- 1 An agent-based model that simulates the spatio-temporal dynamics of sources and
- 2 transfer mechanisms contributing faecal indicator organisms to streams. Part 2:
- 3 Application to a small agricultural catchment

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Highlights

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- First test of agent-based model for simulating faecal indicator organisms (FIOs).
- Model shows skill in capturing transfer of FIOs from livestock to streams.
- Quantifies sources, transfer mechanisms and host animals for FIOs reaching streams.
- Potential to refine process conceptualisation (e.g. simulating FIOs from wildlife).
 - Model likely has scope for eventual incorporation into decision-support frameworks.

Abstract

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The new Model for the Agent-based simulation of Faecal Indicator Organisms (MAFIO) is applied to a small (0.42 km²) Scottish agricultural catchment to simulate the dynamics of E. coli arising from sheep and cattle farming, in order to provide a proof-of-concept. The hydrological environment for MAFIO was simulated by the "best" ensemble run of the tracer-aided ecohydrological model EcH2Oiso, obtained through multi-criteria calibration to stream discharge (MAE: 1.37 L s⁻¹) and spatiallydistributed stable isotope data (MAE: 1.14-3.02‰) for the period April-December 2017. MAFIO was then applied for the period June-August for which twice-weekly E. coli loads were quantified at up to three sites along the stream. Performance in simulating these data suggested the model has skill in capturing the transfer of faecal indicator organisms (FIOs) from livestock to streams via the processes of direct deposition, transport in overland flow and seepage from areas of degraded soil. Furthermore, its agent-based structure allowed source areas, transfer mechanisms and host animals contributing FIOs to the stream to be quantified. Such information is likely to have substantial value in the context of designing and spatially-targeting mitigation measures against impaired microbial water quality. This study also revealed, however, that avenues exist for improving process conceptualisation in MAFIO (e.g. to include FIO contributions from wildlife) and highlighted the need to quantitatively assess how uncertainty in the spatial extent of surface flow paths in the simulated hydrological environment may affect FIO simulations. Despite the consequent status of MAFIO as a research-level model, its encouraging performance in this proof-of-concept study suggests the model has significant potential for eventual incorporation into decision support frameworks.

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Key words:

60 Diffuse pollution; E. coli; EcH₂O-iso; Microbial water quality; Tracer-aided modelling; Water quality

61 modelling

1. Introduction

A prerequisite to improving impaired microbial water quality in agricultural catchments is identification of the sources and transfer mechanisms which contribute faecal indicator organisms (FIOs) to streams at the sub-field scale where mitigation measures can be implemented (Oliver et al., 2007, 2016; also c.f. Greene et al., 2015; Vinten et al., 2017). In a companion paper (Neill et al., in review), limitations were identified in using existing process-based FIO models (e.g. SWAT [Sadeghi and Arnold, 2002] and INCA-Pathogens [Whitehead et al., 2016]) to understand sub-field-scale drivers of in-stream FIO dynamics that emerge at the catchment scale. Specifically, the coarse spatial discretisations often adopted by such models are inconsistent with the scales at which processes affecting FIO fate and transport operate and at which mitigation measures can be employed (Rode et al., 2010; Wellen et al., 2015). Furthermore, as most FIO models are aggregative (i.e. they simulate stores and fluxes of FIOs integrated over spatial units such as grid cells), the ability to account for heterogenity amongst FIOs of different types and to trace pathways taken by individual FIOs to streams is limited (c.f. O'Sullivan et al., 2012; Reaney, 2008). Finally, most FIO models rely on skill in simulating stream discharge to indicate whether catchment hydrological functioning is being adequately captured (Cho et al., 2016). However, such data do not contain information on the velocities of water through a catchment that reflect flow path dynamics and hydrological connectivity, factors to which FIO transport is sensitive (Birkel and Soulsby, 2015; Wellen et al., 2015).

 Drawing on the potential offered by agent-based models for simulating individuals with heterogenous attributes that can be tracked over a simulation, Neill et al. (*in review*) reported the development of a new Model for the Agent-based simulation of Faecal Indicator Organisms (MAFIO) as an alternative approach to FIO modelling. The purpose of the model is to elucidate the sources and transfer mechanisms contributing FIOs to streams at the sub-field scale in small (<10 km²) agricultural catchments through simulating and tracking the fate and transport of agents representing FIOs (FIOagents) in a process-based, spatially-distributed manner. MAFIO consists of six sub-models that allow simulation of the following processes: 1) FIO loading from different livestock, including direct deposition in streams; 2) FIO die-off as a function of temperature and, for above-ground FIOs, solar radiation; 3) Precipitation-induced detachment of FIOs from faeces; 4) Surface routing of FIOs accounting for infiltration, exfiltration and lateral transport in overland flow; 5) Seepage of FIOs to streams from areas of degraded soil; 6) Channel routing with settling modelled by a distance-decay function for sediment-associated FIOs. A further key feature of MAFIO is that the hydrological environment used to simulate hydrological transfer mechanisms is provided by an external model; thus, there is scope for using hydrological models which can be robustly evaluated with respect to their

consistency with internal catchment states and process representation. Full details of the model, its operation and parameterisation can be found in the companion paper (Neill et al., *in review*).

 Here, MAFIO is applied to simulate the dynamics of *E. coli* in a small agricultural catchment in Scotland arising from sheep and cattle farming, in order to provide a proof-of-concept. The following specific questions are addressed:

- 103 1. To what extent can MAFIO resolve the main processes driving observed dynamics of FIOs?
- What potential does MAFIO have for providing processed-based insights into microbial water quality that are relevant for management?

Given the potential of tracer-aided ecohydrological models in providing robust simulations of catchment hydrological functioning (see Neill et al., *in review*), the model EcH₂O-iso (Kuppel et al., 2018a) is used to generate the hydrological environment for MAFIO following multi-criteria calibration to discharge and spatially-distributed isotope data.

2. Study site

The study site was the Tulloch Burn catchment (0.42 km²; Figure 1a), a sub-catchment of the Tarland Burn (71 km²) which is a tributary of the River Dee, NE Scotland. The Dee is a regional water resource, supplying >300,000 people with drinking water, and is designated a Special Area of Conservation due to the freshwater ecosystem it supports. Higher intensities of agriculture in lowland tributaries of the Dee have been linked to impaired water quality (Langan et al., 1997). As the most upstream tributary draining significant areas of agriculture, the Tarland Burn catchment became a research site for assessing diffuse- and point-sources of pollution and evaluating best management practices for mitigation (Bergfur et al., 2012). The selection of the Tulloch Burn catchment for this study was based on work that identified it as a "hot spot" for faecal contamination from 11 years of *E. coli* data (Neill et al., 2018).

Longer-term data (2000-2010) from Aboyne meteorological station ~10 km from the Tulloch shows mean annual precipitation and potential evapotranspiration for the area to be 828 mm and 521 mm, respectively (Dunn et al. 2013). Catchment elevation ranges from 216 m to 453 m. Brown earths (41%)

and humus-iron podzols (34%) are the predominant soil-types (Figure 1b; Soil Survey of Scotland Staff, 2014). These are freely-draining soils; consequently, artificial field drainage is not necessary in the catchment. There are also limited areas of non-calcareous gleys and alluvial soils, with higher elevations dominated by peaty-gleyed podzols (Figure 1b; Soil Survey of Scotland Staff, 2014). Approximately 60% of the catchment is agricultural (Figure 1c). During the study, surveys showed that five fields were used for pastoral (sheep and cattle) farming (Lower/Mid Pasture [L/R] and Top Pasture) and two for arable (Lower/Upper Arable). Apart from Mid-Pasture (R), field boundaries extend beyond the Tulloch Burn catchment. Of the remaining catchment, 24% is mixed-conifer forest and 16% is heather moorland (Figure 1c).

To prevent livestock access, parts of the stream are fenced-off from fields and surrounded by small riparian areas (Figure 1d). However, stretches of the stream running through Mid Pastures (R) and (L) are directly accessible to livestock, with animals able to move between Mid Pastures (R) and (L) at a discrete stream crossing-point depending on whether a gate is open (Figure 1c and e). Other discrete crossing-points can also connect Lower Pastures (R) and (L), and Lower Pasture (L) and Mid Pasture (R), again depending on gates (Figure 1c). Observation during the study found a high degree of soil compaction around all three crossing points (DS1-3 in Figure 1c) due to the concentration of livestock moving through these areas. This resulted in the soils being in a state of semi-permanent saturation (Figure 1f); therefore, these areas of degraded soil can continually seep water to the stream and may be a potential source of chronic faecal contamination (e.g. Bilotta et al., 2007).

3. Data and methods

3.1 Hydrometric and isotope data

Hydrometric monitoring at the Tulloch Burn started in October 2016. The main study period was between 27/04/17 and 31/12/17. Daily average discharge at the catchment outlet and sites T6 and T8 was derived by area-scaling discharge measurements made at the outlet of the 3.9 km^2 Blackmill Burn catchment within which the Tulloch is nested. Specifically, near-concurrent discharge (*Q*) measurements made under identical hydroclimatic conditions at five sites within the Blackmill Burn (including the outlet of the Tulloch) with catchment areas (*A*) of 0.2- 3.9 km^2 revealed a strong relationship between discharge and area ($Q = 1.85e-8 \times A^{0.97}$; Adj. R²: 0.99). Consequently, discharges for sites within Tulloch Burn could be derived as:

 $Q_{TulX} = Q_{BM} \cdot \left(\frac{A_{TulX}}{A_{BM}}\right)^{0.97}$ Eq. 1

where Q_{TulX} and A_{TulX} are discharge and catchment area for Site X in the Tulloch Burn, respectively, and Q_{BM} and A_{BM} are discharge and catchment area of the Blackmill Burn, respectively. This was necessary as the narrow, poorly-defined channel of the Tulloch prevented a reliable stage-discharge rating curve. The Blackmill and Tulloch Burns have comparable soils and land use causing them to exhibit similar hydrological responses. Meteorological data (precipitation, temperature, relative humidity and windspeed) were collected at 15-mintue intervals using an automatic weather station within the catchment (Figure 1a). Short- and long-wave radiation were obtained from ERA-Interim climate reanalysis (Dee et al., 2011). These data were amalgamated into daily timeseries.

Isotope samples were analysed for δ^2H and $\delta^{18}O$ using a Los Gatos laser isotope analyser (precision: \pm 0.4‰ for δ^2H and 0.1‰ for $\delta^{18}O$). Given higher relative precision, δ^2H was used here. Daily streamwater samples at the catchment outlet were collected for isotope analysis using an ISCO-3700 autosampler from 27/04/17 (Figure 1a). A layer of paraffin was added to bottles to prevent evaporation. Synoptic grab-sampling for isotopes occurred on a twice-monthly basis at sites T2-8 (Figure 1a). During the microbial observation period (Section 3.2), samples were taken twice-weekly at T6 and T8 to be coincident with samples taken for *E. coli* analysis. Daily bulk samples of precipitation were also collected using an ISCO-3700 autosampler.

3.2 Microbial and livestock count data

Within the study, a more intense field campaign was carried out between 08/06/17 and 31/08/17 (the "microbial observation period" – MOP) to collect higher-temporal-resolution data for stream *E. coli* concentrations and livestock counts. This corresponded to when most livestock are in the fields and potential for faecal contamination is elevated (Kay et al., 2008). Twice-weekly sampling for *E. coli* occurred during this period at the outlet, and from 06/07/17 at T6 and T8 to characterise spatial variability in concentrations arising from differing catchment characteristics (Figure 1a). Samples for *E. coli* were collected (working upstream to T8) in glass bottles sterilised by autoclaving at 123 °C for 20 minutes. Care was taken not to disturb the channel bed to prevent contamination from *E. coli* stored in the sediment. Samples were placed in cool boxes until processing began within 6 hours of collection. Concentrations of *E. coli* were determined using the Colilert-18 most-probable-number (MPN) method (IDEXX Laboratories, Westbrook, Maine, USA). Samples were well-shaken to ensure uniform distribution of *E. coli* prior to 100 ml being decanted to provide concentrations in MPN 100 ml⁻¹. When

high concentrations of *E. coli* were likely, dilutions were made using sterile Ringers' solution. The limit of detection for undiluted samples was 1 MPN 100 ml⁻¹. *E. coli* loads (MPN d⁻¹) were derived by multiplying observed concentrations of *E. coli* at the outlet, T6 and T8 by average daily discharge for each respective site.

Number and type of livestock in each agricultural land parcel of the catchment (Figure 1c) was recorded each sampling day. Where a land parcel represented a field with boundaries that extended beyond the Tulloch Burn catchment, the total number of livestock in the whole field was scaled by the fraction of the field falling within the land parcel. This assumed livestock would be uniformly distributed within a given field (c.f. Dorner et al., 2006; Haydon and Deletic, 2006). If livestock could move between land parcels, then the total number of livestock in all connected parcels was counted and scaled to each individually based on the fraction of the total connected area they represented. In addition, whether gates prevented livestock access to the stream at crossing points DS1-3 (Figure 1c) was also recorded. The remainder of the stream was either fenced off and inaccessible to livestock, or, for stream sections in Mid Pastures (L) and (R), permanently open to livestock (Figure 1c and e). Daily timeseries of stream access and livestock counts were generated from the twice-weekly observations by assuming any changes occurring between successive observation days did so at the mid-point between them.

3.3 Setup of EcH₂O-iso for the Tulloch Burn

In EcH₂O-iso, the spatial grid for simulations is defined by a digital elevation model (Kuppel et al., 2018a). A 5×5 m resolution LandMap digital terrain model (DTM) was resampled to 30×30 m resolution for delineating the Tulloch Burn catchment and deriving local slopes and drainage directions. The relatively coarse spatial resolution was necessary to keep model runtimes manageable given the long spin-up period needed for simulated water ages to stabilise. Simulation of 22 years with a daily timestep was necessary, with the period 27/04/17 to 31/12/17 for the last year retained for further analysis. The 22-year spinup was achieved by looping meteorological and isotopic inputs for 2016-2017 eleven times (c.f. Hrachowitz et al., 2010). Meteorological data before establishment of a catchment weather station (October 2016) were derived from the adjacent Aboyne and Bruntland Burn stations using statistical relationships from periods of overlapping data, whilst radiation for all of 2016 was available from the ERA-Interim climate reanalysis. For altitudinal effects on precipitation and temperature, a 5.5% increase in precipitation (Ala-aho et al., 2017) and decrease of 0.6 °C (Goody and Yung, 1995) with every 100 metre elevation gain was implemented.

