to be involved in K^+ homeostasis, and many K^+ channels and transporters have been identified. In plants K^+ is highly mobile. Its movement into the xylem is driven by transpiration, and in the phloem by the specific requirements of tissues and organs [3,4]. Environmental conditions also regulate the movement of K^+ . For example, under drought stress the concentration of K^+ in the xylem decreases, possibly because of the closure of stomatal pores induced by abscisic acid (ABA) [5].

The effects of low K^+ have been investigated in some depth, and it has emerged that jasmonic acid (JA) plays a central role in the response of plants to K^+ shortage [6,7]. Nevertheless, whether the application of K^+ is beneficial or not in conferring resistance towards pathogens is controversial [8]. At the molecular level the role of K^+ is clearer. Plant-pathogen interactions cause an increase

in cytosolic Ca^{2+} triggering anion channel activation, plasma membrane depolarization, activation of K^+ permeable efflux channels leading to enhanced K^+ efflux and the initiation of the hypersensitive response (HR) [9,10].

In this study, we describe the characterization of an *A. thaliana* mutant with a 10–30% reduction in leaf K^+ which was previously identified in an ionic screen of fast neutron mutagenized plants [11]. Genetic analysis revealed this mutant to be a new null allele of *CPR5*, a gene originally identified in two independent screens for altered response to pathogens [12,13]. *cpr5* has a high content of salicylic acid (SA) and shows constitutive expression of pathogenesis related genes (*PR*), as well as plant defensin *PDF1.2* which is a marker of the JA-dependent pathway. Therefore, it has been suggested the *CPR5* is a negative regulator of local defence response to pathogens [14]. *CPR5* is also implicated in cell senescence [15–17], cell proliferation and trichome development [18], cell wall biogenesis [19] redox balance [20], and water relations via enhanced ABA sensitivity [21]. Here, we show that *CPR5* is also associated with K^+ homeostasis possibly via modulation of expression of various CNGCs and *HAK5*.

Materials and Methods

Plant growth and mutant screening

Fast neutron-mutagenized M2 *A. thaliana* seeds were purchased from Lehle Seeds (Round Rock, TX) and plants screened for their leaf elemental profile by ICP-MS [11]. Seeds of Col-0 (CS6000) and *cpr5-2* (CS3770) were provided by the Arabidopsis Biological Resource Center (The Ohio State University).

For non axenic conditions, plants were grown in pots containing moist soil (Scotts Potting Medium, Scotts-Sierra Horticultural Products Company, Marysville, OH) in a climate-controlled room (temperature 19–22°C, day-night; humidity 60%; photoperiod 10–14 hours light-dark; light intensity 100±10 μmol m⁻² sec⁻¹) and bottom watered at regular intervals with a solution containing 0.25× Hoagland's macro and micronutrients [11].

For plants grown in axenic conditions, surface sterilized seeds were stratified at 4°C in the dark for five days before sowing. To measure the expression of *HAK5* and *AKT1* plants were grown for 2 weeks on solidified medium containing 1/20th MS salts accordingly to Cheong et al., [22] and containing 20 mM or 100 μM of KCl, and in a second experiment on a minimal medium without NH₄⁺ [23] with 10 g L⁻¹ UltraPure sucrose (Sigma) and solidified with 10 g L⁻¹ pure agarose (Molecular Biology Grade, Research Products International Corp.), which contains negligible amounts of K⁺ (approximately 8 μg Kg⁻¹ as determined by ICP-MS analysis). K⁺ was added as KCl at the final concentration of 10, 50 and 100 μM and the pH adjusted to 5.8 with Ca(OH)₂. K⁺ content in root and shoot was measured in plants grown for 2 weeks on solidified medium containing 1/20th MS salts accordingly to Cheong et al., [22] and containing 20 mM of KCl.

Genetic analysis

cpr5-3 was outcrossed to *A. thaliana Landsberg erecta* (Ler-0) accession, F1 seeds were planted on 0.5× MS medium solidified with 10 g L⁻¹ of agar and seedlings visually screened for hyponastic and early yellowing cotyledons. All seedlings of F1 generation looked wild-type and were transferred to soil and grown to produce F2 seeds. F2 plants grown on soil were visually selected for small size and yellow early senescing leaves and used to map the mutation with a positional cloning approach with single strand length polymorphism (SSLP) markers.

Quantitative real-time PCR

Total RNA was extracted with the Qiagen RNeasyPlant Mini Kit (<http://www.qiagen.com>) from five weeks old plants grown on soil or from two week old plants grown on plates. DNase digestion on column was performed to eliminate possible contamination with DNA. Two micrograms of total RNA were used as a template to synthesize first-strand cDNA with SuperScript VILO cDNA Synthesis Kit (InvitrogenLife Technologies, <http://www.invitrogen.com>). Quantitative real-time PCR was performed with the SYBR Green reagent mix in a StepOnePlus instrument according to the manufacturer's instructions (Applied Biosystems, California, USA). The expression of K⁺ channel/transporter genes was detected with the following set of primers: *HAK5*, 147 (5'-TGCTGATCTAGGTCACCTTCAGTGTT-3') and 148 (5'-AAAGCAGGATATGCGACACATG-3'); *AKT1*, 170 (5'-TCTAAATTGTGTTCTTCTTCTGTTGGA-3') and 171 (5'-CCTTCCGC-GTCTCTGCAA-3'); *CNGC19*, 137 (5'-TTCTCACTTGGTGCCTCTCTTCT-3') and 138 (5'-AATCCCTTTGGTGGCATCTTT-3'); *CNGC10*, 139 (5'-TCATCATTGATCTACTCTCTATCCTTCCCT-3') and 140 (5'-TGGTTGACGCTTGGAA-TAACG-3'); *CNGC12*, 156 (5'-CAACGAACATTCAGGTTA-

TACTCACA-3') and 157 (5'-GCCGCTTGAATGAAGAATGC-3'); *CNGC20*, 160 (5'-TGCCTCGAACGCTCTTCTG-3') and 161 (5'-CCTTTGATGGCATCCTTATCCT-3'); *CNGC11*, 196 (5'-GATAAAAACATGAATCTTCAGAGGAGAA-3') and 197 (5'-CTAACACTTTTCAATTTTCCATCAACTC-3'). As an internal reference the expression of *PP2A* (At1g13320) was used since it showed stable expression throughout the experimental series of development, shoot and root abiotic stress, hormones, nutrient stress, light and biotic stress [24], all of which are affected by loss-of-function of *CPR5*. In addition to *PP2A* the expression of *HAK5* and *AKT1* was also normalized to *UBQ10* (At4g05320). *UBQ10* also shows stable expression in *cpr5*. The average value from real-time PCR measurements from at least three independent biological replicates was used to evaluate transcript abundance. Biological replicates were composed of tissues harvested from between 5 and 10 plants for analysis of shoot tissue, and between 25 and 30 plants for analysis of root tissue, from plants grown on plates. For plants grown in soil 2–3 leaves from at least 3 individual plants were used. Steady state mRNA levels were calculated relative to a reference gene and presented based on the 2^{-ΔΔCt} method [25].

Determination of the elemental content of plant tissues

Plants grown in soil were non-destructively sampled by removing 1–2 leaves (approximately 3 mg dry weight), rinsed with 18 MΩ water, placed into Pyrex digestion tubes and dried at 92°C for 20 hours. Alternatively, shoots and roots were harvested from plants grown on 1/20th MS medium modified accordingly to Cheong et al. [22] as previously described. After cooling, all samples were digested with 0.7 mL concentrated nitric acid (OmniTrace, VWR) and diluted to 6.0 mL with 18 MΩ water. Acid used for digestion was spiked with gallium (Ga) to act as an internal standard to control for errors in dilution, variations in sample introduction, and plasma stability in the ICP-MS instrument. Sample sets also contained analytical blanks, standard reference material (NIST SRM 1547) digested in the same manner as the plant samples, and quantitative calibration standards. Calibration standards and standard reference material samples were included at the beginning and end of the sample sets to control for drift during the analysis. Samples were introduced into an inductively coupled plasma mass spectrometer (ICP-MS) (Elan DRCE, PerkinElmer) and analyzed for Li, B, Na, Mg, P, K, Ca, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Mo and Cd. All samples were normalized to calculated weights, as determined with an iterative algorithm using the best-measured elements, the weights of the seven weighed samples, and the solution concentrations, detailed at www.ionomicshub.org.