Parameterisation of soil hydrological properties in EcH₂O-iso was based on the five mapped soil types in the catchment (Soil Survey of Scotland Staff, 2014; Figure 1b). Soil properties were assumed to be uniform within each type. To facilitate parameterisation of vegetation, land parcels of the Tulloch Burn in Figure 1c were divided into three categories: agriculture, forest and heather moorland. Based on local knowledge, agricultural areas were assumed to comprise 95% grass and 5% bare soil, forested land was assumed to comprise 68% conifers, 30% grasses and 2% bare soil, and heather moorland was assumed to comprise 95% heather and 5% bare soil. To identify parameters for calibration, an initial sensitivity analysis was undertaken following the method of Morris (1991) and Sohier et al. (2014) using eight trajectories and a radial step for evaluating the parameter space. This identified 11 soil-related, 13 vegetation-related and two channel-related parameters as sensitive (Table S1), resulting in the need to calibrate 96 individual parameter values ([11×5]+[13×3]+2=96). Values of fixed parameters are given in Table S2.

3.4 Multi-criteria calibration of EcH₂O-iso

Calibration of the 96 parameter values followed a multi-criteria approach incorporating stream discharge (outlet) and δ^2H (outlet + sites T2-8) as calibration targets. Latin Hypercube Sampling was used to generate 100,000 parameter sets for EcH₂O-iso, based on the sampling ranges given in Table S1. For each model run, mean absolute errors (MAEs; Willmott and Matsuura, 2005) were calculated to quantify the skill of the run in simulating the dynamics of each calibration target for the period 27/04/17 to 31/12/17. For discharge, use of MAE avoided overemphasis on high-flows typical of alternatives such as the Nash-Sutcliffe efficiency statistic (Krause et al., 2005; Legates and McCabe, 1999), whilst for isotopes the limited variability in observations and daily timestep of the model necessitated use of a measure of average error (c.f. Gupta et al., 2009; Schaefeli and Gupta, 2007). A single performance metric for each model run was then derived by combining MAEs for individual calibration targets via a weighted-addition (e.g. Beven, 2012). This allowed the number of observations for each calibration target to determine the influence of its associated MAE on defining overall performance of the run and enabled identification of a "best" run for use in providing the hydrological environment for MAFIO (Section 3.5.2).

As MAE is dimensional with an optimal value of 0, it was necessary to convert the MAEs associated with each calibration target into dimensionless metrics that monotonically increase with model performance, prior to implementing the weighted addition. The latter need was met by calculating the metric (1-MAE), which increases with model performance to an optimum of 1 (in both instances 1 is in units of the calibration target). To remove dimensionality and obtain a metric (MAE^*) for use in the weighted addition, the following equation was applied:

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$$MAE_{i,j}^* = \frac{\left(1 - MAE_{i,j}\right) - \min(1 - MAE)_j}{\max(1 - MAE)_j - \min(1 - MAE)_j}$$
 Eq. 2

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263 where $MAE_{i,j}$ is the MAE associated with calibration target j for model run i and $(1-MAE)_j$ is the 264 complete set of (1-MAE) associated with calibration target j from all 100,000 model runs. For a given 265 run, a final goodness of fit in the range [0,1] was obtained through the weighted addition:

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$$GOF_i = \sum_{j=1}^{n} W_j . MAE_{i,j}^*$$
 Eq. 3

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269 where GOF_i is the goodness of fit value of run i and W_j is the weighting given to the performance metric 270 MAE^* associated with calibration target j. The weighting of a performance metric associated with a 271 given calibration target was the fraction of observations for all calibration targets that it contained. 272 Numbers of observations and consequent weightings for each target used in the calibration are detailed 273 in Table 1. Following calibration, an ensemble of the 100 model runs with the highest GOF values

(behavioural runs) were retained to examine model performance and uncertainty.

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- 276 3.5 Setup of MAFIO for the Tulloch Burn
- As a first test of the model, MAFIO was used to simulate the behaviour and transport of E. coli in the
- Tulloch Burn during the MOP. This section describes the setup of the catchment and hydrological
- environments of the model (see Section 3.2 of Neill et al., *in review*) and its parameterisation for E.
- 280 *coli*.

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- 282 3.5.1 Catchment environment
- environment. Catchment extent and local drainage directions were defined from the 30×30 m DTM (identical to EcH₂O-iso). The spatial distribution of land parcels and of cells containing degraded soil / the channel were defined as shown in Figure 1c. To facilitate representation of sub-grid heterogeneity in the latter, channel widths were derived from linear interpolation between field measurements of bankfull width. In addition, land parcels either adjacent to the stream (stream-associated land parcels)

Table 3 of Neill et al. (in review) outlines the inputs necessary to characterise the catchment

or from which livestock could contribute to soil degradation were defined as those falling within the 30×30 m footprint of a given cell as determined from field observations and aerial imagery (Table S3). The cell(s) immediately upslope of those containing degraded soil / the channel were defined from local drainage directions (Figure S1). Timeseries of livestock counts and access to the stream were as described in Section 3.2.

3.5.2 Hydrological environment

The hydrological environment of MAFIO was simulated by the "best" overall ensemble run of EcH₂O-iso. To qualitatively assess the potential for uncertainty in the outputs of EcH₂O-iso to impact MAFIO simulations, spatial patterns of surface and groundwater flow paths and soil saturation deficit simulated over the MOP by the "best" run and by the 100 behavioural runs were compared. This indicative approach was taken as a full quantitative uncertainty analysis would require using the 100 behavioural EcH₂O-iso runs to generate the hydrological environment for an ensemble of MAFIO runs, the latter necessary to account for the effect of stochasticity (see Section 3.6). Such an uncertainty analysis was beyond the scope of this initial proof-of-concept test of MAFIO; however, this will be a focus of future work. To aid in assessing process representation in MAFIO, the simulated hydrological environment was characterised by generating spatial summaries of discharge, δ^2 H and water ages in the stream, and of overland flow, soil saturation deficit and groundwater fluxes within the catchment, for the whole MOP and for exemplar "dry" (10/08/17) and "wet" (15/08/17) days. Here, streamwater ages denote how long water contributing to discharge spent travelling through the catchment since entering it as precipitation (Sprenger et al., 2019).

3.5.3 Parameterisation of MAFIO sub-models

Parameter values for the MAFIO sub-models are presented in Table 4 of Neill et al. (*in review*). Sheep and cattle were the only livestock reared in the catchment. Most parameter values were taken from the extensive literature on *E. coli* as an FIO. Exceptions were the parameters *faecesConc* (concentration of FIOs per gram of faeces) and *agentsRepresent* (number of FIOs shed in reality for which an FIO-agent is introduced into the simulation). For the former, geometric mean concentrations of *E. coli* in sheep and cattle faeces were determined from faecal samples collected in the catchment during 2017 and 2018 (Avery et al., *unpublished data*). Meanwhile, the minimum value of *faecesConc* (4.18×10⁵) was used for *agentsRepresent*. This was the minimum permissible value given available computational resources; however, similarities between simulations obtained using this value and a value of 1×10⁸ *E. coli* (*Data not shown*) suggested that significant changes to model outputs would be unlikely if a smaller value were to be used. This provided confidence that a sufficient quantum of *E. coli* was simulated by MAFIO.

Despite potential uncertainty in the values of model parameters (Oliver et al., 2016), calibration was not undertaken in this initial application. This was primarily because use of stochasticity to model processes conceptualised in ABMs hinders the use of "fit-to-data" metrics (i.e. those quantifying model skill in reproducing dynamics of observed data) often used in automated calibration (Polhill and Salt, 2017). This is further described in Section 3.6. A benefit to using the model uncalibrated is that it offers insight into the process consistency of the model if physically-meaningful parameter values are used (c.f. Kuppel et al., 2018a), as compensatory parameter effects on model structural deficiencies from calibration are avoided (c.f. Beven, 2019).

3.6 Application of MAFIO

MAFIO was applied for the MOP on a 30×30 m spatial grid using a daily time step, consistent with the spatio-temporal resolution of data characterising the catchment and hydrological environments. For initialisation, values for the fraction of damaged soil (dFrac) at DS1, DS2 and DS3 were set to 0.72, 0.53 and 0.7155, respectively, based on application of Eq. 3 of Neill et al. ($in\ review$) with livestock counts made in the catchment since 01/01/17. As these counts also showed all pasture land parcels to have been grazed to some extent since the start of 2017, all were set to have FIO-agents already in the soil at initialisation as a significant soil reservoir of $E.\ coli$ may persist for several months even after cessation of grazing (Muirhead, 2009). Initial numbers of FIO-agents were based on estimates of the total number of $E.\ coli$ in the upper soil of each land parcel (between 4.1×10^9 - $9.3\times10^{10}\ E.\ coli$). These were approximated from concentrations of $E.\ coli$ (in MPN g^{-1}) measured in the top 5 cm of soil at five locations within the Lower and Mid Pasture fields (Avery et al., $unpublished\ data$) and estimates of the weight of soil in each land parcel over the same depth.

An ensemble of 30 model runs using the same parameterisation and input was made for the MOP to characterise stochastic variability in MAFIO outputs (Abdou et al., 2012). Combined with the fact that natural variability intrinsic to complex systems can cause observed data to be conditional on a particular trajectory having been taken by the system, of which many may have been possible (Refsgaard et al., 2007; Windrum et al., 2007), this stochastic variability characteristic of ABM outputs can complicate assessments of model performance (Brown et al., 2005). In particular, traditional "fit-to-data" metrics become inappropriate as exclusive means of evaluating ABM performance as a model that adequately represents the processes giving rise to observations could be unfairly penalised if, due to stochastic treatment of system processes, it simulates a range of plausible scenarios which may or may not include what was observed (Polhill and Salt, 2017). Whilst a "benchmark" alternative is yet to emerge, other ABM work has highlighted the value of combining quantitative performance evaluation with qualitative "checks" that focus on understanding how simulated characteristics at larger scales emerge

from the processes influencing the behaviour of individual agents and assessing the plausibility of such processes similarly effecting the phenomena under investigation in reality (Moss and Edmonds, 2005; Polhill et al., 2010; Polhill and Salt, 2017).

Consequently, the following approach to performance assessment was adopted. For each ensemble run, observed and simulated E. coli loads were compared quantitatively at the outlet, T6 and T8. Since MAFIO simulates fluxes of FIO-agents, simulated loads were approximated by multiplying FIO-agent fluxes by the value of agentsRepresent. As this parameter dictates the precision to which observed loads can be simulated (c.f. Parry and Bithell, 2012), the skill of MAFIO in capturing periods of relatively more or less impaired microbial water quality was also assessed (c.f. Oliver et al., 2010; Porter et al., 2017). This was achieved by calculating Z-scores showing the number of standard deviations an observed E. coli load or simulated flux of FIO-agents was away from the mean of its associated timeseries. As observations were not available for all dates, Z-scores for simulated FIO-agent fluxes were based only on simulations that overlapped with observations. Spearman's rank correlation coefficients were also used to assess how well MAFIO captured the relative order of observed E. coli loads at each site (c.f. Porter et al., 2017). For a qualitative "check" on the model, the plausibility of simulated outputs given the potential processes influencing E. coli dynamics in the Tulloch Burn was the subject of a literature-based discussion (Section 5.1) that also considered the performance of EcH₂Oiso in simulating the hydrological environment. Whilst such qualitative evaluation is less robust than alternative methods based on consultation of independent experts (e.g. Moss and Edmonds, 2005; Polhill et al., 2010), the latter was beyond the scope of this initial MAFIO application.

As a basis for assessing the potential of MAFIO in providing insights relevant to management, the overall flux of FIO-agents leaving the catchment in the stream ("exported FIO-agents") was observed and further disaggregated into contributions from different livestock types and transfer mechanisms. The latter were derived from the attributes *Domain type* and *Livestock type* (see Table 2 of Neill et al., *in review*) of exported FIO-agents. By observing the *Location memory* attribute, areas where exported FIO-agents entered the stream via direct deposition and pathways taken by exported FIO-agents reaching the stream in overland flow or seepage were also identified. For each timestep, pathways were derived by quantifying the total number of exported FIO-agents that had passed through each grid cell in overland flow or seepage at any point whilst being transported from their original spawning location to the stream (i.e. pathways reflect the transport of exported FIO-agents over their entire existence, not just during the timestep in which they were exported). Meanwhile, spatial patterns of direct deposition were characterised by quantifying the total number of exported FIO-agents entering the stream via direct deposition for each cell containing a channel. For each of the 30 ensemble runs of MAFIO, timestep

totals were extracted for the example dry and wet days and further summed over the whole MOP. Median totals across the ensemble runs for each cell were then used to generate "average" maps of direct deposition and pathways taken to the stream by exported FIO-agents for each period of interest, whilst maps using ranges in totals were generated to evaluate the effect of stochastic variability in ensemble simulations.

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4. Results

- 4.1 Hydrometric and isotope observations
- 401 Hydroclimatic conditions in 2017 were typical for the region (Hannaford et al., 2018). The study period
- started out relatively dry, with only 23 mm of precipitation falling by the end of May (Figure 2a).
- 403 Consequently, summer baseflows were established by mid-June (Figure 2b), despite the largest
- 404 precipitation event of the study (37.2 mm d⁻¹) occurring at the beginning of that month, two days before
- 405 the MOP commenced. For the remaining summer, precipitation fell in low-intensity events that
- 406 generated small discharge responses (Figure 2a-b and 3a). Sustained periods of precipitation in mid-
- September re-wetted the catchment leading to a rise in baseflows and the largest discharge responses
- 408 (up to 31 L s⁻¹) being observed in November (Figure 2a-b).

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- The δ^2 H composition of precipitation ranged between -153.1‰ and -14.9‰ (Figure 2a). By contrast,
- the δ^2 H composition of stream water at the outlet was substantially damped (range -64.2% to -55.0%),
- 412 though deviations in the direction of the precipitation signal were observed during events (Figure 2b).
- At T2-8, δ^2 H was similarly damped, ranging between -61.0% and -54.8% across all sites (Figure 2c).
- These sites behaved similarly to the outlet in terms of variability; however, the upper T7 and T8 sites
- had slightly more enriched δ^2 H values, with values then becoming more depleted towards the outlet
- 416 (Figure 2c). Daily average air temperatures peaked at ~18 °C in May and June and fluctuated ~12 °C
- 417 until September when temperatures fell towards a minimum of -3 °C in December (Figure 2d). Solar
- 418 radiation followed a similar trajectory (Figure 2d).

- 420 *4.2 Microbial observation period (MOP)*
- 421 Microbial observations began as the hydrograph experienced a small secondary peak in response to
- precipitation that occurred following the 37.2 mm d⁻¹ precipitation event (Figures 2a-b and 3a). Peak
- daily average temperature and solar radiation for the overall study period fell within the MOP, with the
- 424 former averaging 12.7 °C and the latter 176 W m⁻² (Figure 3b).

 Temporal dynamics of *E. coli* concentrations and loads at individual sites were similar (Figure 3c-d). At the outlet, concentrations ranged from 2.3×10^1 to 2.0×10^3 MPN 100 ml⁻¹ (Figure 3c), whilst loads varied between 4.9×10^7 and 4.1×10^9 MPN d⁻¹ (Figure 3d). Both were highest towards the end of the MOP and lowest during early July. The latter coincided with no livestock present in fields closest to the catchment outlet (Figures 1c and 3f), suggesting that this period may have been characterised by background concentrations of *E. coli*. At T6, concentrations and loads varied between 4.4×10^1 and 9.1×10^3 MPN 100 ml⁻¹ and 5.0×10^7 and 8.7×10^9 MPN d⁻¹, respectively (Figure 3c-d). Thus, whilst T6 and the outlet generally experienced similar concentrations and loads of *E. coli*, these could be higher at the former site. T8 was generally the least-contaminated site as concentrations of *E. coli* were <10 MPN 100 ml⁻¹ on over half the sampling days (Figure 3c). However, concentrations could increase to 10^2 MPN 100 ml⁻¹, most frequently towards the end of the MOP. Loads varied between 5.8×10^5 and 3.9×10^8 MPN d⁻¹ (Figure 3d) and were, consequently, often much smaller than loads observed at the outlet and T6 (exceptions are the last three sample dates of the MOP where loads at T8 were greater than at T6). A clear response of *E. coli* concentrations and loads to discharge was elusive; however, a link to livestock counts was more apparent at the outlet and T6.

Timeseries showing stream accessibility to livestock at the three discrete crossing points are shown in Figure 3e. Sheep were the main livestock in the catchment and were in Lower Pasture (L), Mid Pasture (R), Top Pasture or, for a short time late in the MOP, Lower Pasture (R) (Figure 3f). Cattle were briefly present in Mid Pasture (R) from late June to early July (Figure 3f).

4.3 Multi-criteria calibration of EcH₂O-iso

The results of calibrating EcH₂O-iso to discharge and spatially-distributed isotope data are summarised in Table 2 and Figure 4; calibrated parameter ranges are given in Table S1. Discharge was generally well-simulated (Table 2), with behavioural model runs successfully capturing the summer baseflows and small events that characterised the MOP alongside the re-wetting of the catchment in September and timing of the largest discharges in November (Figure 4b). The magnitudes of the latter were, however, under-estimated, as were discharges at the start of the simulation period (Figure 4b). For isotopes, the model could reproduce the markedly-damped composition of streamwater at the outlet (Table 2; Figure 4c). Isotopic variability in response to precipitation was also generally well-captured, though more extreme excursions could be simulated. Skill in simulating isotope dynamics at the synoptic sampling sites was more variable (Table 2; Figure 4d-j), likely reflecting their lower weighting in the multi-criteria calibration (Table 1) and the sparser temporal resolution of observations.

Performance was best for sites closer to the outlet, whilst simulations for sites further upstream showed greater uncertainty with some ensemble runs exhibiting poorer performance. However, performance was still maintained in the "best" overall run, with MAEs not exceeding ~3‰ at the upstream sites (Table 2). Overall, it was encouraging that EcH₂O-iso generally reproduced the damped isotope signals observed at the synoptic sampling sites.

4.4 Characterisation of the hydrological environment

Median discharges over the MOP (Figure 5a) simulated by the "best" run of EcH₂O-iso decreased in age with distance downstream (Figure 5e) but did not exhibit a clear spatial pattern in δ^2 H (Figure 5c). Median streamwater ages were relatively old (averaging ~1.5 years), reflecting the dominant simulation of groundwater fluxes over surface water fluxes. Overland flow was only simulated for very restricted areas, mainly limited to stream-proximal cells in the lower part of the catchment (Figure 5b). Over the MOP, cells for which overland flow was simulated generated total fluxes <1000 mm. By contrast, simulated groundwater fluxes over the MOP could be up to 4700 mm for individual cells and occurred across much of the catchment (Figure 5f). The limited extent of simulated overland flow arose from much of the soil in the catchment being in saturation deficit (Figure 5d). Highest deficits were generally simulated in the upper catchment in areas of forest and heather (Figure 1c) underlain by podzolic soils (Figure 1b); here, overland and groundwater fluxes were consequently minimal (Figure 5b and f).

Stream discharges simulated for the example wet day were higher than for the dry day (Figure 6a) and consisted of younger water (median age of streamwater 399 days vs. 522 days; Figure 6e). The isotopic composition of the stream was also generally more enriched in wet conditions (Figure 6c). This, together with the lower streamwater ages, indicates increased contributions of younger overland flow and soil water to streamflow during summer wet conditions compared with the dominance of older groundwater during drier periods. However, even when wet, overland flow remained spatially limited, with just a few cells adjacent to the stream and a small number of distal, unconnected cells simulating fluxes of up to 35.7 mm d⁻¹ (Figure 6b). In dry conditions, overland flow was generated from more restricted areas that maintained saturation by virtue of their position in flatter parts of the riparian area (Figure 6b). Overland flow was limited due to most soil being in saturation deficit in both dry and wet conditions (Figure 6d). In contrast, groundwater fluxes were active across similar spatial areas on the wet and dry days, with fluxes of up to 57.8 mm d⁻¹ and 35.7 mm d⁻¹ simulated for each day, respectively (Figure 6f).

 Assessment of the spatial outputs from the 100 behavioural runs of EcH₂O-iso revealed that generation of overland flow from areas proximal to the stream and the dominance of groundwater simulated by the

"best" run was also simulated with reasonable certainty (i.e. in >50% of runs) by the ensemble (Figure 7a-c). In addition, relative spatial patterns of moisture deficits were comparable (Figure 7d). However, it was possible for some behavioural runs to simulate larger areas of overland flow generation, leading to uncertainty in the exact spatial extent of surface flow paths (Figure 7c). The implications of this will be discussed with respect to assessing the adequacy of process conceptualisation in MAFIO.

4.5 Performance of MAFIO

Simulated *E. coli* loads at the outlet captured the main dynamics of observations quite well; in particular, the observed decrease in loads in early July and subsequent increase, relatively constant loads in early-to mid-August, and brief dip in loads towards the end of the MOP were all simulated (Figure 8a). However, there was a general tendency for loads to be over-predicted, with the main exceptions being at the end of the MOP and when observed loads decreased in July (Figure 8a). Z-scores at the outlet show that the model was more successful in capturing when loads were above- and below-average, with the sign of the Z-scores simulated correctly in the majority of cases (Figure 8b). Exceptions were at the end of the simulation (reflecting over-prediction of observed loads earlier on despite absolute loads at the end of the MOP being successfully captured) and on 03/07/17. Spearman's rank correlations ranged between 0.21 and 0.30 with an average of 0.26 across the 30 ensemble runs. Stochastic variability in outlet simulations was minimal (Figure 8a-b).

At T6, all ensemble runs simulated loads of zero and Z-scores below 0 whenever livestock were absent from Mid Pastures (R) or (L) (Figures 3f and 8c-d). Non-zero loads were only simulated when Mid Pasture (R) had livestock present (Figures 3f and 8c); however, simulated loads and Z-scores exhibited a high degree of stochastic variability (Figure 8c-d). Consequently, Spearman's rank correlations varied between runs, ranging from -0.12 to 0.32. When livestock were present in Mid Pasture (R), observations generally fell within simulation bands; however, loads were under-estimated when livestock were absent (Figure 3f and 8c). For relative performance, observed Z-scores were often within the range simulated (Figure 8d). When non-zero loads were simulated, the large range in simulated Z-scores meant that MAFIO was not consistent between ensemble runs in terms of simulating loads above- or below-average. At the end of the period, observed and simulated Z-scores were similar, suggesting that MAFIO successfully captured this as a time of relatively less-impaired microbial water quality.

Ensemble runs always simulated zero loads at T8 (Figure 8e). Consequently, neither Z-scores for MAFIO simulations nor Spearman's rank correlations could be calculated. Qualitatively, the simulated behaviour was largely consistent with the low concentrations and loads of *E. coli* at T8 (Figure 3c-d),

and negative observed Z-scores (Figure 8f). Quantitatively, however, the simulation of zero loads was not consistent with observations.

4.6 Sources and mechanisms contributing E. coli to streams

To help interpret timeseries relating to exported FIO-agents, simulated effective precipitation, discharge at the outlet and storage of FIO-agents are shown in Figures 9a-b. The simulated flux of exported FIOagents (Figure 9c) strongly reflected storage dynamics of FIO-agents (Figure 9b) and livestock counts (Figure 3f). Exported FIO-agents predominantly entered the stream via seepage from degraded soil (Figure 9d) as simulated overland flow was limited (Figure 5). When overland flow did transfer FIOagents to the stream, localised spikes in export fluxes were simulated (Figure 9c-d). Contributions were non-linear due to the changing storage of FIO-agents. When cattle were in Mid Pasture (R), both seepage and direct deposition made their greatest contributions of FIO-agents to the stream (Figures 3f and 9d), reflecting the higher loading rates of E. coli from cattle (see Table 4 in Neill et al., in review) and the extensive stream access in this field (Figure 1c). Variability in flux magnitudes between ensemble runs was also greatest at this time (Figure 9c). Contributions from direct deposition were otherwise minimal, reflecting the lower loading rates of sheep (see Table 4 in Neill et al., in review) when present in Mid Pasture (R) or the limited number of discrete crossing points allowing stream access to livestock in other fields (Figures 1c and 3e). Contributions from different livestock types largely corresponded to their presence in the catchment (Figures 3f and 9e); however, a limited "memory-effect" in contributions reflected survival of FIO-agents in areas of degraded soil.

Over the MOP, similar median numbers of exported FIO-agents entered the stream via direct deposition wherever livestock were present with stream access (Figures 1c, 3f and 10a). However, stochastic variability across the ensemble runs was evident (Figure 10b). Pathways taken by exported FIO-agents reaching the stream in overland flow or seepage were constrained by the limited area over which the former was generated in the catchment (Figures 5 and 10c). Fluxes of exported FIO-agents along individual pathways increased towards the stream with generation of overland flow (Figures 5b and 10c); however, short pathways between areas of degraded soil and the stream were consistently followed by the largest numbers of exported FIO-agents (Figures 1c and 10c). Overall, source areas contributing exported FIO-agents to the stream over the MOP were always restricted to stream-proximal locations (Figure 10c-d). Stochastic variability in numbers of exported FIO-agents following particular pathways to the stream was usually less than for numbers being directly deposited in the stream (Figure 10b and d).

On the example dry and wet days, there was a difference in the spatial distribution of cells for which the median number of exported FIO-agents entering the stream via direct deposition was non-zero (Figure 11a and c). However, similar counts of sheep in Lower Pasture (L) and Mid Pasture (R) on both days (Figure 3f) meant that when non-zero, median numbers were comparable (Figure 11a and c). In addition, stochastic variability across ensemble runs showed that on either day, direct deposition of exported FIO-agents could occur wherever livestock had stream access (Figure 11b and d). More extensive overland flow on the wet day (Figure 6b) increased the spatial extent of pathways taken by exported FIO-agents to the stream, although contributing areas were always limited to near-stream locations (Figure 11e and g). Fluxes of FIO-agents along individual pathways were elevated in wet conditions (Figure 11g) due to the increased generation of overland flow (Figure 6b). For both the dry and wet days, "average" maps indicated pathways taken by exported FIO-agents to the stream (Figure 11e and g) that fully consisted of cells where overland flow was simulated (Figure 6b) or seepage from degraded soil was possible (Figure 1c). However, from considering the effect of stochastic variability between ensemble simulations (Figure 11f and h), it is apparent that in some instances, exported FIOagents had followed paths that included cells not hydrologically connected to the stream during the timestep in question (e.g. in Lower Pasture [L]; Figures 1c and 6b). This indicates that FIO-agents previously moved and infiltrated into the soil could be exfiltrated and further transported. Stochastic variability in numbers of exported FIO-agents following particular pathways to the stream was generally lower overall for the dry event (Figure 11f and h).

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5. Discussion

5.1 To what extent does MAFIO resolve the main processes driving observed FIO dynamics?

Using models to explore issues of water quality, especially in a decision-making context, requires confidence that processes governing the determinand of interest are adequately captured (c.f. Vaché and McDonnell, 2006; Wellen et al., 2015). Consequently, the new agent-based model MAFIO was applied to the Tulloch Burn for a relatively data-rich period, in order to assess the adequacy of process representation in the model. The spatially-distributed, tracer-aided ecohydrological model EcH₂O-iso provided the hydrological environment to improve confidence in the robustness of simulated hydrological processes underpinning FIO simulations (c.f. Birkel and Soulsby, 2015; Neill et al., 2019). However, whilst multi-criteria calibration to discharge and isotope data allowed elements of catchment hydrological functioning to be reasonably constrained (i.e. dominance of groundwater, generation of overland flow proximal to the stream), uncertainty persisted in the exact spatial extent of overland flow paths (Figure 7c). This can obscure whether deficiencies in FIO simulations arise from uncertainty in simulated hydrology or the need to refine process conceptualisation in MAFIO; where this may be an issue is highlighted in the discussion that follows. This also necessitates that a full quantitative

uncertainty analysis be the subject of future work (c.f. Beven and Lamb, 2017), and reinforces the need for collection of diverse datasets for use in constraining highly-parametrised models (Kelleher et al., 2017; Kuppel et al., 2018b).

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The consistent simulation of zero loads of E. coli at T8 despite non-zero loads being observed likely indicates that process conceptualisation in MAFIO itself requires refinement (Figure 8e). This assertion arises from confidence in the lack of surface connectivity between T8 and Top Pasture (the only upstream source of livestock-derived FIOs) simulated by the "best" run of EcH₂O-iso (Figure 5b), despite the overall uncertainty in the exact extent of overland flow paths (Figure 7c). Specifically, the skill of this run in capturing observed isotope dynamics at T8 (Figure 4j) indicates that no overland flow upstream of this site is plausible. Furthermore, increased tree water use in the forest separating T8 and Top Pasture (Douinot et al., 2019) combined with likely enhanced filtering of FIOs in overland flow by the forest floor (Kay et al., 2012) suggests limited opportunities for surface transport of FIOs between these locations. Given that deer and hares have been observed in the forest around T8, a possible refinement to MAFIO could be inclusion of wild animals as sources of FIOs. Indeed, wildlife (including gastropods, frogs and fish as previously unrecognised sources; Frick et al., 2018) have previously been found to significantly impact microbial water quality, even in agricultural areas (Muirhead et al., 2011). An alternative refinement could be accounting for possible sources of "naturalised" FIOs that have adapted to persist and grow in the environment (e.g. Jang et al., 2017). A key issue to consider when conceptualising the former would be the increased uncertainty in loading and die-off parameters associated with FIOs from wildlife (Guber et al., 2015). Furthermore, difficulties also exist in characterising population levels and movement of wild animals in the landscape (Tetzlaff et al., 2010). This latter issue could likely be overcome by representing wildlife movements stochastically in MAFIO. Whilst this would probably result in greater stochastic variability across ensemble simulations, it would help increase confidence that the potential impacts of wildlife as a source of FIOs are being represented.