Determination of SA content

Total SA content was quantified in leaves of five weeks old plants using a Waters Alliance HPLC system equipped with Millennium software, 2695 Separation Module, 2475 Fluorescence Detector, and 2996 Photodiode array detector. A Nova-Pack C-18 column was used with a flow rate and methanol gradient as described previously [26]. SA (Sigma; catalogue no. S-6271) was used to develop the standard curve for quantification.

Grafting

cpr5-2 and Col-0 seeds were germinated in the dark on 0.5× MS plates containing 1 mL L⁻¹ of MS Vitamins (Caisson Laboratories, Inc.), 3 mg L⁻¹ Benomyl (methyl 1-(butylcarbamoyl)-2-benzimidazolecarbamate; Sigma), 0.04 mg L⁻¹ BA (6-benzylaminopurine; Sigma), 0.02 mg L⁻¹ IAA (indole acetic acid; Sigma), and 12 g L⁻¹ agar. Plates were held vertically and after seven days

seedlings were grafted as previously described [27] and then grown for an additional seven days on plates before transfer into soil. Plants were grown for a further four weeks in soil before being analyzed for their elemental content. Plants that from a visual inspection showed adventitious roots coming from the shoot above the graft were excluded from the experiment.

Statistical analysis

ANOVA was conducted using the software CoStat 6.2 (CoHort Software, CA, USA). Separation of means was performed using LSD test at $P=0.05$ significance level.

Results

Mutant identification

Fast neutron mutagenized *A. thaliana* Col-0 plants were previously screened for altered leaf elemental composition [11]. In this screen *12645* was identified as a low K^+ mutant with a reduction in leaf K^+ of approximately 20% compared to wild-type Col-0 (raw data are available at www.ionomicshub.org, experimental tray 229) (Fig. 1). In addition, *12645* was also smaller in size than wild-type Col-0 and developed symptoms of hypersenescence in cotyledons and mature leaves. Moreover, cotyledons of mutant plants were hyponastic when seeds were germinated in soil, as well as in plates.

Genetic analysis

To map and define the Mendelian character of the mutation, *12645* was out-crossed to *Ler-0*, and F1 and F2 populations scored for mutant like plants. All F1 plants from this cross looked wild-type, while in the F2 small plants with early senescent leaves segregated with a ratio of 3:1 (wild-type to mutant phenotype), indicating that *12645* is inherited as a single recessive nuclear mutation. Small hypersenescent plants from the F2 population also showed reduced K^+ content ($38.3 \mu\text{g g}^{-1}$ dry weight) compared to those that looked wild-type ($42.8 \mu\text{g g}^{-1}$ dry weight), indicating that the traits of early senescence and small size co-segregate with reduced leaf K^+ ($P<0.001$).

The chromosomal position of the *12645* mutation was determined using a positional cloning approach with SSLP markers in 171 F2 plants from the outcross with *Ler-0*. The recombinant population scored for markers in the region between nucleotides 25,500,000 and 26,100,000 on chromosome 5 revealed that only one plant had *Ler-0* alleles on the BAC clone MUB3 and two plants on F15O5, which placed the mutation on the BAC clone MXK3 (Fig. 2A). DNA sequencing of this region revealed that *12645* contains a 972 base pair insertion at nucleotide 1474 in the fourth exon of the At5g64930 gene (Fig. 2B). This insertion originated from nucleotides 77,802–78,764 on chromosome 1, a region that does not contain any annotated loci. *A. thaliana* mutants carrying recessive loss-of-function alleles of At5g64930 have been previously and independently identified in screens for constitutive expression of pathogen resistance, and the early appearance of hypersenescence symptoms, and named *cpr5-1* [13], *cpr5-2* [12] and *hys1* [17]. Based on the previous nomenclature we renamed *12645* as *cpr5-3*. As expected *CPR5* expression is lost in *cpr5-3* (Fig. S1A). All *cpr5* mutants (*cpr5-1*, *cpr5-2* and *cpr5-3*) were grown together, along with the wild-type Col-0 and the concentration of Li, B, Na, Mg, P, S, K, Ca, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Mo, Cd in leaves of each genotype determined by ICP-MS ($n=10-22$ replicate plants per genotype). Of all the elements measured only K^+ was found to be significantly different between the wild-type Col-0 and all the *cpr5* alleles ($P<0.005$ after Bonferroni correction for multiple testing). Wild-type Col-0 had a leaf K^+ concentration of $40,067 \pm 6222 \mu\text{g g}^{-1}$ dry weight, compared to $27,490 \pm 3575 \mu\text{g g}^{-1}$ dry weight for *cpr5-1*, $29,751 \pm 2616 \mu\text{g g}^{-1}$ dry weight for *cpr5-2*, and $27,324 \pm 2502 \mu\text{g g}^{-1}$ dry for *cpr5-3* (raw data are available at www.ionomicshub.org, experimental tray 1020).

Low leaf K^+ in *cpr5* is not dependent on SA or JA, and it is a unique trait of *cpr5* among lesion mimic mutants

CPR5 acts as a negative regulator of local resistance to pathogens by partially repressing the accumulation of SA, so that *cpr5* mutants have elevated SA content [13,14], which we also confirmed to be the case for *cpr5-3* (Fig. S1C). This raised the

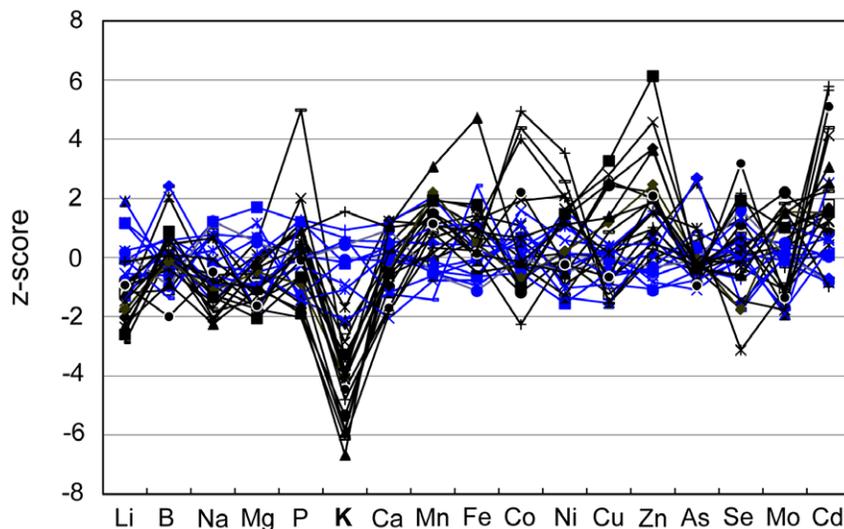


Figure 1. Leaf ionic profile of *cpr5-3* (*12645*). The concentration of each element was measured (in leaves of five weeks old plants of both *12645* (M3 generation) (black lines; $N=21$) and wild type Col-0 (blue lines; $N=12$) plants. For each element given on the abscissa the correspondent z-score value is given on the ordinate. The z-score represents the number of standard deviations of the wild-type Col-0 each plant differs from the mean of the wild-type Col-0 (raw data are available at www.ionomicshub.org, experimental tray 743). doi:10.1371/journal.pone.0026360.g001

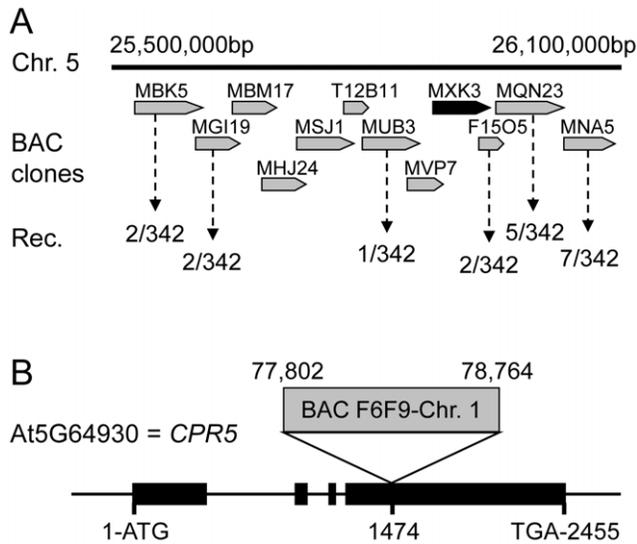


Figure 2. Mapping and gene structure of *cpr5-3* (12645). **A.** BAC clones covering the *cpr5-3* (12645) locus. Frequency of recombination is expressed as number of recombinant chromosomes over the total number of examined chromosomes. **B.** 972 bp insertion from BAC clone F6F9 from chromosome 1 inserted at nucleotide 1474 in the fourth exon of At5g64930 gene. Exons are shown by boxes and introns by lines.

doi:10.1371/journal.pone.0026360.g002

question, whether low leaf K^+ in *cpr5* results from the constitutively high SA in this mutant. To address this we measured K^+ content in three different *cpr5* alleles, in the single *eds5-1* mutant, and in the double mutant *cpr5eds5* prepared from *cpr5-1* and *eds5-1* [14]. The mutation in *eds5-1* suppresses the biosynthesis of SA and in the double mutant *cpr5eds5* the elevated SA content of *cpr5-1* is returned to the level of the wild-type [14,28,29]. As shown in Fig. 3A, K^+ content in *eds5-1* leaves is slightly reduced but not significantly different from Col-0 whereas in *cpr5eds5*, which has low SA but carries the *cpr5-1* mutation, the content of K^+ is similar to that of single *cpr5* alleles (raw data are available at www.ionomicshub.org, experimental tray 1020).

We also followed a similar approach to determine if the low leaf K^+ of *cpr5* is dependent on the elevated JA in this mutant [16]. The *jasmonate resistant 1* (*jar1*) mutant is unable to conjugate JA with isoleucine to form the active jasmonoyl-L-isoleucine form of JA that is required to elicit the JA response [30], and therefore this mutant is insensitive to JA [31]. Fig. 3B shows that the double mutant *cpr5jar1*, prepared from *cpr5-1* and *jar1-1* [14], and single allelic mutants *cpr5-1*, *cpr5-2* and *cpr5-3* all share a similar reduction of leaf K^+ which is not observed in the single *jar1-1* mutant (raw data are available at www.ionomicshub.org; experimental tray 1840). From this we conclude that the low leaf K^+ of *cpr5* is not dependent on JA signalling. Taken together these experiments support the conclusion that the reduced leaf K^+ of *cpr5* is independent of both SA and JA.

cpr5 is easily distinguishable from wild-type plants from the presence of necrotic and chlorotic spots on the leaves, a trait which characterizes plants infected by bacteria and/or fungi, as well as mutants with a constitutively active response to pathogens. Hypothesizing that low leaf K^+ was associated with the response to pathogens, we measured K^+ content in leaves of lesion mimic mutants (LMM) selected from the two major classes of initiation (*dnd1*, *dnd2*, *agd2*, *acd6*, *lsd6*) and propagation (*acd1*, *acd2*, *vad1*) of lesion mutants [32]. None of the lesion mimic mutants tested

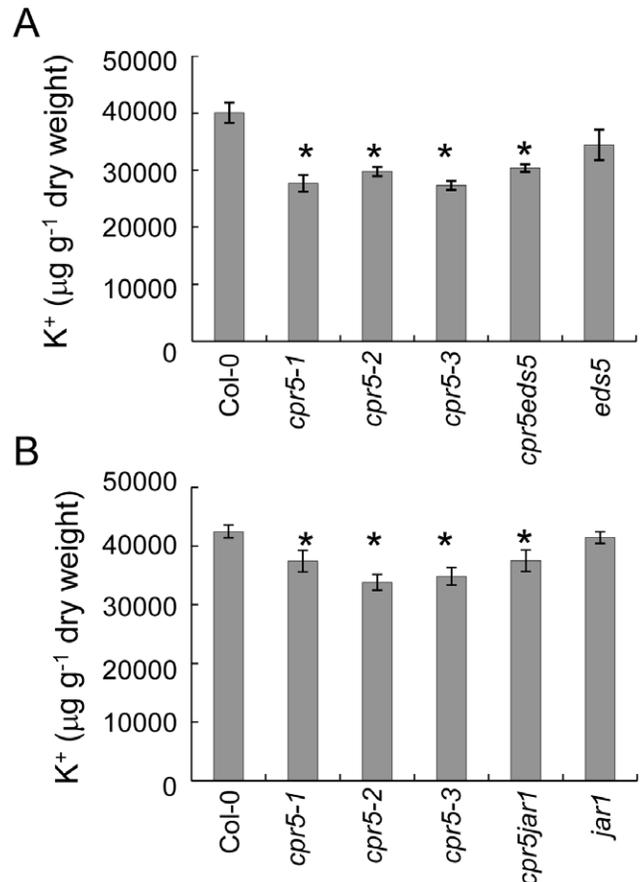


Figure 3. K^+ content in salicylic acid and jasmonic acid defective mutants. K^+ ($\mu\text{g g}^{-1}$ of dry weight) was measured in leaves of five weeks old plants of wild-type Col-0, *cpr5-1*, *cpr5-2*, *cpr5-3* and (A) *eds5* and *cpr5eds5*, and (B) *jar1* and *cpr5jar1*. Wild-type Col-0 was used as a control. Values represent means of independent biological replicates (between 10 and 22 plants per genotype). Error bars represent standard errors. An asterisk (*) indicates values significantly different from the mean of the wild-type Col-0 at 99% confidence interval (raw data are available at www.ionomicshub.org; experimental trays 838, 839, 1020 and 1840).

doi:10.1371/journal.pone.0026360.g003

showed reduced leaf K^+ (Fig. 4A, B). From this we conclude that the low leaf K^+ observed in *cpr5* is not a result of the presence of lesions.

Low leaf K^+ in *cpr5* is primarily driven by the shoot but roots also play a role

Low K^+ in *cpr5* leaves could be caused by reduced uptake from roots, impaired root-to-shoot translocation, or enhanced shoot-to-root circulation through the phloem. *CPR5* is equally expressed in both roots and leaves (Fig. S1B) making it possible that *CPR5* could contribute to K^+ homeostasis in either organ. Therefore, we performed reciprocal grafting of *cpr5* and wild-type Col-0 to determine in which tissue *cpr5* exerts its influence. Shoots of *cpr5-2* were grafted onto wild-type Col-0 roots and vice versa, grafted plants allowed to grow for four weeks in soil and the K^+ content measured in leaves. When *cpr5-2* shoots were grafted on wild-type Col-0 roots we observed the K^+ content in leaves to be significantly reduced by 43% compared to self-grafted Col-0 (Fig. 5A). Moreover, plants with *cpr5-2* shoot and wild-type Col-0 root also retained the hypersensitive phenotype of chlorotic and necrotic