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This last point is relevant when inferring process adequacy from the large spread in non-zero loads simulated at T6 (Figure 8c-d). As there were again no surface flow paths upstream of this site in the simulated hydrological environment (Figure 5b), FIO-agents could only be directly deposited in the stream by livestock in Mid Pasture (R). Given the stochastic treatment of livestock movement and, consequently, direct deposition in MAFIO, it should be expected that different ensemble runs will collectively simulate a range of possible *E. coli* loads dependent on when and where livestock were simulated to directly defecate in the stream (c.f. Abdou et al., 2012). Given the plausibility of direct deposition influencing microbial water quality at T6 when livestock are present in Mid Pasture (R) due to stream accessibility (Figure 1c), observed loads may reflect one particular realisation of how

livestock entered the stream and directly deposited (c.f. Windrum et al., 2007). Consequently, that observations could fall within the spread of simulated loads (Figure 8c-d) likely suggests that MAFIO is adequately representing the process of direct deposition. Thus, it cannot be concluded that the large spread in simulated non-zero loads is indicative of model inadequacy (c.f. Parker and Meretsky, 2004).

A more problematic feature of simulations at T6 is the consistent simulation of zero loads when livestock were absent from Mid Pasture (R) despite observed non-zero loads (Figures 3f and 8c). This could reflect incorrect simulation of zero loads at T8, depending on the extent to which upstream locations influence microbial water quality at T6 (Neill et al., 2018; Vitro et al., 2017). Qualitative similarity between observed *E. coli* dynamics at T6 and T8 when livestock were absent from Mid Pasture (R) lends some support to this possibility (Figures 3c-d and f). However, uncertainty in the exact spatial extent of flow paths simulated by EcH₂O-iso also means that surface connectivity between Mid Pasture (R) and the stream could have plausibly been simulated by some behavioural model runs. Such connectivity could facilitate transfer of FIOs to the stream in the absence of livestock, depending on longevity of survival (Martinez et al., 2013). Consequently, further work to reduce uncertainty in simulated flow paths is necessary to determine whether under-predictions at T6 have a hydrological or microbiological cause. A further possibility could be that a streambed reservoir of *E. coli* from livestock in Mid Pasture (R) exists that can be mobilised by streamflow (McDonald et al., 1982). However, this mechanism would be unlikely to explain under-predictions that occurred during times of recessional and base flows (Figures 3a and 8c; Nagels et al., 2002).

Despite the tendency to over-estimate loads in absolute terms, MAFIO had most skill in capturing observed *E. coli* dynamics at the outlet (Figure 8a-b). Furthermore, stochastic variability in simulations was reduced compared to T6 (Figure 8a-d). Combined, these results suggest that potential process deficiencies impacting upstream sites had less influence here. This likely reflected greater opportunities for FIO-agents to reach the stream via seepage and transport in overland flow generated by areas proximal to the stream in the lower catchment (Figures 1c, 5 and 9). The promising performance at the outlet suggests the model reasonably well-captures these highly-localised mechanisms of FIO transfer as dominant drivers of impaired microbial water quality at this location. This is further supported by relative confidence in EcH₂O-iso simulations of overland flow generation close to the stream and spatial patterns of soil saturation deficit underpinning seepage (Figure 7a and d) and would be consistent with the likely importance of localised sources of FIOs in the wider Tarland Burn (Neill et al., 2018). The over-estimation of loads at the outlet, however, highlights there is scope for improving the degree to which simulations capture the detail in observations. Consequently, several avenues for model

refinement are identified that may help increase correspondence between observed and simulated loads at the outlet and lead to more nuanced simulation of general FIO dynamics.

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Many parameters relating to the loading, die-off, and transport of FIOs are characterised by uncertainty (Cho et al., 2016). Therefore, in addition to a quantitative assessment of how flow path uncertainty in EcH₂O-iso affects MAFIO simulations, a sensitivity analysis and calibration of MAFIO parameters should also be conducted. This may help better-constrain parameters and improve simulations whilst enhancing understanding of how parameter uncertainty is propagated into model outputs (e.g. Beven, 2006). However, limitations in using "fit-to-data" metrics to assess ABM performance likely means that alternative automated calibration procedures will need developing (c.f. Polhill and Salt, 2017). MAFIO has also been applied at relatively coarse spatial (30×30 m) and temporal (1-day timestep) resolutions in this initial application. Whilst this spatial scale is finer than permitted by most processbased FIO models (e.g. Dorner et al., 2006; Whitehead et al., 2016), resolving non-linearities in the fate and transport of FIOs arising from the effects of small-scale heterogenity in the landscape (e.g. microtopography influencing flow paths; Frei et al., 2010) or processes operating at sub-daily timescales (e.g. intra-storm precipitation dynamics; McKergow and Davies-Colley, 2010) may be necessary for more nuanced simulation of FIO dynamics. Finally, alternative methods of allowing MAFIO to simulate the large populations of FIOs found in catchments could be trialled. One possibility is use of "superindividuals" (Scheffer et al., 1995). Like FIO-agents, these are introduced into a simulation for every given number of real individuals. However, this number is then assigned to the super-individual as an attribute that is influenced by controlling processes, which may give a more complete simulation of dynamics that would be observed if all individuals were represented explicitly (Scheffer et al., 1995). Exploring these avenues for model refinement will be a focus of future work.

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- 5.2 What potential does MAFIO have for providing process-based insights into microbial water quality that are relevant for management?
- The preceding discussion highlights the status of MAFIO as a research-level model. However, application to Tulloch Burn still provided insight into how an agent-based approach has significant potential for identifying drivers of microbial water quality at scales relevant for management.

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Management of microbial water quality at the farm scale is always likely to have financial implications for the farmer (Oliver et al., 2007). Consequently, information aiding spatial targeting of cost-effective and efficient mitigation measures is desirable (c.f. Oliver et al., 2018, Vinten et al., 2017). The facility to interrogate the *Domain type* and *Location memory* attributes of exported FIO-agents in MAFIO

permits insights into the transfer mechanisms and source areas contributing FIOs to streams, which may have significant value in this respect. For the Tulloch Burn, for example, it was revealed that whilst overland flow could cause spikes in the flux of exported FIO-agents during events, its limited capacity to transport FIO-agents to the stream in both time and space meant that seepage from areas of degraded soil was always the dominant transfer mechanism (Figures 9-11). Overall, this led to exported FIO-agents being sourced from locations highly proximal to the stream under all conditions (Figures 10-11). As the skill of MAFIO in simulating observed *E. coli* dynamics at the outlet suggests these localised mechanisms are dominant drivers of microbial water quality (Figure 8a-b), an implication is that small-scale interventions (e.g. building bridges between fields separated by the stream or preventing livestock access to stream-proximal locations capable of generating overland flow) could result in significant improvements to microbial water quality without the need for larger-scale and more costly measures (e.g. reducing stocking densities or extensive use of buffer strips; Cuttle et al., 2006).

Quantitative microbial risk assessment (QMRA) can further assist in selection of mitigation measures by providing a basis for assessing how risks presented by faecal pathogens to human health can be reduced through management (Haas et al., 2014; Strachan et al., 2005). Usually, a dose-response model estimates the likelihood of an adverse health effect occurring based on an input dosage of pathogens (Haas et al., 2014). Direct quantification of pathogens in water is not common, however, due to their lower occurrence with respect to FIOs and the costly methods necessary for their enumeration (Geldreich, 1996). Therefore, it may be necessary to estimate exposure based on the prevalence of pathogens in animals responsible for contaminating the medium humans come into contact with (c.f. Strachan et al., 2002). In this regard, MAFIO simulations attributing contributions of exported FIO-agents to different livestock types (Figure 9e) could prove useful. Furthermore, the agent-based model structure could allow direct simulation of pathogenic organisms alongside non-pathogenic FIOs, subject to sufficient data availability to inform parameterisations or rule sets associated with pathogenic FIO-agents (c.f. Hipsey et al., 2008).

A characteristic of MAFIO which also has potential management value is the ability to model processes stochastically. This can enable greater representation of how both natural variability and uncertainty in simulated processes propagate to predictions of microbial water quality, which may be useful for decision making (c.f. Brouwer and De Blois, 2008). Indeed, application of MAFIO to the Tulloch Burn highlighted how simulated *E. coli* loads may demonstrate considerable spread due to variability and uncertainty in how livestock use the landscape and, consequently, directly defecate in streams (c.f. Oliver et al., 2010). However, full realisation of this value would be contingent on the development of

appropriate calibration methods for ABMs which quantify how parameter and structural uncertainty also propagate to model outputs, as discussed earlier.

A final important point relates to the data requirements of MAFIO and its consequent transferability. As previously highlighted, availability of diverse observations corresponding to model outputs will benefit calibration / validation of the model and its associated hydrological environment simulator (c.f. Kelleher et al., 2017; Kuppel et al., 2018b). However, where such observations exist, it is not necessary that model inputs be derived from site-specific data such as those available in this study following intensive monitoring at the Tulloch Burn. For example, nationally-available datasets combined with simple assumptions regarding grazing practices could provide the spatial arrangement of fields within a catchment along with livestock counts that vary in space and time (e.g. Oliver et al., 2010, 2018). Furthermore, use of local datasets (e.g. regarding stream fencing as in Dymond et al., 2016) alongside one-off farm surveys or farmer interviews (Oliver et al., 2007, 2009) could sufficiently characterise the spatial distributions of stream accessibility to livestock and areas of degraded soil. Other necessary catchment characteristics can be derived from widely-available data (e.g. elevation, soil types, etc.) integrated into a geographical information system. Finally, where site-specific data on FIO concentrations in faeces and soil are unavailable, estimates may be derived from literature values (e.g. Dorner et al., 2006; Hipsey et al., 2008; Whitehead et al., 2016). Consequently, it should be possible to apply MAFIO to less intensively-studied catchments. It is also important to emphasise that any hydrological model could be used to provide the hydrological environment for MAFIO as long as its consistency with catchment hydrological functioning can be robustly assessed.

6. Conclusions

This work provided a proof-of-concept application for MAFIO, an agent-based model designed to unravel the spatio-temporal dynamics of sources and transfer mechanisms contributing FIOs to streams at the sub-field scale. Performance in simulating observed *E. coli* dynamics in the Tulloch Burn catchment showed that the model has skill in capturing the transfer of FIOs from livestock to streams via the processes of direct deposition, overland flow and seepage from areas of degraded soil. This assessment was aided by EcH₂O-iso, the hydrological environment simulator for MAFIO, identifying generation of overland flow close to the stream and dominance of groundwater in the catchment with some confidence following multi-criteria calibration to discharge and isotope data. However, uncertainty in the exact spatial extent of overland flow paths simulated by EcH₂O-iso meant it was not always clear whether deficiencies in MAFIO performance reflected a hydrological or microbiological cause. This identified the need for a quantitative assessment of uncertainty propagation from EcH₂O-

iso to MAFIO to be the subject of future work. Nonetheless, under-prediction of observed *E. coli* loads in the upper catchment implied the need to consider "naturalised" or wildlife sources of FIOs in the model, and it was further possible to identify several avenues relating to issues of scale and calibration that could be explored to improve model performance.

Despite the present status of MAFIO as a research-level model, this application revealed how the agent-based structure of the model allowed it to have significant potential for informing management. Interrogation of the attributes of FIO-agents exported from the catchment could reveal insights into source areas, transfer mechanisms and livestock contributing FIOs to the stream, providing information that could inform implementation of efficient, cost-effective mitigation measures. Furthermore, the potential to model processes stochastically in MAFIO allowed the effects of natural variability and uncertainty in processes influencing microbial water quality to be characterised, which may have value in a decision support context. Whilst this proof-of-concept study identified possible refinements that could be made to MAFIO, once addressed, it is likely that the model could have substantial value in underpinning decision support frameworks aimed at mitigating impaired microbial water quality.

Software and data availability

- The source code for MAFIO as used in this work is available via the University of Aberdeen PURE
- 785 repository: https://doi.org/10.20392/66f74663-ece3-4a52-8bed-f0cf52d0831a.
- The source code for EcH₂O-iso is available at: https://bitbucket.org/sylka/ech2o_iso/src/master_2.0/.
- 787 The Tulloch Burn datasets used in this study are available from the lead author on request.

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798	References
799	Abdou, M., Hamill, L., Gilbert, N., 2012. Designing and Building an Agent-Based Model: 141-166 in Heppenstall,
800	A.J., Crooks, A.T., See, L.M., Batty, M. (eds). Agent-Based Models of Geographical Systems. Springer:
801	Dordrecht, Heidelberg, London, New York.
802	Ala-aho, P., Tetzlaff, D., McNamara, J.P., Laudon, H., Soulsby, C., 2017. Using isotopes to constrain water flux
803	and age estimates in snow-influenced catchments using the STARR (Spatially distributed Tracer-Aided
804	Rainfall-Runoff) model. Hydrological Earth Systems Science 21, 5089-5110.
805	Bergfur, J., Demars, B.O.L., Stutter, M.I., Langan, S.J., Friberg, N., 2012. The Tarland Catchment Initiative and
806	Its Effect on Stream Water Quality and Macroinvertebrate Indices. Journal of Environmental Quality 41:
807	314-321.
808	Beven, K., 2006. A manifesto for the equifinality thesis. Journal of Hydrology 320, 18-36.
809	Beven, K., 2012. Rainfall-runoff Modelling: The Primer (Second Edition). John Wiley and Sons: Chichester, UK.
810	Beven, K., 2019. Towards a methodology for testing models as hypotheses in the inexact sciences. Proceedings
811	of the Royal Society A 475: 20180862.
812	Beven, K., Lamb, R., 2017. The uncertainty cascade in model fusion. Geological Society, London, Special
813	Publications 408: 255-266.
814	Bilotta, G.S., Brazier, R.E., Haygarth, P.M., 2007. The impacts of grazing animals on the quality of soils,
815	vegetation and surface waters in intensively managed grasslands. Advances in Agronomy 94: 237-280.
816	Birkel, C., Soulsby, C., 2015. Advancing tracer-aided rainfall-runoff modelling: a review of progress, problems
817	and unrealised potential. Hydrological Processes 29: 5227-5240.
818	Brouwer, R., De Blois, C., 2008. Integrated modelling of risk and uncertainty underlying the cost and effectiveness
819	of water quality measures. Environmental Modelling and Software 23: 922-937.
820	Brown, D.G., Page, S., Riolo, R., Zellner, M., Rand, W., 2005. Path dependence and the validation of agent-based
821	spatial models of land use. International Journal of Geographical Information Science 19: 153-174.
822	Cho, K.H., Pachepsky, Y.A., Oliver, D.M., Muirhead, R.W., Park, Y., Quilliam, R.S., Shelton, D.R., 2016.
823	Modelling fate and transport of fecally-derived microorganisms at the watershed scale: State of the
824	science and future opportunities. Water Research 100: 38-56.

825	Cuttle, S.P., Macleod, C.J.A., Chadwick, D.R., Scholefield, D., Haygarth, P.M., Newell-Price, P., Harris, D.
826	Shepherd, M.A., Chambers, B.J., Humphrey, R., 2006. An inventory of methods to control diffuse water
827	pollution from agriculture (DWPA): User Manual. Accessed July 2019 from
828	http://randd.defra.gov.uk/Document.aspx?Document=es0203_4145_FRA.pdf
829	Dee, D.P., Uppala, S.M., Simmons, A.J., Berrisford, P., Poli, P., Kobayashi, S., Andrae, U., Balmaseda, M.A.
830	Balsamo, G., Bauer, P., Bechtold, P., Beljaars, A.C.M., van de Berg, L., Bidlot, J., Bormann, N., Delsol
831	C., Dragani, R., Fuentes, M., Geer, A.J., Haimberger, L., Healy, S.B., Hersbach, H., Holm, E.V., Isaksen,
832	L., Kållberg, P., Kohler, M., Matricardi, M., McNally, A.P., Monge-Sanz, B.M., Morcrette, JJ., Park
833	BK., Peubey, C., de Rosnay, P., Tavolato, C., Thepaut, JN., Vitart, F., 2011. The ERA-Interim
834	reanalysis: configuration and performance of the data assimilation system. Quarterly Journal of the Royal
835	Meteorological Society 137: 553-597.
836	Dorner, S.M., Anderson, W.B., Slawson, R.M., Kouwen, N., Huck, P.M., 2006. Hydrologic modeling of pathogen
837	fate and transport. Environmental Science and Technology 40, 4746-4753.
838	Douinot, A., Tetzlaff, D., Maneta, M., Kuppel, S., Schulte-Bisping, H., Soulsby, C., 2019. Ecohydrological
839	modelling with EcH2O-iso to quantify forest and grassland effects of water partitioning and flux ages.
840	Hydrological Processes 33: 2174-2191.
841	Dunn, S.M., Johnston, L., Taylor, C., Watson, H., Cook, Y., Langan, S.J., 2013. Capability and limitations of a
842	simple grid-based model for simulating land use influences on stream nitrate concentrations. Journal of
843	Hydrology 507: 110-123.
844	Dymond, J.R., Serezat, D., Ausseil, A.G.E., Muirhead, R.W., 2016. Mapping of Escherichia coli sources
845	connected to waterways in the Ruamahanga catchment, New Zealand. Environmental Science and
846	Technology 50(4), 1897-1905.
847	Frei, S., Lischeid, G., Fleckenstein, J.H., 2010. Effects of micro-topography on surface-subsurface exchange and
848	runoff generation in a virtual riparian wetland – A modeling study. Advances in Water Resources 13(11)
849	1388–1401.
850	Frick, C., Vierheilig, J., Linke, R., Savio, D., Zornig, H., Antensteiner, R., Baumgartner, C., Bucher, C., Blaschke
851	A.P., Derx, J., Kirschner, A.K.T., Ryzinska-Paier, G., Mayer, R., Seidl, D., Nadiotis-Tsaka, T., Sommer
852	R., Farnleitner, A.H., 2018. Poikilothermic Animals as a Previously Unrecognized Source of Fecal
853	Indicator Bacteria in a Backwater Ecosystem of a Large River. Applied and Environmental Microbiology
854	84: e00715-18.

856 Goody, R.M., Yung, Y.L., 1995. Atmospheric Radiation: Theoretical Basis. Oxford University Press: Oxford, 857 UK. 858 Greene, S., Johnes, P.J., Bloomfield, J.P., Reaney, S.M., Lawley, R., Elkhatib, Y., Freer, J., Odoni, N., Macleod, 859 C.J.A., Percy, B., 2015. A geospatial framework to support integrated biogeochemical modelling in the 860 United Kingdom. Environmental Modelling and Software 68: 219-232. 861 Guber, A.K., Fry, J., Ives, R.L., Rose, J.B., 2015. Escherichia coli Survival in, and release from, White-Tailed Deer feces. Applied and Environmental Microbiology 81, 1168-1176. 862 863 Gupta, H.V., Kling, H., Yilmaz, K.K., Martinez, G.F., 2009. Decomposition of the mean squared error and NSE 864 performance criteria: Implications for improving hydrological modelling. Journal of Hydrology 377: 80-865 91. 866 Hannaford, J., Muchan, K., Lewis, M., Clemas, S., 2018. Hydrological summary for the United Kingdom: 867 December 2017. Wallingford, UK: NERC/Centre for Ecology & Hydrology. Accessed June 2019 from 868 http://nora.nerc.ac.uk/id/eprint/518984/ 869 Haas, C.N., Rose, J.B., Gerba, C.P., 2014. Quantitative microbial risk assessment (Second Edition). Wiley: 870 Hoboken, New Jersey. 871 Haydon, S., Deletic, A., 2006. Development of a coupled pathogen-hydrologic catchment model. Journal of 872 Hydrology 328: 467-480. 873 Hipsey, M.R., Antenucci, J.P., Brookes, J.D., 2008. A generic, process-based model of microbial pollution in 874 aquatic systems. Water Resources Research 44, W07408. 875 Hrachowitz, M., Soulsby, C., Tetzlaff, D., Speed, M., 2010. Catchment transit times and landscape controls— 876 does scale matter? Hydrological Processes 24: 117-125. 877 Jang, J., Hur, H.-G., Sadowsky, M.J., Byappanahalli, M.N., Yan, T., Ishii, S., 2017. Environmental Escherichia 878 coli: ecology and public health implications—a review. Journal of Applied Microbiology 123: 570-581. 879 Kay, D., Crowther, J., Kay, C., Mcdonald, A.T., Ferguson, C., Stapleton, C.M., Wyer, M.D., 2012. Effectiveness 880 of best management practices for attenuating the transport of livestock-derived pathogens within 881 catchments: 195-255 in Dufour, A., Bartram, J., Bos, R., Gannon, V. (eds.). Animal waste, water quality 882 and human health. IWA Publishing: London.

883 Kay, D., Crowther, J., Stapleton, C.M., Wyer, M.D., Fewtrell, L., Anthony, S., Bradford, M., Edwards, A., 884 Francis, C.A., Hopkins, M., Kay, C., McDonald, A.T., Watkins, J., Wilkinson, J., 2008. Faecal indicator 885 organism concentrations and catchment export coefficients in the UK. Water Research 42: 2649-2661. 886 Kelleher, C., McGlynn, B., Wagener, T., 2017. Characterising and reducing equifinality by constraining a 887 distributed catchment model with regional signatures., local observations and process understanding. 888 Hydrology and Earth Systems Science 21: 3325-3352. 889 Krause, P., Boyle, D.P., Bäse, F., 2005. Comparison of different efficiency criteria for hydrological model 890 assessment. Advances in Geosciences 5: 89-97. 891 Kuppel, S., Tetzlaff, D., Maneta, M.P., Soulsby, C., 2018a. EcH2O-iso 1.0: water isotopes and age tracking in a 892 process-based, distributed ecohydrological model. Geoscientific Model Development 11: 3045-3069. 893 Kuppel, S., Tetzlaff, D., Maneta, M.P., Soulsby, C., 2018b. What can we learn from multi-criteria calibration of 894 a process-based ecohydrological model? Environmental Modelling and Software 101: 301-316. 895 Langan, S.J., Wade, A.J., Smart, R.P., Edwards, A.C., Soulsby, C., Billett, M.F., Jarvie, H.P., Cresser, M.S., 896 Owen, R., Ferrier, R.C., 1997. The prediction and management of water quality in a relatively unpolluted 897 major Scottish catchment: current issues and experimental approaches. Science of the Total Environment 898 194-195: 419-435. 899 Legates, D.R., McCabe, G.J., 1999. Evaluating the use of "goodness-of-fit" measures in hydrologic and 900 hydroclimatic model validation. Water Resources Research 35: 233-241. Martinez, G., Pachepsky, Y.A., Shelton, D.R., Whelan, G., Zepp, R., Molina, M., Panhorst, K., 2013. Using the 901 902 Q₁₀ model to simulate E. coli survival in cowpats on grazing lands. Environmental International 54: 1-903 10. 904 McDonald, A., Kay, D., Jenkins, A., 1982. Generation of fecal and total coliform surges by stream flow 905 manipulation in the absence of normal hydrometeorological stimuli. Applied and Environmental 906 Microbiology 44: 292-300. 907 McKergow, L.A., Davies-Colley, R.J., 2010. Stormflow dynamics and loads of Escherichia coli in a large mixed 908 land use catchment. Hydrological Processes 24: 276-289. 909 Morris, M.D., 1991. Factorial sampling plans for preliminary computational experiments. Technometrics 33: 161-910 174.

911	Moss, S., Edmonds, B., 2005. Sociology and Simulation: Statistical and Qualitative Cross-Validation. American
912	Journal of Sociology 110: 1095-1131.
913	Muirhead, R.W., 2009. Soil and faecal material reservoirs of Escherichia coli in a grazed pasture, New Zealand
914	Journal of Agricultural Research 52: 1-8.
915	Muirhead, R.W., Elliot, A.H., Monaghan, R.M., 2011. A model framework to assess the effect of dairy farms and
916	wild fowl on microbial water quality during base-flow. Water Research 45: 2863-2874.
917	Nagels, J.W., Davies-Colley, R.J., Donnison, A.M., Muirhead, R.W., 2002. Faecal contamination over flood
918	events in a pastoral agricultural stream in New Zealand. Water Science and Technology 45: 45-52.
919	Neill, A.J., Tetzlaff, D., Strachan, N.J.C., Soulsby, C., 2019. To what extent does hydrological connectivity
920	control dynamics of faecal indicator organisms in streams? Initial hypothesis testing using a tracer-aided
921	model. Journal of Hydrology 570: 423-435.
922	Neill, A.J., Tetzlaff, D., Strachan, N.J.C., Hough, R.L., Avery, L.M., Watson, H., Soulsby, C., 2018. Using spatial-
923	stream-network models and long-term data to understand and predict dynamics of faecal contamination
924	in a mixed land-use catchment. Science of the Total Environment 612: 840-852.
925	Neill, A.J., Tetzlaff, D., Strachan, N.J.C., Hough, R.L., Avery, L.M., Kuppel, S., Maneta, M.P., Soulsby, C., in
926	review. An agent-based model that simulates the spatio-temporal dynamics of sources and transfer
927	mechanisms contributing faecal indicator organisms to streams. Part 1: Background and model
928	description. Journal of Environmental Management.
929	O'Sullivan, D., Millington, J., Perry, G. & Wainwright, J., 2012. Agent-based models – because they're worth it?:
930	109-123 in Heppenstall, A.J., Crooks, A.T., See, L.M., Batty, M. (eds). Agent-Based Models of
931	Geographical Systems. Springer: Dordrecht, Heidelberg, London, New York.
932	Oliver, D.M., Heathwaite, A.L., Hodgson, C.J., Chadwick, D.R., 2007. Mitigation and current management
933	attempts to limit pathogen survival and movement within farmed grassland. Advances in Agronomy 93:
934	95-152.
935	Oliver, D.M., Bartie, P.J., Heathwaite, A.L., Reaney, S.M., Parnell, J.A.Q., Quilliam, R.S., 2018. A catchment-
936	scale model to predict spatial and temporal burden of E. coli on pasture from grazing livestock. Science
937	of the Total Environment 616-617: 678-687.

938 939 940	Oliver, D.M., Fish, R.D., Hodgson, C.J., Heathwaite, A.L., Chadwick, D.R., Winter, M., 2009. A cross-disciplinary toolkit to assess the risk of faecal indicator loss from grassland farm systems to surface waters. Agriculture, Ecosystems and Environment 129: 401-412.
941 942 943	Oliver, D.M., Page, T., Hodgson, C.J., Heathwaite, A.L., Chadwick, D.R., Fish, R.D., Winter, M., 2010. Development and testing of a risk indexing framework to determine field-scale critical source areas of faecal bacteria on grassland. Environmental Modelling and Software 25: 503-512.
944 945 946 947	Oliver, D.M., Porter, K.D.H., Pachepsky, Y.A., Muirhead, R.W., Reaney, S.M., Coeffey, R., Kay, D., Milledge, D.G., Hong, E., Anthony, S.G., Page, T., Bloodworth, J.W., Mellander, P., Carbonneau, P., McGrane, S.J., Quilliam, R.S., 2016. Predicting microbial water quality with models: Over-arching questions for managing risk in agricultural catchments. Science of the Total Environment 544: 39-47.
948 949	Parker, D.C., Meretsky, V., 2004. Measuring pattern outcomes in an agent-based model of edge-effect externalities using spatial metrics. Agriculture, Ecosystems and Environment 101: 233-250.
950 951 952	Parry, H.R., Bithell, M., 2012. Large scale agent-based modelling: A review and guidelines for model scaling: 271-308 in Heppenstall, A.J., Crooks, A.T., See, L.M., Batty, M. (eds). Agent-Based Models of Geographical Systems. Springer: Dordrecht, Heidelberg, London, New York.
953 954 955	Polhill, J.G., Sutherland, L., Gotts, N.M., 2010. Using qualitative evidence to enhance and agent-based modelling system for studying land use change. Journal of Artificial Societies and Social Simulation 13 (2) 10. http://jasss.soc.surrey.ac.uk/13/2/10.html
956 957 958	Polhill, J.G., Salt, D., 2017. The importance of ontological structure: why validation by 'fit-to-data' is insufficient: 141-172 in Edmonds, B., Meyer, R. (eds.) Simulating Social Complexity: A Handbook (Second Edition). Springer: Dordrecht, Heidelberg, London, New York.
959 960 961	Porter, K.D.H., Reaney, S.M., Quilliam, R.S., Burgess, C., Oliver, D.M., 2017. Predicting diffuse microbial pollution risk across catchments: The performance of SCIMAP and recommendations for future development. Science of the Total Environment 609: 456-465.
962 963 964	Reaney, S.M., 2008. The use of agent based modelling techniques in hydrology: determining the spatial and temporal origin of channel flow in semi-arid catchments. Earth Surface Processes and Landforms 33, 317-327.
965 966	Refsgaard, J.C., van der Sluijs, J.P., Højberg, A.L., Vanrolleghem, P.A., 2007. Uncertainty in the environmental modelling process – A framework and guidance. Environmental Modelling and Software 22: 1543-1556.