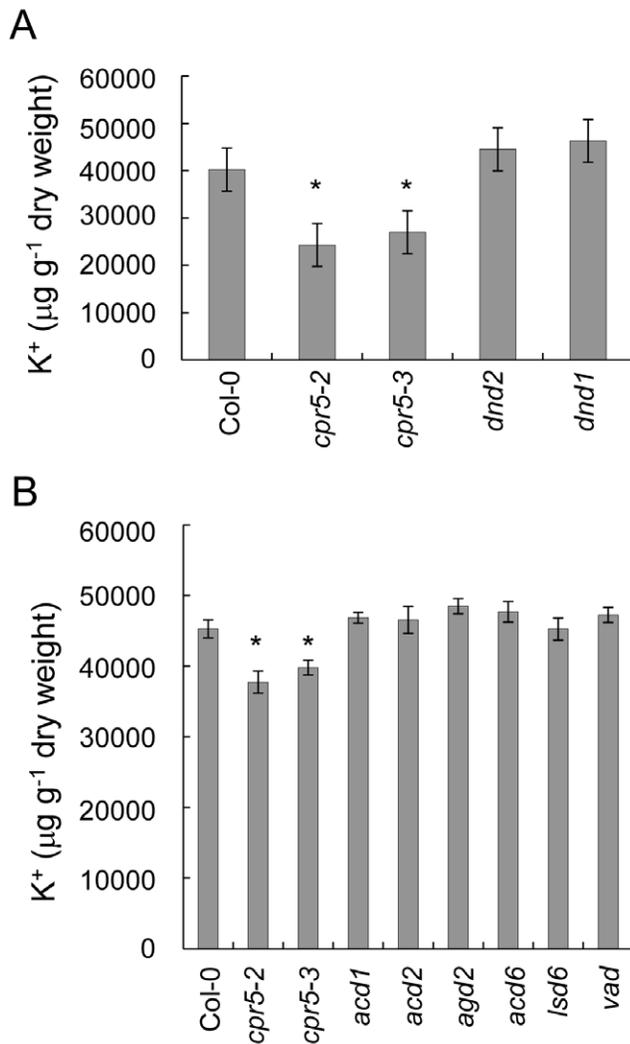


Figure 4. K⁺ content in lesion mimic mutants. K⁺ (μg g⁻¹ of dry weight) content was measured in leaves of five weeks old plants of *cpr5-1*, *cpr5-2*, *cpr5-3* and (A) initiation of lesion mimic mutants *dnd1* and *dnd2*, and (B) initiation of lesion mimic *agd2*, *acd6*, *lsd6*, and propagation of lesion mimic mutants *acd1*, *acd2*, *vad1*. Wild-type Col-0 was used as a control. Values represent means of independent biological replicates (between 10 and 22 plants per genotype). Error bars represent standard errors. An asterisk (*) indicates values significantly different from wild-type Col-0 at 99% confidence interval (raw data are available at www.ionomicshub.org; experimental trays 1770 and 1771).

doi:10.1371/journal.pone.0026360.g004

spots observed on leaves of non-grafted *cpr5-2* plants. In contrast, plants with wild-type Col-0 shoots grafted on *cpr5-2* roots showed only an 11% reduction in leaf K⁺ and were indistinguishable from Col-0 in respect to symptoms of hypersenescence. Interestingly, self-grafted *cpr5-2* plants showed the lowest leaf K⁺ of all the grafting types tested, with a reduction in leaf K of 58%. These results suggest that loss-of-function of *CPR5* in shoots plays a primary role in the reduced leaf K⁺ phenotype of *cpr5*, but loss-of-function of *CPR5* in roots also exerts a lesser yet significant influence.

To further understand the role of roots versus shoots in the low leaf K⁺ phenotype of *cpr5* we measured the K⁺ content of shoot and root tissue of plants grown on solidified MS medium in plates. This experiment revealed no significant difference in the root

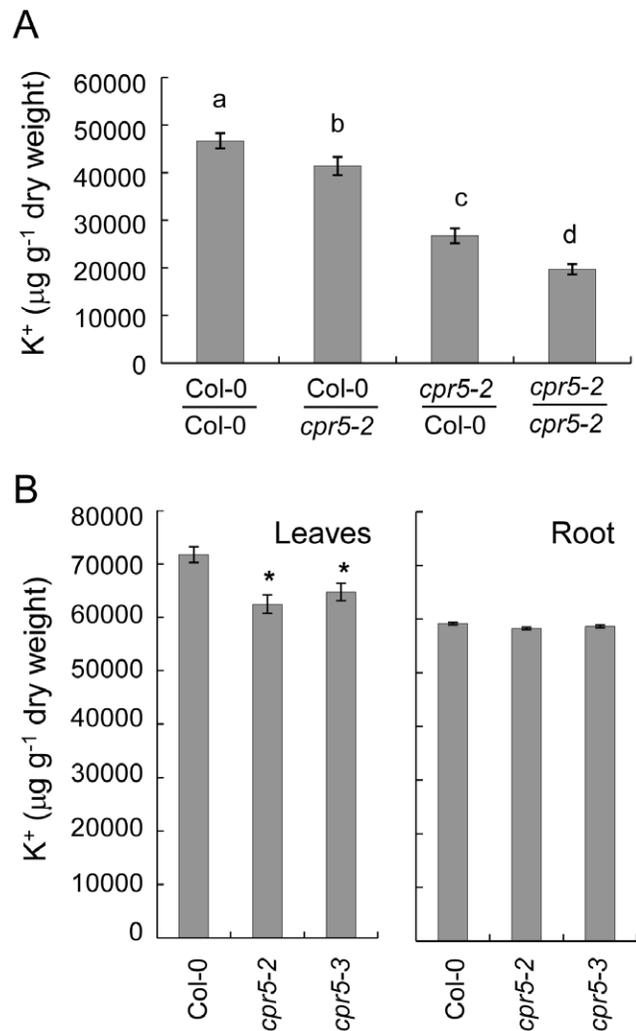


Figure 5. K⁺ content in *cpr5* root and leaves and in *cpr5* grafted plants. A. K⁺ (μg g⁻¹ of dry weight) in leaves of grafted plants transferred to soil after grafting and grown for four weeks. Values represent means of independent measurements performed on 19 to 32 different plants per graft combination. Error bars show standard errors. Bars with different letters are significantly different at $P=0.05$ level (LSD test). Col-0/Col-0, self-grafted wild-type Col-0; Col-0/*cpr5-2*, wild-type Col-0 shoot grafted on *cpr5-2* root; *cpr5-2*/Col-0, *cpr5-2* shoot grafted on wild-type Col-0 root; *cpr5-2*/*cpr5-2*, self-grafted *cpr5-2*. B. K⁺ (μg g⁻¹ of dry weight) in leaves and roots of wild-type Col-0, *cpr5-2* and *cpr5-3* plants grown for two weeks on solidified 1/20th MS medium supplemented with 20 mM of K⁺. Data represents the mean of 10 measurements of a pool of 10 to 15 shoots or roots per genotype grown on 15 different plates. Error bars represent standard errors. An asterisk (*) indicates values significantly different from the mean of wild-type Col-0 at $P=0.005$.

doi:10.1371/journal.pone.0026360.g005

concentration of K⁺ between *cpr5* and Col-0, while *cpr5* shoots retained the low K⁺ phenotype observed in plants grown in soil (Fig. 5B).

Expression of Cyclic Nucleotide Gated Channels is elevated in shoots of *cpr5*

Based on the importance of the shoot in driving the reduced leaf K⁺ in *cpr5* we investigated expression of *Cyclic Nucleotide Gated Channels* (CNGCs) that may be directly or indirectly involved in K⁺ efflux during the response to pathogens in *A. thaliana*. An initial

survey of transcriptional data publicly available in experiment 175 on Genevestigator [33] revealed that the expression of numerous *CNGCs* was up-regulated in *cpr5* leaves. We confirmed the differences initially observed in the database using qRT-PCR, and performed similar measurements in root tissue. Our analysis revealed that steady state levels of *CNGC10*, *CNGC11*, *CNGC12*, *CNGC19* and *CNGC20* mRNA are all elevated in leaves of both *cpr5-2* and *cpr5-3* compared to wild-type Col-0 (Fig. 6). Analogous measurements performed in roots did not reveal any substantial differences from wild-type Col-0 for the *cpr5* mutants (Fig. 6).

Expression of *HAK5* encoding a high affinity K^+ transporter is reduced in roots of *cpr5*

The K^+ transporter *HAK5* is known to contribute to K^+ uptake in *A. thaliana* primarily at low K^+ supply (0–0.25 mM) [23,34–37], whereas the K^+ channel *AKT1* [38,39] contributes to K^+ uptake at both low and intermediate K^+ supply (0.01–5 mM) [35–37,40]. Above 5 mM external K^+ the transport processes involved in K^+ uptake are currently undefined [36]. Given the importance of both *HAK5* and *AKT1* in K^+ uptake in *A. thaliana* we used qRT-PCR to quantify the steady state levels of *HAK5* and *AKT1* mRNA in *cpr5* roots to test if altered expression of these genes may be involved in the low leaf K^+ phenotype we observe in *cpr5*. Steady state levels of *AKT1* mRNA in *cpr5* were found to be the same as wild-type Col-0 after growth on medium supplemented with either high (20 mM) or low (100 μ M) K^+ (Fig. 7C). A slight increase in *AKT1* mRNA was observed in wild-type Col-0 plants grown on medium supplemented with 100 μ M K^+ compared to 20 mM K^+ . Enhanced expression of *AKT1* was not previously observed [41], though this is possibly due to the fact that the previous authors used RT-PCR to determine expression of *AKT1*. Interestingly, steady state levels of *HAK5* mRNA were observed to be reduced in *cpr5* compared to wild-type Col-0 grown in medium with either high and low K^+ supply (Fig. 7B). Further, the increase in *HAK5* mRNA observed in wild-type Col-0 grown on medium containing 100 μ M K^+ was essentially abolished in *cpr5* (Fig. 7B). Interestingly, root growth of *cpr5* on medium supplemented with 100 μ M K^+ was found to be reduced compared to wild-type (Fig. 7A). However, in medium supplemented with 20 mM K^+ growth of *cpr5* and wild-type was similar (Fig. 7A).

To confirm and extend these results we performed a more detailed dose response experiment with solidified medium supplemented with 10, 50, and 100 μ M KCl lacking NH_4^+ given that NH_4^+ is known to suppress activity [38,39] and expression [23] of *HAK5*. Pure agarose was used to solidify the growth medium to avoid any extra K^+ supply [42]. In wild-type Col-0 root steady state levels of *HAK5* mRNA were observed to increase as external K^+ was reduced (Fig. 8A), as expected [34]. In roots of *cpr5-2* and *cpr5-3* steady state levels of *HAK5* mRNA were also increased as the external K^+ concentration was reduced (Fig. 8A). However, steady state levels of *HAK5* mRNA in *cpr5-2* and *cpr5-3* were consistently lower than wild-type Col-0 at 10, 50 and 100 μ M K^+ in the growth medium (Fig. 8A). In agreement with our previous observations no consistent differences in expression of *AKT1* between wild-type Col-0 and *cpr5* were observed (Fig. 8B).

Discussion

The data presented here establishes that the wild-type allele of *CPR5* is required to maintain normal K^+ homeostasis in *A. thaliana* Col-0. We observed that loss-of-function alleles of *CPR5* show a consistent and specific reduction in leaf K^+ of 10–30% in plants grown in both soil or on solidified MS medium with high K^+ supply. Further, this defect leads to reduced growth at low K^+ supply. Genetic analysis confirmed that this reduction in leaf K^+ is

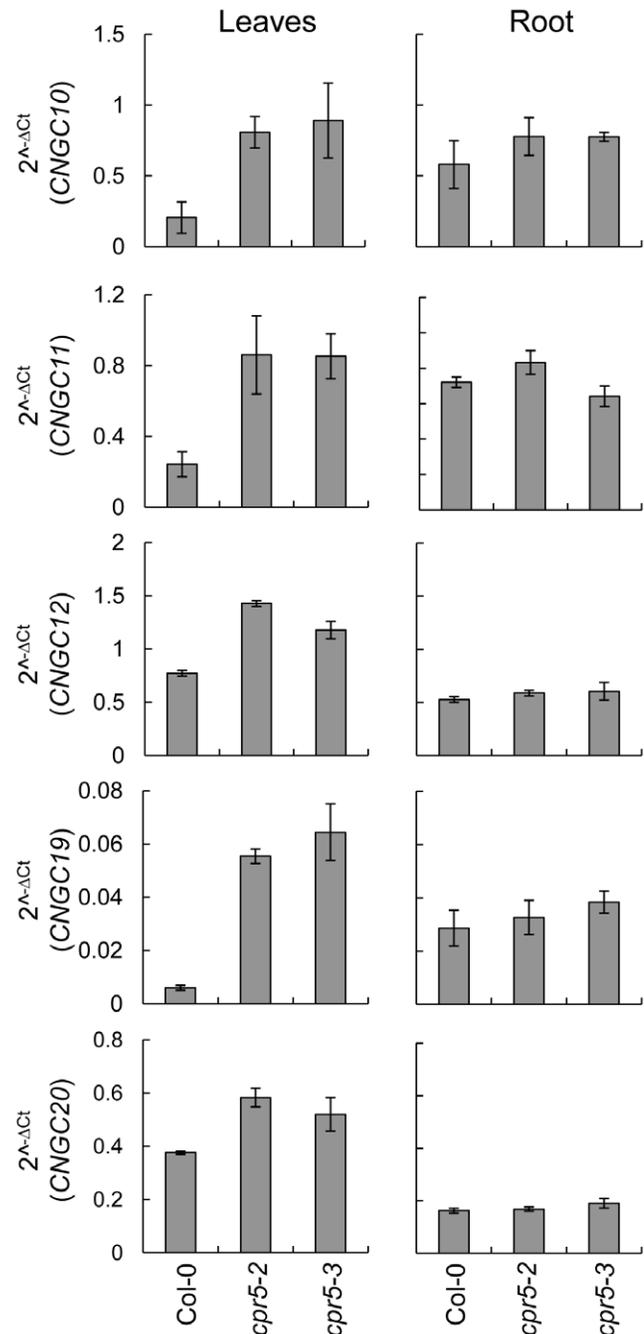


Figure 6. Expression of *CNGCs* in *cpr5*. Steady state levels of *CNGC10*, *CNGC11*, *CNGC12*, *CNGC19*, *CNGC20* mRNA in leaves and roots of *cpr5-2*, *cpr5-3* and wild-type Col-0 evaluated by qRT-PCR. Seedlings were germinated and grown on 1/20th MS medium supplemented with 20 mM KCl. After two-week of growth shoot and root tissue was harvested and RNA extract for qRT-PCR. *PP2A* (*At1g13320*) was used as an endogenous reference gene for normalization across samples, and data presented as $2^{-\Delta\Delta CT}$. Error bars represent standard deviations calculated following [29].

doi:10.1371/journal.pone.0026360.g006

not dependent on the elevated SA or JA known to occur in *cpr5*. Further, genetic analysis confirmed that reduced leaf K^+ is not a feature of lesion mimic mutants in general. Through reciprocal grafting we establish that the reduced leaf K^+ observed in *cpr5* is primarily driven by the shoot (74%), with the root playing a