967 Rode, M., Arhonditsis, G., Balin, D., Kebede, T., Krysanova, V., van Griensven, A., van der Zee, S.E.A.T.M., 968 2010. New challenges in integrated water quality modelling. Hydrological Processes 24: 3447-3461. 969 Sadeghi, A., Arnold, J., 2002. A SWAT/Microbial Sub-Model for Predicting Pathogen Loadings in Surface and 970 Groundwater at Watershed and Basin Scales. In Total Maximum Daily Load (TMDL): Environmental 971 Regulations, Proceedings of 2002 Conference (p. 56). American Society of Agricultural and Biological 972 Engineers. 973 Schaefli, B., Gupta, H.V., 2007. Do Nash values have value? Hydrological Processes 21: 2075-2080. 974 Scheffer, M., Baveco, J.M., DeAndelis, D.L., Rose, K.A., van Nes, E.H., 1995. Super-individuals a simple 975 solution for modelling large populations on an individual basis. Ecological Modelling 80: 161-170. 976 Soheir, H., Farges, J-L., Piet-Lahanier, H., 2014. Improvement of the representativity of the Morris Method for 977 air-launch-to-orbit separation. IFAC Proceedings Volumes 47(3): 7954-7959. 978 Soil Survey of Scotland Staff, 2014. Digitized soil map of Scotland, scale 1:25 000. James Hutton Institute. 979 Sprenger, M., Stumpp, C., Weiler, M., Aeschbach, W., Allen, S.T., Benettin, P., Dubbert, M., Hartmann, A., 980 Hrachowitz, M., Kirchner, J.W., McDonnell, J.J., Orlowski, N., Penna, D., Pfahl, S., Rinderer, M., 981 Rodriguez, N., Schmidt, M., Werner, C., 2019. The demographics of water: A review of water ages in 982 the critical zone. Reviews of Geophysics 57: 800-834. 983 Strachan, N.J.C., Dunn, G.M., Ogden, I.D., 2002. Quantitative risk assessment of human infection from 984 Escherichia coli O157 associated with recreational use of animal pasture. International Journal of Food 985 Microbiology 75: 39-51. 986 Strachan, N.J.C., Doyle, M.P., Kasuga, F., Rotariu, O., Ogden, I.D., 2005. Dose response modelling of 987 Escherichia coli O157 incorporating data from foodborne and environmental outbreaks. International 988 Journal of Food Microbiology 103: 35-47. 989 Tetzlaff, D., Soulsby, C., Birkel, C., 2010. Hydrological connectivity and microbiological fluxes in montane 990 catchments: the role of seasonality and climatic variability. Hydrological Processes 24: 1231-1235. 991 Vaché, K.B., McDonnell, J.J., 2006. A process-based rejectionist framework for evaluating catchment runoff 992 model structure. Water Resources Research 42, W02409. 993 Vinten, A., Sample, J., Ibiyemi, A., Abdul-Salam, Y., Stutter, M., 2017. A tool for cost-effectiveness analysis of

field scale sediment-bound phosphorus mitigation measures and application to analysis of spatial and

995	temporal targeting in the Lunan Water catchment, Scotland. Science of the Total Environment 586: 631-
996	641.
997	Vitro, K.A., BenDor, T.K., Jordanova, T.V., Miles, B., 2017. A geospatial analysis of land use and stormwater
998	management on fecal coliform contamination in North Carolina streams. Science of the Total
999	Environment 603-604: 709-727.
1000	Wellen, C., Kamran-Disfani, A.R., Arhonditsis, G.B., 2015. Evaluation of the current state of distributed
1001	watershed nutrient water quality modeling. Environmental Science and Technology 49: 3278-3290.
1002	Whitehead, P.G., Leckie, H., Rankinen, K., Butterfield, D., Futter, M.N., Bussi, G., 2016. An INCA model for
1003	pathogens in rivers and catchments: Model structure, sensitivity analysis and application to the River
1004	Thames catchment, UK. Science of the Total Environment 572: 1601-1610.
1005	Willmott, C.J., Matsuura, K., 2005. Advantages of the mean absolute error (MAE) over the root mean square error
1006	(RMSE) in assessing average model performance. Climate Research 30: 79-82.
1007	Windrum, P., Fagiolo, G., Moneta, A., 2007. Empirical validation of agent-based models: Alternatives and
1008	perspectives. Journal of Artificial Societies and Social Simulation 10 (2) 8.
1009	http://jasss.soc.surrey.ac.uk/10/2/8.html
1010	

1011 Figures

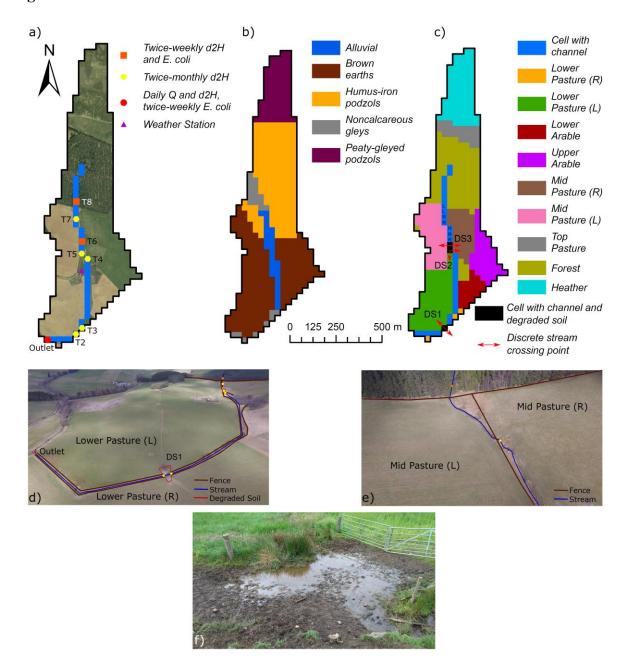


Figure 1: The Tulloch Burn catchment, with maps showing a) An overview of the catchment and monitoring locations; b) Soil types based on Soil Survey of Scotland Staff (2014); c) Designations of land parcels and cells containing a channel (potentially with an area of degraded soil) as provided to MAFIO. In the latter, the appearance of "L" or "R" in cells containing a channel denote the permanent direct accessibility of the stream to livestock in Mid Pastures (L) and (R), respectively, whilst DS1-3 are the designations given to the three areas of degraded soil associated with discrete stream crossing points. The stream in (a) and all data in (b) and (c) are presented on the 30×30 m grid utilised by EcH₂O-iso and MAFIO. Also provided are drone-based aerial images of the catchment showing examples of d) Fencing-off of the stream from adjacent fields and areas of soil degradation; e) Sections of the stream

directly accessible to livestock. For context, selected field and degraded soil designations, and approximate sampling locations are provided in the aerial images using the same symbols as in a). An example of soil degradation (DS3) in the catchment is shown in (f).

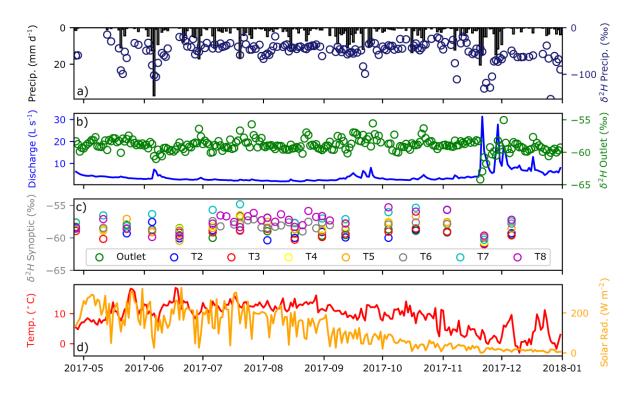


Figure 2: For the full study period, observed timeseries of a) Precipitation and its associated isotopic composition; b) Daily average discharge at the catchment outlet and its associated isotopic composition; c) Isotopic compositions of streamwater at synoptic sampling sites; d) Daily average temperature and solar radiation.

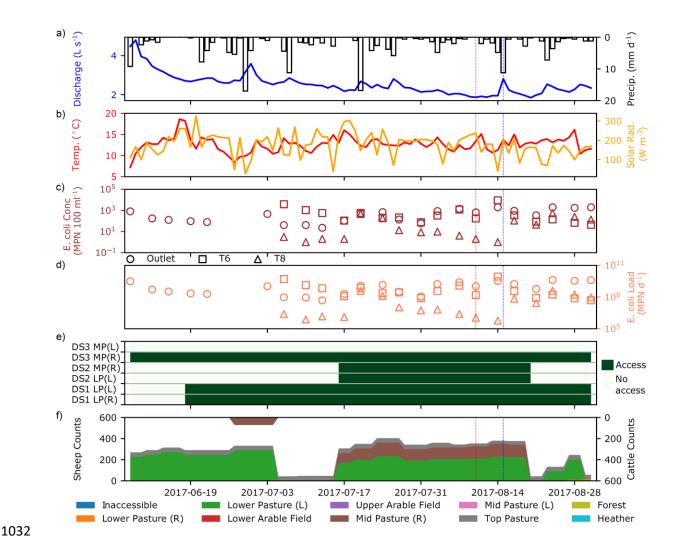


Figure 3: For the microbial observation period, timeseries of observed a) Precipitation and daily average discharge at the catchment outlet; b) Daily average temperature and solar radiation; c) Concentrations of *E. coli* (plotted on a log scale); d) *E. coli* loads (plotted on a log scale); e) Accessibility of the stream to livestock at the discrete crossing points (DS1-3 in Figure 1c); f) Sheep and cattle counts. The red and blue dashed lines denote the example dry and wet days, respectively. In (c) and (d), MPN = most-probable-number. In (e), the abbreviations LP and MP refer to Lower Pasture and Mid Pasture, respectively.

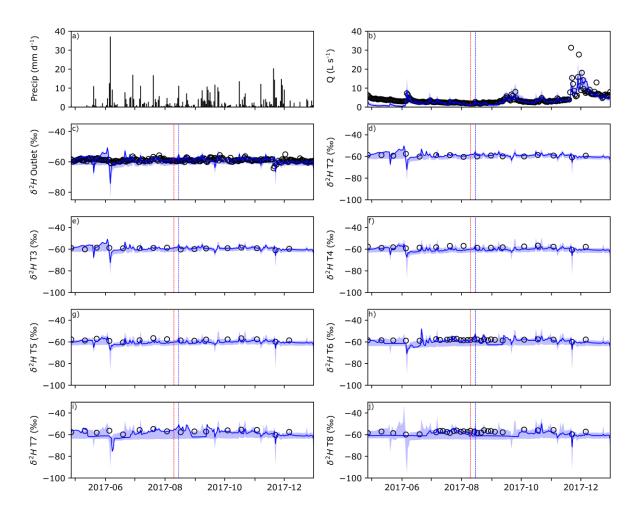


Figure 4: Timeseries of a) Precipitation; b) Observed and modelled discharge at the catchment outlet; c-j) Observed and modelled isotopes at the catchment outlet and synoptic sampling sites. Shaded areas show the 90% spread of simulations from the behavioural ensemble, whilst the solid blue line shows the "best" simulation. The red and blue dashed lines denote the example dry and wet days, respectively. Note the different y-scales between isotope data for the outlet (c) and for the synoptic sampling sites (d-j), necessary to show the greater spread of the simulations for the latter.

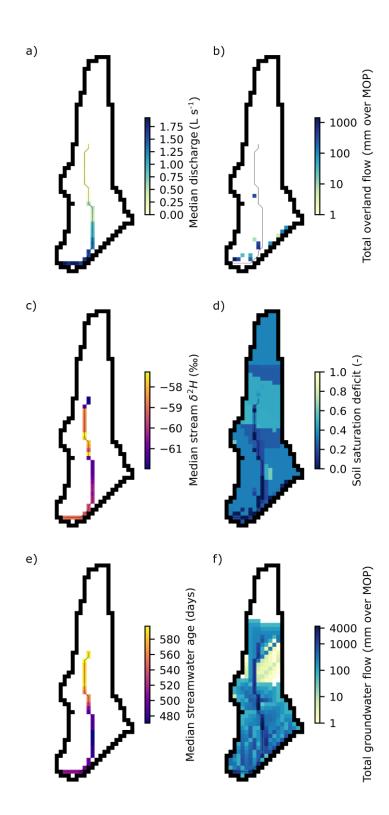


Figure 5: Maps showing for the microbial observation period a) Median stream discharge; b) Total overland flow; c) Median stream δ^2H ; d) Median soil saturation deficit; e) Median streamwater age; f) Total groundwater flow, based on the "best" ensemble run of EcH₂O-iso. Overland (b) and groundwater (f) flows are plotted on log scales for clarity, with areas of white denoting fluxes of 0. MOP = Microbial observation period.