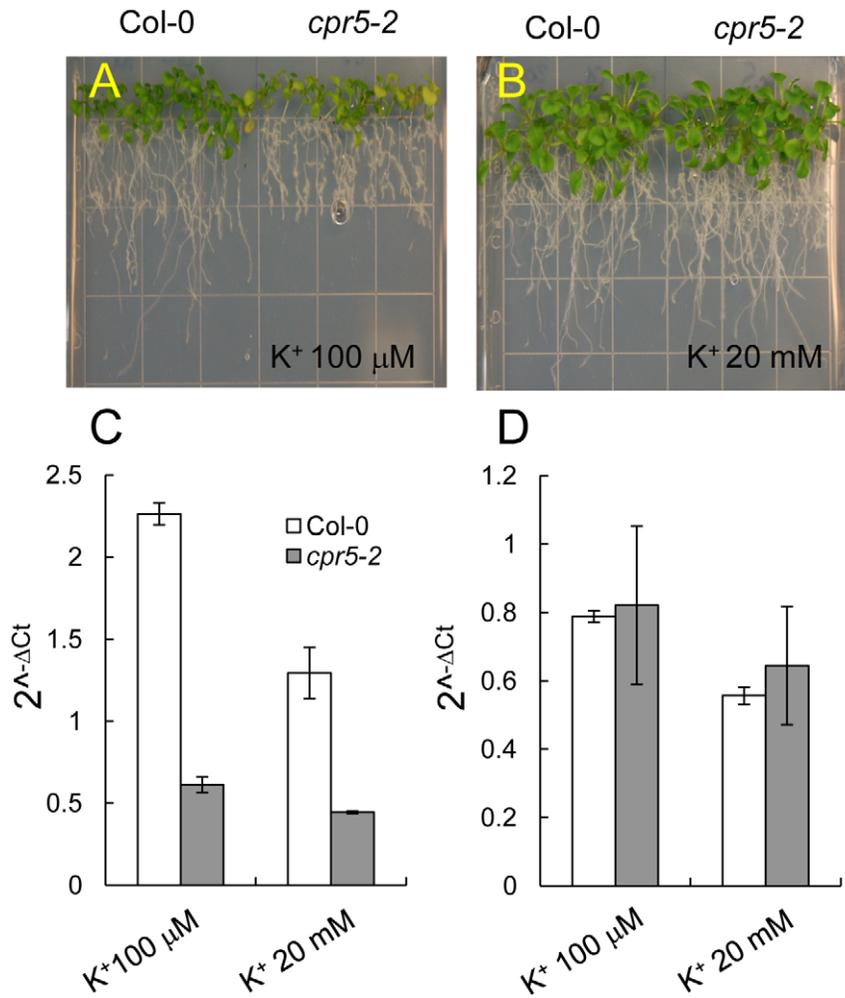


Figure 7. Expression of *HAK5* and *AKT1* in roots of *cpr5*. Seeds were germinated on solidified 0.5 × MS salts, and after five days seedlings transferred to solidified 1/20th MS medium supplemented with either 100 μM (A) or 20 mM KCl (B). Pictures were taken after two weeks of growth. Steady state levels of *HAK5* (C) and *AKT1* (D) mRNA were quantified using qRT-PCR in roots of wild-type Col-0 (white bars) and *cpr5-2* (gray bar). RNA was extracted from roots of three independent samples generated from between 25 and 30 plants per plates. *UBQ10* (At4g05320) was used as an endogenous reference gene for normalization across the samples, and data presented as 2^{-ΔΔCt}. Error bars represent standard deviations calculated following [29]. doi:10.1371/journal.pone.0026360.g007

significant but smaller role (19%). Interestingly, the presence of the *cpr5* allele in both roots and shoots is required to produce the full low leaf K⁺ phenotype, suggesting that feedback between both organs is needed. In investigating the factors that in *cpr5* cause the reduction of leaf K⁺, we surveyed the experiment AT-175 performed on *cpr5* shoots (www.geneinvestigator.ethz.ch) and this revealed that genes belonging to the *CNGC* family are highly expressed in *cpr5*. Using qRT-PCR we confirmed that *CNGC10*, *CNGC11*, *CNGC12*, *CNGC19*, *CNGC20* are indeed highly expressed in leaves, but not roots, of *cpr5*. The expression of *CNGC1* and *CNGC13* which showed a milder increase in the AT-175 array did not show any difference when expression was analyzed by qRT-PCR. *CNGC2* and *CNGC4*, null mutants of which (*dnd1* and *dnd2*) have enhanced disease resistance, also did not show any misregulation when analyzed by qRT-PCR. In the interaction between plant and pathogens the recognition of factors of avirulence by the host plant induces fluxes of Ca²⁺ that initiate the immune response and leads to enhanced K⁺ efflux [10]. *CNGCs* are believed to be the channels that deliver the Ca²⁺ signal required for pathogen recognition [43], and *dnd1*, *dnd2* and the gain of function chimeric *CNGC11/12* mutant *cpr22* strongly support this conclusion [44–50]. Moreover, heterologous

expression of specific *CNGCs* provides direct evidence of Ca²⁺ transport activity [51,52]. It is therefore possible that in *cpr5* the constitutively high level of expression of *CNGCs* we observe leads to a persistent activation of this Ca²⁺ initiated signalling cascade that in turn leads to constitutively enhanced K⁺ efflux [10]. Such an enhanced K⁺ efflux could increase shoot-to-root K⁺ export, and may explain the leaf-driven portion of the reduced leaf K⁺ observed in *cpr5*. The transporters/channels involved in this K⁺ efflux are currently unknown. However, there is evidence that *CNGC10* transports K⁺ [53], and steady state levels of *CNGC10* mRNA are elevated in *cpr5*, making it possible that *CNGC10* could play a role in the proposed enhanced K⁺ efflux and shoot-to-root export in *cpr5*.

HAK5 encodes the primary high affinity root K⁺ transporter in *A. thaliana* [23,34–37], and its expression in wild-type Col-0 roots is induced at low K⁺ supply [34]. Significantly, *cpr5* roots show a consistent reduction in the steady state levels of *HAK5* mRNA under both high and low K⁺ supply. Though *HAK5* primarily plays a role in K⁺ uptake at low K⁺ supply, it is possible that the reduced steady state levels of *HAK5* mRNA in *cpr5* roots we observe even at high K⁺ supply is responsible for the small (18%) but significant contribution of roots to the reduced leaf K⁺

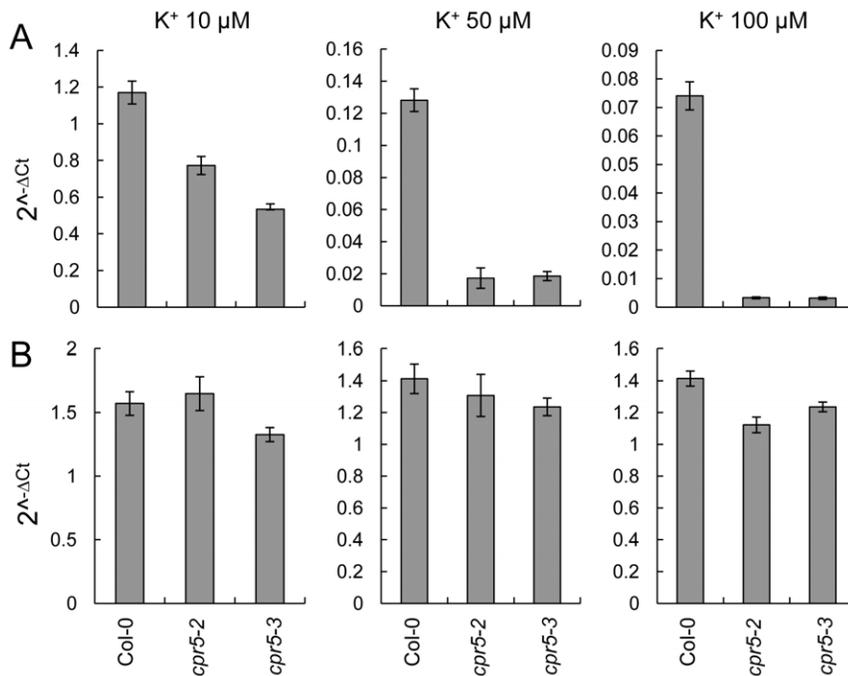


Figure 8. Expression of *HAK5* and *AKT1* in roots of *cpr5* grown in the absence of NH_4^+ . Wild-type Col-0, *cpr5-2* and *cpr5-3* seedlings were grown on solidified defined medium without NH_4^+ and supplemented with different concentrations of KCl (10, 50 and 100 μM). RNA was extracted from roots of two week old plants and steady state levels of *HAK5* (A) and *AKT1* (B) quantified using qRT-PCR. Data represents the mean of four biological replicates each composed of a pool of 5 to 10 plants. *PP2A* (*At1g13320*) was used as an endogenous reference gene for normalization across samples, and data presented as $2^{-\Delta\text{Ct}}$. Error bars represent standard deviations calculated following [29]. doi:10.1371/journal.pone.0026360.g008

phenotype of *cpr5*. It is also interesting to speculate that expression of *HAK5* in *cpr5* roots is suppressed by an increased flux of K^+ arriving in the roots from the shoots driven by elevated expression of *CNGCs* in shoots.