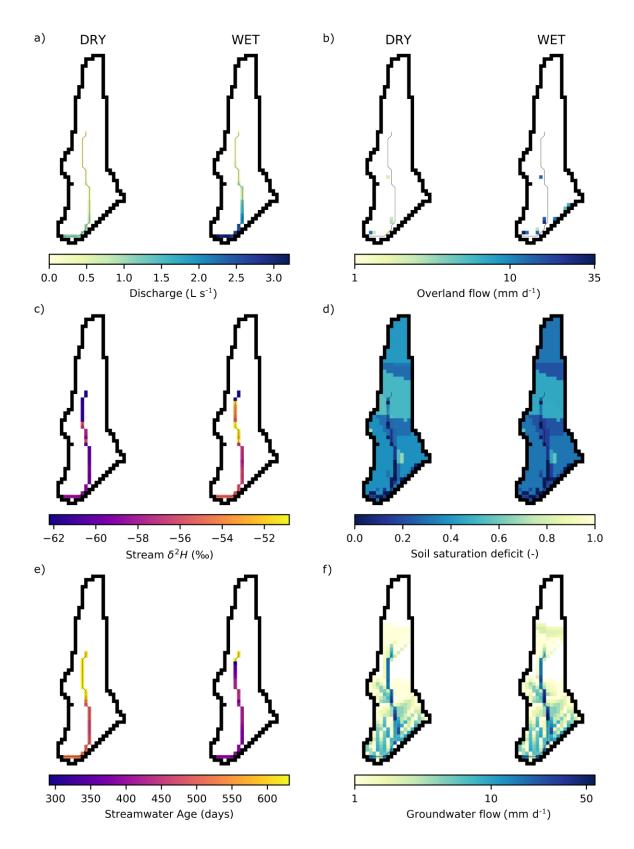


Figure 6: For the example dry and wet days, maps showing a) Discharge; b) Overland flow; c) Streamwater $\delta^2 H$; d) Soil saturation deficit; e) Streamwater age; f) Groundwater flow, based on the "best" ensemble run of EcH₂O-iso. Overland (b) and groundwater (f) flows are plotted on log scales for clarity, with areas of white denoting fluxes of 0.

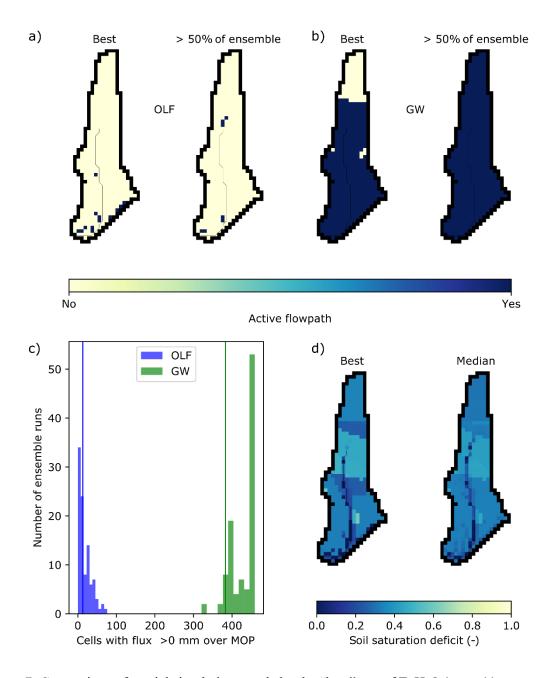


Figure 7: Comparison of spatial simulations made by the "best" run of EcH₂O-iso and by an ensemble of 100 behavioural runs: a) Spatial extent of cells with total overland flow (OLF) fluxes greater than 0 mm over the microbial observation period (i.e. "Active flowpath") simulated by the "best" run and >50% of the ensemble; b) As (a) but for groundwater (GW); c) Histogram quantifying the number of ensemble runs in which different numbers of cells were simulated to have total OLF or GW fluxes greater than 0 mm over the microbial observation period (solid lines denote the "best" run); d) Spatial patterns of median soil saturation deficit over the microbial observation period simulated by the "best" run and of the median of the median deficits simulated by the behavioural ensemble. MOP = Microbial observation period.

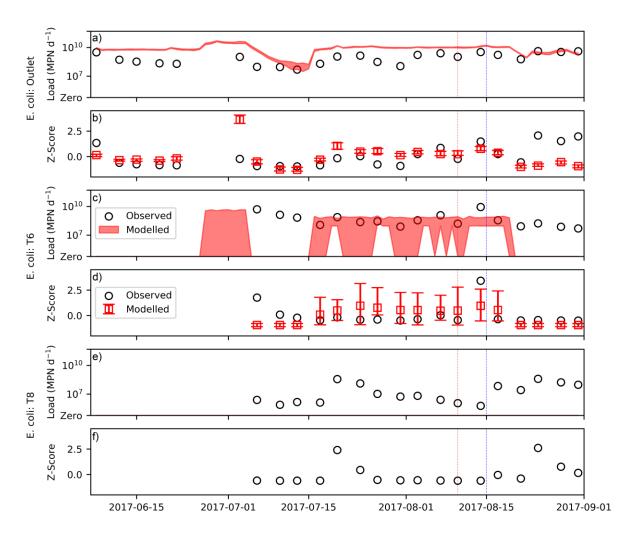


Figure 8: Comparison of observed and simulated E. coli loads and Z-scores for a-b) The catchment outlet; c-d) T6; e-f) T8. In the latter, only observed Z-scores are given due to all ensemble runs of MAFIO simulating fluxes of zero FIO-agents for all timesteps. For modelled data, the square marker represents the median Z-score across the 30 ensemble runs of MAFIO, whilst the error bars/shaded areas denote the range of Z-scores/simulated loads. The red and blue dashed lines denote the example dry and wet days, respectively. Loads are plotted on a log scale.

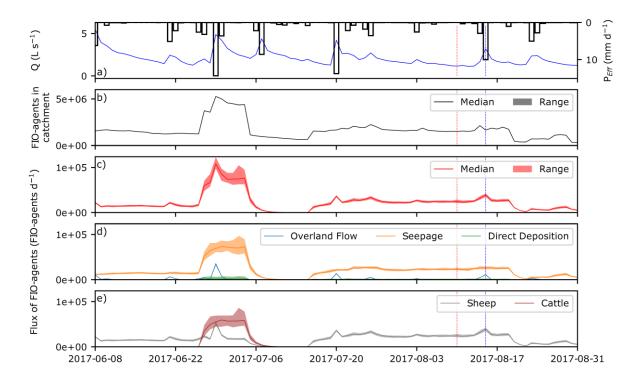


Figure 9: Timeseries of a) Effective precipitation and discharge simulated by the "best" ensemble run of EcH₂O-iso; and b) FIO-agents stored in the catchment at the end of each timestep; c) Flux of FIO-agents exported from the catchment; d) Mechanisms by which exported FIO-agents reached the stream; e) Contributions of exported FIO-agents from sheep and cattle, based on the 30 ensemble runs of MAFIO. The red and blue dashed lines denote the example dry and wet days, respectively. All scales are linear.

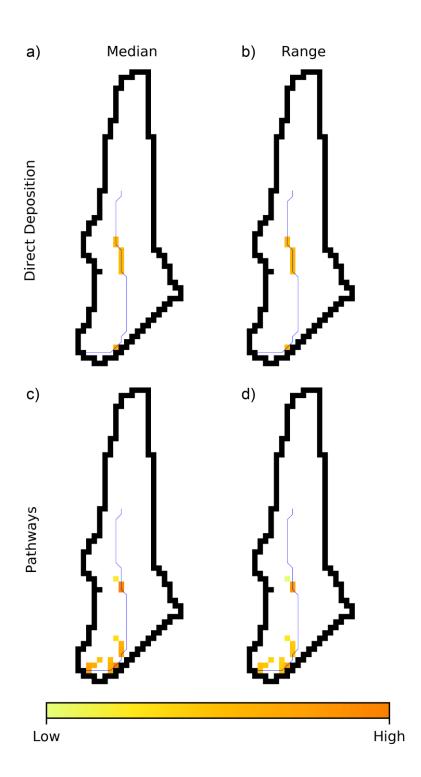


Figure 10: For the whole microbial observation period and based on the 30 ensemble runs of MAFIO, maps showing the median number or range in numbers of exported FIO-agents a-b) Directly deposited in the stream for each cell containing a channel or c-d) That passed through each grid cell in the course of being transported to the stream in overland flow or seepage. The scale reflects the log₁₀-transformed median number/range in numbers and is common to all maps. Areas of white denote medians/ranges of 0.

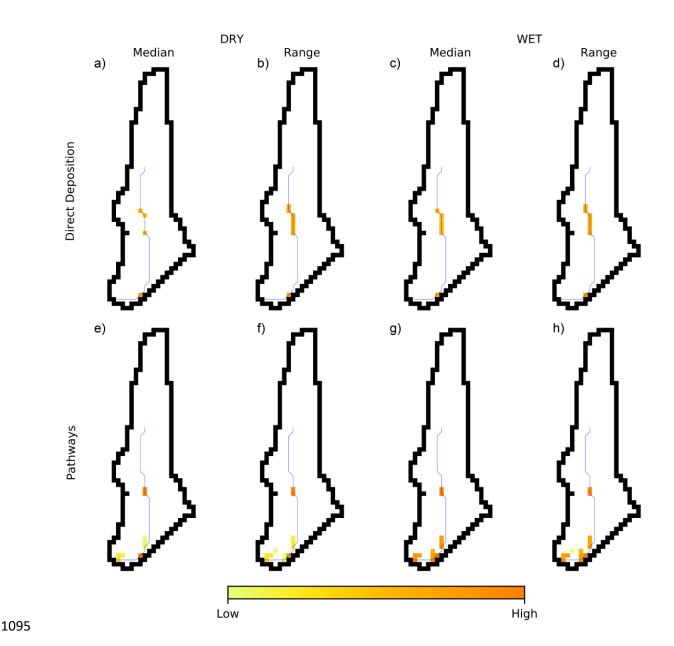


Figure 11: For the exemplar dry day and based on the 30 ensemble runs of MAFIO, maps showing the median number or range in numbers of exported FIO-agents a-b) Directly deposited in the stream for each cell containing a channel or e-f) That passed through each grid cell in the course of being transported to the stream in overland flow or seepage. Equivalent maps for the wet day are shown in (c-d) and (g-h), respectively. The scale reflects the log₁₀-transformed median number/range in numbers and is common to all maps. Areas of white denote medians/ranges of 0.

1103 Tables

Table 1: The number of observations for each calibration target and the weighting for each target in the multi-criteria calibration.

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Dataset	Number of observations	Calibration weighting
Discharge: Outlet	249	0.396
Isotopes: Outlet	242	0.385
Isotopes: T2	16	0.025
Isotopes: T3	16	0.025
Isotopes: T4	16	0.025
Isotopes: T5	16	0.025
Isotopes: T6	28	0.045
Isotopes: T7	16	0.025
Isotopes: T8	29	0.046
Total	628	1.0

Table 2: The mean absolute error (MAE) for simulations of discharge and isotope data by the "best" run of EcH₂O-iso (Best MAE), and summary statistics for simulations by the 100 behavioural runs.

Dataset	Best MAE	Mean MAE	Min - Max MAE
Discharge: Outlet	1.37 L s ⁻¹	1.56 L s ⁻¹	1.24 - 2.04 L s ⁻¹
Isotopes: Outlet	1.41‰	1.99‰	1.12 - 3.68‰
Isotopes: T2	1.30‰	2.06‰	1.02 - 3.82‰
Isotopes: T3	1.14‰	2.13‰	0.95 - 3.82‰
Isotopes: T4	2.73‰	3.57‰	0.83 - 6.20‰
Isotopes: T5	2.69‰	3.56‰	1.00 - 6.25‰
Isotopes: T6	2.00‰	3.36‰	0.86 - 6.51‰
Isotopes: T7	3.02‰	4.64‰	1.55 - 8.21‰
Isotopes: T8	2.52‰	4.23‰	1.30 - 8.08‰

Supplementary Material

Appendix A: Parameterisation of EcH₂O-iso

The sampling and calibrated ranges of parameters found to be sensitive in EcH₂O-iso are given in Table S1, whilst fixed values of insensitive parameters are detailed in Table S2.

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Table S1: The sampling and 90%-spread calibrated ranges of soil, vegetation and channel parameters identified as sensitive in the application of EcH_2O -iso to Tulloch Burn. Additional information on parameter definitions can be found at: https://ech2o-iso.readthedocs.io/en/latest/Setup.html

Sampling range

1122

Parameter

	[90% spread of c	calibrated range]						
Soil	Brown earths	Humus-iron podzol	Noncalcareous gley	Peaty-gleyed podzol	Alluvial			
Total soil depth (m)	1-4 [1.1-3.9]	1-4 [1.1-3.8]	1-4 [1.2-3.8]	1-4 [1.2-3.8]	1-4 [1.3-3.9]			
Depth of 1st hydrological	0.05-0.25	0.05-0.25	0.05-0.25	0.05-0.25	0.05-0.25			
layer (m)	[0.066-0.25]	[0.071-0.24]	[0.059-0.24]	[0.069-0.24]	[0.068-0.24]			
Depth of 2 nd hydrological	0.05-0.5	0.05-0.5	0.05-0.5	0.05-0.5	0.05-0.5			
layer (m)	[0.082-0.49]	[0.073-0.48]	[0.088-0.49]	[0.067-0.45]	[0.096-0.45]			
Porosity (m ³ m ⁻³)	0.4-0.6	0.4-0.6	0.4-0.6	0.4-0.8	0.4-0.6			
	[0.41-0.59]	[0.42-0.60]	[0.41-0.58]	[0.43-0.76]	[0.41-0.58]			
Porosity exponential decay	1-20	1-20	1-20	1-20	1-20			
constant (m ⁻¹)	[1.8-18.7]	[2.1-18.4]	[1.7-18.0]	[1.6-19.0]	[1.9-19.5]			
Saturated horizontal hydraulic	1×10 ⁻⁵ -0.01							
conductivity (ms ⁻¹)	[7.05×10 ⁻⁵ -	[9.79×10 ⁻⁵ -	[3.72×10 ⁻⁴ -	[1.53×10 ⁻⁴ -	[2.84×10 ⁻⁴ -			
	0.0022]	0.0073]	0.0087]	0.0093]	0.0095]			
Hydraulic conductivity	1-20	1-20	1-20	1-20	1-20			
exponential decay constant (m ⁻¹)	[1.5-18.4]	[1.6-18.6]	[2.2-19.3]	[1.4-19.0]	[1.6-19.2]			
Anisotropy (-)	1×10 ⁻³ -0.6							
	[0.035-0.55]	[0.027-0.58]	[0.029-0.57]	[0.043-0.56]	[0.022-0.57]			