Based on sequence analysis *CPR5* encode a membrane protein with five transmembrane domains and a nuclear localization signal (NLS). *CPR5* appears to localize to the nucleus [54] where it has been proposed to be targeted to the inner nuclear membrane [19,54]. It has been further suggested that proteolytic cleavage can release the nucleosolic domain of *CPR5* from the membrane to allow its participation in transcriptional processes [19,54]. Recent studies have identified the transcription factors *TPR1* and *EDS1* that direct repression of expression of *CNGC2* and *CNGC4* [55,56], and we speculate that *CPR5* may play a similar role in the nucleus to negatively regulate the expression of the various *CNGCs* affected on the *cpr5* mutant.

In summary, we have identified a new low leaf K^+ phenotype for *cpr5* that is primarily driven by the shoot, and is independent of the elevated levels of SA and JA found in this mutant. We suggest that the reduced leaf K^+ of *cpr5* is caused by the elevated expression of various *CNGCs* in shoots and the reduced expression of *HAK5* in roots, driving both an enhanced K^+ export from shoots and a reduced K^+ uptake in roots. Our observation of altered K^+ homeostasis in *cpr5* may also be an important piece of evidence linking the function of *CPR5* as a negative regulator of local defence responses to pathogens with the role K^+ efflux plays in these responses; likely through the direct or indirect regulation of *CNGCs* by *CPR5*.

Supporting Information

Figure S1 *CPR5* expression and salicylic acid (SA) levels in *A. thaliana* leaves and roots of wild-type Col-0 and

***cpr5*.** A. Steady state levels of *CPR5* mRNA in wild-type Col-0 and *cpr5-3* quantified using qRT-PCR. RNA was extracted from leaves of five-week old plants grown in soil. Data represents the mean of measurements of four independent biological replicates. Each biological replicate consisted of 2-3 leaves from individual plants. Errors bars represent standard deviation. B. Steady state levels of *CPR5* mRNA in root and shoots of wild-type Col-0 quantified using qRT-PCR. RNA was extracted from shoots and roots of two week old wild-type Col-0 plants grown on 0.5 \times MS media solidified with 1% agar (w/v). Values represent mean of measurements from at least three independent replicates. Errors bars represent standard deviation. Steady state mRNA levels (A & B) are presented as $2^{-\Delta\text{Ct}}$. *UBQ10* (*At4g05320*) was used as an endogenous reference gene for normalization across samples. C. SA content (mg g^{-1} of fresh weight) in leaves of wild-type Col-0, *cpr5-2* and *cpr5-3*. Data represents the mean of three independent leaf samples harvested from individual plants grown in soil for five weeks. Error bars represent the standard error. (TIF)

Acknowledgments

The authors wish to acknowledge Brett Lahner for ICP-MS analysis and Hyeong Cheol Park for salicylic acid determination. We wish to acknowledge Xinniang Dong's laboratory for providing seeds of *cpr5-1*, *eds5-1* and *cpr5eds5* and Fred Ausubel for giving us permission to use them. We also acknowledge Shannon Clarke and Luis Mur for providing seeds of *cpr5jar1*.

Author Contributions

Conceived and designed the experiments: MB AR DES. Performed the experiments: MB AR. Analyzed the data: MB AR DES. Wrote the paper: MB DES.

References

- Marschner H (2002) Mineral nutrition of higher plants: New York: Academic Press. pp 299–312.
- Ashley MK, Grant M, Grabov A (2006) Plant responses to potassium deficiencies: a role for potassium transport proteins. *J Exp Bot* 57: 425–436.
- Grignon C, Sentenac H (1991) pH and ionic conditions in the apoplast. *Annu Rev Plant Biol* 42: 103–128.
- White P (1997) The regulation of K⁺ influx into roots of rye (*Secale cereale* L.) seedlings by negative feedback via the K⁺ flux from shoot to root in the phloem. *J Exp Bot* 48: 2063–2073.
- Shabala S (2007) Transport from root to shoot. In: Yeo A, Flowers T, eds. Plant solute transport Blackwell Publishers. pp 214–234.
- Armengaud P, Breitling R, Amtmann A (2004) The potassium-dependent transcriptome of Arabidopsis reveals a prominent role of jasmonic acid in nutrient signaling. *Plant Physiol* 136: 2556–2576.
- Armengaud P, Sulpice R, Miller AJ, Stütt M, Amtmann A, et al. (2009) Multilevel analysis of primary metabolism provides new insights into the role of potassium nutrition for glycolysis and nitrogen assimilation in Arabidopsis roots. *Plant Physiol* 150: 772–785.
- Amtmann A, Troufflard S, Armengaud P (2008) The effect of potassium nutrition on pest and disease resistance in plants. *Physiol Plantarum* 133: 682–691.
- Jabs T, Tschöpe M, Colling C, Hahlbrock K, Scheel D (1997) Elicitor-stimulated ion fluxes and O₂⁻ from the oxidative burst are essential components in triggering defense gene activation and phytoalexin synthesis in parsley. *Proc Natl Acad Sci U S A* 94: 4800–4805.
- Jeworutzki E, Roelofsma MRG, Anschütz U, Krol E, Elzenga JTM, et al. (2010) Early signaling through the Arabidopsis pattern recognition receptors FLS2 and EFR involves Ca²⁺-associated opening of plasma membrane anion channels. *Plant J* 62: 367–378.
- Lahner B, Gong J, Mahmoudian M, Smith EL, Abid KB, et al. (2003) Genomic scale profiling of nutrient and trace elements in *Arabidopsis thaliana*. *Nature Biotech* 21: 1215–1221.
- Boch J, Verbsky M, Robertson T, Larkin J, Kunkel B (1998) Analysis of resistance gene-mediated defense responses in *Arabidopsis thaliana* plants carrying a mutation in *CPR5*. *Mol Plant Microbe In* 11: 1196–1206.
- Bowling S, Clarke J, Liu Y, Klessig D, Dong X (1997) The *cpr5* mutant of Arabidopsis expresses both NPR1-dependent and NPR1-independent resistance. *Plant Cell* 9: 1573–1584.
- Clarke JD, Volko SM, Ledford H, Ausubel FM, Dong X (2000) Roles of salicylic acid, jasmonic acid, and ethylene in *cpr*-induced resistance in Arabidopsis. *Plant Cell* 12: 2175–2190.
- Jing H, Sturre M, Hille J, Dijkwel P (2002) Arabidopsis onset of leaf death mutants identify a regulatory pathway controlling leaf senescence. *Plant J* 32: 51–63.
- Jing HC, Anderson L, Sturre MJ, Hille J, Dijkwel PP (2007) Arabidopsis *CPR5* is a senescence-regulatory gene with pleiotropic functions as predicted by the evolutionary theory of senescence. *J Exp Bot* 58: 3885–3894.
- Yoshida S, Ito M, Nishida I, Watanabe A (2002) Identification of a novel gene *HYS1/CPR5* that has a repressive role in the induction of leaf senescence and pathogen-defense responses in *Arabidopsis thaliana*. *Plant J* 29: 427–437.
- Kirik V, Bouyer D, Schobinger U, Bechtold N, Herzog M, et al. (2001) *CPR5* is involved in cell proliferation and cell death control and encodes a novel transmembrane protein. *Curr Biology* 11: 1891–1895.
- Brininstool G, Kasili R, Simmons LA, Kirik V, Hulskamp M, et al. (2008) *Constitutive Expressor Of Pathogenesis-related Genes5* affects cell wall biogenesis and trichome development. *BMC Plant Biol* 8: 58.
- Jing HC, Hebel R, Oeljeklaus S, Sitek B, Stuhler K, et al. (2008) Early leaf senescence is associated with an altered cellular redox balance in Arabidopsis *cpr5/old1* mutants. *Plant Biol* 10(Suppl 1): 85–98.
- Gao G, Zhang S, Wang C, Yang X, Wang Y, et al. (2011) Arabidopsis *CPR5* Independently Regulates Seed Germination and Postgermination Arrest of Development through LOX Pathway and ABA Signaling. *PLoS One* 6: e19406.
- Cheong YH, Pandey GK, Grant JJ, Batistic O, Li L, et al. (2007) Two calcineurin B-like calcium sensors, interacting with protein kinase CIPK23, regulate leaf transpiration and root potassium uptake in Arabidopsis. *Plant J* 52: 223–239.
- Qi Z, Hampton CR, Shin R, Barkla BJ, White PJ, et al. (2008) The high affinity K⁺ transporter AtHAK5 plays a physiological role in planta at very low K⁺ concentrations and provides a caesium uptake pathway in Arabidopsis. *J Exp Bot* 59: 595–607.
- Czechowski T, Stütt M, Altmann T, Udvardi MK, Scheible WR (2005) Genome-wide identification and testing of superior reference genes for transcript normalization in Arabidopsis. *Plant Physiol* 139: 5–17.
- Livak KJ, Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the 2^{-ΔΔCT} method. *Methods* 25: 402–408.
- Freeman JL, Garcia D, Kim D, Hopf A, Salt DE (2005) Constitutively elevated salicylic acid signals glutathione-mediated nickel tolerance in *Thlaspi* nickel hyperaccumulators. *Plant Physiol* 137: 1082–1091.
- Rus A, Baxter I, Muthukumar B, Gustin J, Lahner B, et al. (2006) Natural variants of *AtHKT1* enhance Na accumulation in two wild populations of Arabidopsis. *PLoS Genetics* 2: e210.
- Nawrath C, Metraux J (1999) Salicylic acid induction-deficient mutants of Arabidopsis express *PR-2* and *PR-5* and accumulate high levels of camalexin after pathogen inoculation. *Plant Cell* 11: 1393–1404.
- Rogers E, Ausubel F (1997) Arabidopsis enhanced disease susceptibility mutants exhibit enhanced susceptibility to several bacterial pathogens and alterations in *PR-1* gene expression. *Plant Cell* 9: 305–316.
- Staswick PE (2008) JAZing up jasmonate signaling. *Trends Plant Sci* 13: 66–71.
- Staswick PE, Su W, Howell SH (1992) Methyl jasmonate inhibition of root growth and induction of a leaf protein are decreased in an Arabidopsis thaliana mutant. *PNAS* 89: 6837–6840.
- Lorrain S, Vaillau F, Balagué, Roby D (2003) Lesion mimic mutants: keys for deciphering cell death and defense pathways in plants? *Trends Plant Sci* 8: 263–271.
- Zimmermann P, Hirsch-Hoffmann M, Hennig L, Gruissem W (2004) GENEVESTIGATOR. Arabidopsis Microarray Database and Analysis Toolbox. *Plant Physiol* 136: 2621–2632.
- Gierth M, Maser P, Schroeder JI (2005) The potassium transporter AtHAK5 functions in K⁺ deprivation-induced high-affinity K⁺ uptake and AKT1 K⁺ channel contribution to K⁺ uptake kinetics in Arabidopsis roots. *Plant Physiol* 137: 1105–1114.
- Rubio F, Nieves-Cordones M, Aleman F, Martínez V (2008) Relative contribution of AtHAK5 and AtAKT1 to K⁺ uptake in the high-affinity range of concentrations. *Physiol Plantarum* 134: 598–608.
- Rubio F, Alemán F, Nieves-Cordones M, Martínez V (2010) Studies on Arabidopsis *athak5*, *atakt1* double mutants disclose the range of concentrations at which AtHAK5, AtAKT1 and unknown systems mediate K uptake. *Physiol Plant* 139: 220–228.
- Pyo YJ, Gierth M, Schroeder JI, Cho MH (2010) High-Affinity K⁺ transport in Arabidopsis: AtHAK5 and AKT1 are vital for seedling establishment and postgermination growth under low-potassium conditions. *Plant Physiol* 153: 863–875.
- Hirsch RE, Lewis BD, Spalding EP, Sussman MR (1998) A role for the AKT1 potassium channel in plant nutrition. *Science* 280: 918–921.
- Spalding EP, Hirsch RE, Lewis DR, Qj Z, Sussman MR, et al. (1999) Potassium uptake supporting plant growth in the absence of AKT1 channel activity: Inhibition by ammonium and stimulation by sodium. *J Gen Physiol* 113: 909–918.
- Xu J, Li H, Chen L, Wang Y, Liu L, et al. (2006) A protein kinase, interacting with two calcineurin B-like proteins, regulates K⁺ transporter AKT1 in Arabidopsis. *Cell* 125: 1347–1360.
- Lagarde D, Basset M, Lepetit M, Conejero G, Gaymard F, et al. (1996) Tissue-specific expression of Arabidopsis *AKT1* gene is consistent with a role in K⁺ nutrition. *Plant J* 9: 195–203.
- Jain A, Poling MD, Smith AP, Nagarajan VK, Lahner B, et al. (2009) Variations in the composition of gelling agents affect morphophysiological and molecular responses to deficiencies of phosphate and other nutrients. *Plant Physiol* 150: 1033–1049.
- Moeder W, Urquhart W, Ung H, Yoshioka K (2010) The role of cyclic nucleotide-gated ion channels in plant immunity. *Mol Plant* 4: 442–452.
- Balague C, Lin B, Alcon C, Flottes G, Malmstrom S, et al. (2003) HLM1, an essential signaling component in the hypersensitive response, is a member of the cyclic nucleotide-gated channel ion channel family. *Plant Cell* 15: 365–379.
- Clough SJ, Fengler KA, Yu IC, Lippok B, Smith RK, Jr., et al. (2000) The Arabidopsis *dnd1* “defense, no death” gene encodes a mutated cyclic nucleotide-gated ion channel. *PNAS* 97: 9323–9328.
- Jurkowski GI, Smith RK, Jr., Yu IC, Ham JH, Sharma SB, et al. (2004) Arabidopsis *DND2*, a second cyclic nucleotide-gated ion channel gene for which mutation causes the “defense, no death” phenotype. *Mol Plant Microbe In* 17: 511–520.
- Yoshioka K, Kachroo P, Tsui F, Sharma SB, Shah J, et al. (2001) Environmentally sensitive, SA-dependent defense responses in the *cpr22* mutant of Arabidopsis. *Plant J* 26: 447–459.
- Yoshioka K, Moeder W, Kang HG, Kachroo P, Masmoudi K, et al. (2006) The chimeric Arabidopsis *CYCLIC NUCLEOTIDE-GATED ION CHANNEL1/12* activates multiple pathogen resistance responses. *Plant Cell* 18: 747–763.
- Ali R, Zielinski R, Berkowitz G (2006) Expression of plant cyclic nucleotide-gated cation channels in yeast. *J Exp Bot* 57: 125–138.
- Guo KM, Babourina O, Christopher DA, Borsics T, Rengel Z (2008) The cyclic nucleotide-gated channel, AtCNGC10, influences salt tolerance in Arabidopsis. *Physiol Plantarum* 134: 499–507.
- Frietsch S, Wang Y-F, Sladek C, Poulsen LR, Romanowsky SM, et al. (2007) A cyclic nucleotide-gated channel is essential for polarized tip growth of pollen. *Proc Natl Acad Sci U S A* 104: 14531–14536.
- Leng Q, Mercier RW, Yao W, Berkowitz GA (1999) Cloning and first functional characterization of a plant cyclic nucleotide-gated cation channel. *Plant Physiol* 121: 753–761.
- Li X, Borsics T, Harrington HM, Christopher DA (2005) Arabidopsis AtCNGC10 rescues potassium channel mutants of *E. coli*, yeast and Arabidopsis and is regulated by calcium/calmodulin and cyclic GMP in *E. coli*. *Funct Plant Biol* 32: 643–653.

54. Perazza D, Laporte F, Balagué C, Chevalier F, Remo S, et al. (2011) GeBP/ GPL transcription factors regulate a subset of CPR5-dependent processes. *Plant Physiol*;doi: 10.1104/111.179804.
55. Zhu Z, Xu F, Zhang Y, Cheng YT, Wiermer M, et al. (2010) Arabidopsis resistance protein SNC1 activates immune responses through association with a transcriptional corepressor. *pollen Proc Natl Acad Sci U S A* 107: 13960–13965.
56. García AV, Blanvillain-Baufumé S, Huibers RP, Wiermer M, Li G, et al. (2010) Balanced nuclear and cytoplasmic activities of EDS1 are required for a complete plant innate immune response. *PLoS Pathog* 6: e1000970.