Brooks-Corey lambda (-)	3-15	3-15	3-15	3-15	3-15
	[3.4-13.6]	[3.4-13.9]	[3.6-14.6]	[3.6-14.2]	[3.5-13.9]
Air-entry pressure head (m)	0.05-0.8	0.05-0.8	0.05-0.8	0.05-0.8	0.05-0.8
	[0.11-0.76]	[0.12-0.76]	[0.15-0.75]	[0.11-0.74]	[0.080-0.75]
Tension threshold for mobile	1-100	1-100	1-100	1-100	1-100
water (m)	[11.3-96.4]	[12.1-94.8]	[5.1-94.0]	[10.4-92.8]	[6.9-95.3]
Vegetation	Grasses	Conifers	Heather		
Root profile exponential	1-20	1-20	1-20		
decay constant (m ⁻¹)	[2.0-18.9]	[1.8-18.9]	[1.9-18.5]		
Maximum stomatal	3×10 ⁻³ -0.05	3×10 ⁻³ -0.05	3×10 ⁻³ -0.05		
conductance (ms ⁻¹)	[0.0039-0.045]	[0.0068-0.047]	[0.0048-0.046]		
Soil water potential below	2-6	2-6	2-6		
which there is complete	[2.2-5.9]	[2.3-5.7]	[2.6-5.8]		
stomatal closure (-MPa)					
Soil water potential above	0.1-1	0.1-1	0.1-1		
which there is no soil water	[0.18-0.96]	[0.12-0.94]	[0.12-0.93]		
limitation to stomatal	[0.10 0.50]	[0.12 0.5 .]	[0.12 0.55]		
conductance (-MPa)					
Stomatal sensitivity to light	200-500	200-500	200-500		
(-)	[228.9-482.6]	[223.6-492.0]	[208.5-488.8]		
Stomatal sensitivity to vapour	1×10 ⁻³ -3×10 ⁻³	1×10 ⁻³ -3×10 ⁻³	1×10 ⁻³ -3×10 ⁻³		
pressure deficit (-)	[1.2×10 ⁻³ -	[1.1×10 ⁻³ -	$[1.1 \times 10^{-3}]$		
	2.9×10^{-3}]	2.9×10^{-3}]	2.9×10^{-3}]		
Minimum temperature of	-5-5	-5-5	-5-5		
comfort (°C)	[-4.1-4.4]	[-4.5-4.7]	[-4.2-4.6]		
Optimal temperature (°C)	6-24	6-24	6-24		
	[7.6-23.3]	[6.9-22.8]	[6.9-23.2]		
Maximum temperature of	25-40	25-40	25-40		
comfort (°C)	[26.0-38.9]	[26.0-39.9]	[25.9-38.8]		
Maximum interception	5×10 ⁻⁵ -5×10 ⁻³	5×10 ⁻⁵ -5×10 ⁻³	5×10 ⁻⁵ -5×10 ⁻³		
storage per unit leaf area	[8.81×10 ⁻⁵ -	[1.51×10 ⁻⁴ -	[4.00×10 ⁻⁴ -		
index (m)	0.0031]	0.0048]	0.0047]		
			-		
Albedo (-)	0.1-0.25	0.05-0.25	0.1-0.25		
	[0.11-0.25]	[0.062-0.24]	[0.11-0.24]		
Light attenuation coefficient	0.4-0.7	0.4-0.7	0.4-0.8		
(-)	[0.42-0.69]	[0.42-0.68]	[0.42-0.76]		

Emissivity (-)	0.9-0.99	0.9-0.99	0.9-0.99
	[0.91-0.99]	[0.90-0.99]	[0.90-0.98]
Channel			
Resistance to groundwater	$5 \times 10^{-3} - 0.5$		
seepage to channel (-)	[0.028-0.47]		
Manning's n	1-50		
	[8.2-47.5]		

Parameter	Value					Source
Soil	Brown earths	Humus- iron	Non - calcareous	Peaty- gleyed	Alluvial	
		podzol	gley	podzol		
Terrain random roughness (-)			0.05			Maneta and Silverman, 2013
Residual soil moisture (m³ m⁻³)	N	Min{0.05, 0.25	5 * Hydrologica	l layer porosit	y}	Local expertise
Soil to bedrock leakance (-)			0			Local expertise
Soil albedo			0.3			Maneta and Silverman (2013)
Soil emissivity			0.98			Maneta and Silverman (2013)
Heat capacity of dry soil (J m ³ K ⁻¹)	1.12×10 ⁶	1.19×10 ⁶	1.11×10 ⁶	1.20×10 ⁶	1.10×10 ⁶	Local expertise
Thermal conductivity of dry soil (W m ⁻¹ K ⁻¹)	0.137	0.125	0.138	0.124	0.140	Local expertise
Soil depth with negligible heat exchange (m)			2			Maneta and Silverman (2013)
Temperature at bottom thermal layer (°C)			10			Maneta and Silverman (2013)
Snowmelt coefficient (m s ⁻¹ °C ⁻¹)			4.1×10 ⁻⁸			Maneta and Silverman (2013)
Snow to rain temperature threshold (°C)			2			Maneta and Silverman (2013)
Vegetation	Grasses	Conifers	Heather			

Initial LAI (m ² m ⁻²)	2	2.9	1.6	Albrektson
muai LAI (III III)	2	2.)	1.0	(1984); Calder
				et al. (1984);
				Moors et al.
				(1998)
Stem density (m ⁻²) §	-	0.1	200	Local expertise
Vegetation age (years) §	-	30	7	Local expertise
Basal area (m ²) §	-	0.0195	1.3×10 ⁻⁵	Albrektson
				(1984); Wallén
				(1980)
Vegetation height (m)	0.5	10	0.4	Local expertise
Root mass (g m ⁻²)	1000	800	250	Aerts et al.
				(1989);
				Oleksyn et al.
				(1999)
NPP/GPP Ratio (-)	0.35	0.47	0.47	Lozano-Parra et
				al. (2014)
	1.0.10-6	20 10-6	1.0.106	
Canopy quantum	1.8×10^{-6}	3.0×10 ⁻⁶	1.8×10 ⁻⁶	Landsberg et al.
efficiency (gC J ⁻¹)				(2005)
Max forest age (years) §	-	500	40	Landsberg et al.
				(2005)
Leaf carbon allocation	_	2.235	2.235	Lozano-Parra et
coefficient a (-)§				al. (2014)
		0.004	0.004	
Leaf carbon allocation	-	0.006	0.006	Lozano-Parra et
coefficient b (-)§				al. (2014)
Stem carbon allocation	-	3.3	3.3	Lozano-Parra et
coefficient a (-)§				al. (2014)
Stem carbon allocation	_	3.0×10 ⁻⁷	3.0×10 ⁻⁷	Lozano-Parra et
coefficient b (-)§				al. (2014)
		0.44	0.04	
Wilting point (m ³ m ⁻³)	0.05	0.12	0.06	Maneta and
				Silverman
				(2013)
Specific leaf area (m ² g ⁻	2.3×10 ⁻²	8.0×10^{-3}	9.5×10 ⁻³	Landsberg et al.
1)				(2005); Poorter
				and De Jong
				(1999)

Specific root area (m^2 kg^{-1})	1.1×10 ⁻²	6.5×10 ⁻²	2.2×10 ⁻²	Kuppel et al. (2018)
Crown to stem diameter ratio (-)§	-	0.25	0.25	Landsberg and Waring (1997)
Tree shape coefficient (-)§	-	0.4	0.4	Landsberg and Waring (1997)
Wood density (gC m ⁻²) §	-	2.2×10 ⁵	2.2×10 ⁵	Landsberg and Waring (1997)
Maximum tree height to stem diameter ratio (-)§	-	15	150	Landsberg and Waring (1997); Wallén (1980)
Minimum tree height to stem diameter ratio (-)§	-	5	5	Landsberg and Waring (1997)
Leaf turnover rate (s ⁻¹)	3.17×10 ⁻⁸	1.20×10 ⁻⁸	1.00×10 ⁻⁸	Lozano-Parra et al. (2014); Maneta and Silverman (2013)
Max leaf turnover rate under water stress (s^{-1}) §	-	1.8×10 ⁻⁸	1.8×10 ⁻⁸	Landsberg and Waring (1997)
Sensitivity of leaf turnover rate to water stress (-)§	-	0.2	0.2	Landsberg and Waring (1997)
Max leaf turnover rate under temperature stress (s^{-1}) §	-	1.8×10 ⁻⁸	1.8×10 ⁻⁸	Landsberg and Waring (1997)
Sensitivity of leaf turnover rate to temperature (-)§	-	1.8×10 ⁻⁸	1.8×10 ⁻⁸	Landsberg and Waring (1997)
Cold temperature stress threshold (°C)§	-	1	1	Landsberg and Waring (1997)
Root turnover rate (s ⁻¹)	3.17×10 ⁻⁸	2.7×10 ⁻⁸	1.0×10 ⁻⁸	Landsberg and Waring (1997)
Canopy-scale water use efficiency (gC m ⁻¹)	800	680	3335	Gordon et al. (1999); Hobbie and Colpaert, (2004)

Empirical tree water use	-	0.6	0.6	Landsberg and
coefficient (-)§				Waring (1997)
Empirical tree water use	-	7	7	Landsberg and
exponent (-)§				Waring (1997)
Dry grass	8.5×10 ⁻⁷	-	-	Lozano-Parra et
decomposition rate (s ⁻¹)				al. (2014)
¢				
Temperature threshold	18	-	-	Lozano-Parra et
triggering dry grass				al. (2014)
decay (°C) ¢				

[§] Ligneous species only; [¢] Herbaceous species only

Appendix B: Sub-grid characterisation of channel cells in MAFIO

To enable sub-grid heterogeneity in MAFIO to be represented, it is necessary to specify several characteristics for cells containing an area of degraded soil / the channel. Figure S1 shows the Seepage and Channel IDs associated with such cells and their corresponding upslope contributing cells. The latter are used in deriving attributes of the spatial grid for the former. Channel widths, land parcels and livestock access associated with each cell are given in Table S3.

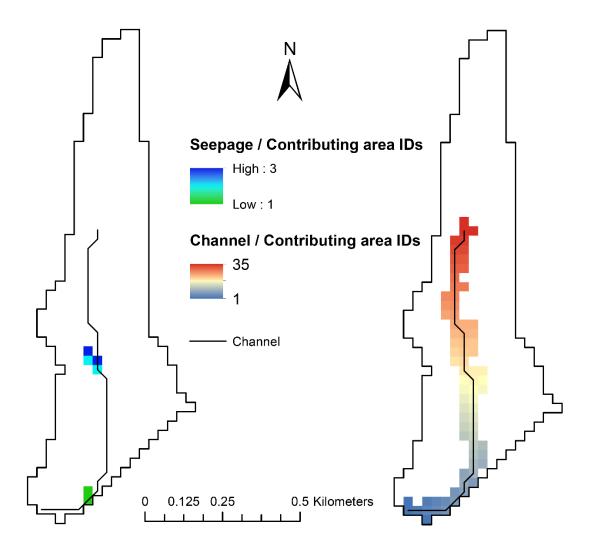


Figure S1: Maps showing a) The seepage IDs of cells with areas of degraded soil and their associated immediate upslope contributing areas; b) The channel IDs of cells containing a channel and their associated immediate upslope contributing areas. Cells containing the area of degraded soil / channel are those intersected by the "Channel" line.

Table S3: The properties of cells containing areas of degraded soil / the channel necessary for the representation of subgrid heterogeneity.

Channel ID	Seepage ID	Channel Width (m)	Land parcel(s)	Livestock Access
1	-	0.37	Lower Pasture (L) and (R)	No
2	-	0.37	Lower Pasture (L) and (R)	No
3	-	0.40	Lower Pasture (L) and (R)	No
4	-	0.45	Lower Pasture (L) and (R)	No
5	-	0.48	Lower Pasture (L) and (R)	No
6	1	0.51	Lower Pasture (L) and (R)	Gate*
7	-	0.55	Lower Pasture (L) and (R)	No
8	-	0.57	Lower Pasture (L) and (R)	No
9	-	0.62	Lower Pasture (L) and (R)	No
10	-	0.64	Lower Pasture (L) and Lower Arable	No
11	-	0.61	Lower Pasture (L) and Lower Arable	No
12	-	0.59	Lower Pasture (L) and Lower Arable	No
13	-	0.57	Lower Pasture (L) and Lower Arable	No
14	-	0.54	Lower Pasture (L) and Lower Arable	No
15	-	0.53	Lower Pasture (L) and Forest	No
16	-	0.52	Lower Pasture (L) and Forest	No
17	-	0.49	Lower Pasture (L) and Forest	No
18	-	0.43	Forest	No
19	-	0.37	Forest	No
20	2	0.32	Lower Pasture (L) and Mid Pasture (R)	Gate*

21	3	0.40	Mid Pastures (L) and (R)	Yes for (R); Gate* for (L)
22	-	0.48	Mid Pastures (L) and (R)	Yes for (R)
23	-	0.52	Mid Pastures (L) and (R)	Yes for (R)
24	-	0.50	Mid Pastures (L) and (R)	Yes for (R)
25	-	0.46	Mid Pastures (L) and (R)	Yes for (R)
26	-	0.44	Mid Pastures (L) and (R)	Yes for (R)
27	-	0.41	Mid Pasture (L)	Yes
28	-	0.35	Mid Pasture (L)	Yes
29	-	0.30	Forest	Yes
30	-	0.26	Forest	Yes
31	-	0.21	Forest	Yes
32	-	0.17	Forest	Yes
33	-	0.12	Forest	Yes
34	-	0.06	Forest	Yes
35	-	0.01	Forest	Yes

^{*} Variable livestock access depending on gate opening

1144 References

- 1145 Aerts, R., Berendse, F., Klerk, N. M. and Bakker, C., 1989. Root production and root turnover in two dominant
- species of wet heathlands, Oecologia 81(3): 374–378.
- Albrektson, A. (1984). Sapwood basal area and needle mass of Scots pine (Pinus sylvestris L.) trees in central
- 1148 Sweden. Forestry 57(1): 35–43.
- 1149 Calder, I.R., Hall, R.L., Harding, R.J., Wright, I.R., 1984. The use of a wet-surface weighing lysimeter system in
- rainfall interception studies of heather (Calluna vulgaris). Journal of Climate and Applied Meteorology
- **1151** 23(3): 461–473.
- Gordon, C., Woodin, S.J., Mullins, C.E., Alexander, I.J., 1999. Effects of environmental change, including
- drought, on water use by competing Calluna vulgaris (heather) and Pteridium aquilinum (bracken).
- 1154 Functional Ecology 13(s1): 96–106.
- Hobbie, E.A., Colpaert, J.V., 2004. Nitrogen availability and mycorrhizal colonization influence water use
- efficiency and carbon isotope patterns in Pinus sylvestris. New Phytologist 164(3): 515–525.
- Kuppel, S., Tetzlaff, D., Maneta, M.P., Soulsby, C., 2018. What can we learn from multi-criteria calibration of a
- process-based ecohydrological model? Environmental Modelling and Software 101: 301-316.
- Landsberg, J., Mäkelä, A., Sievänen, R., Kukkola, M., 2005. Analysis of biomass accumulation and stem size
- distributions over long periods in managed stands of Pinus sylvestris in Finland using the 3-PG model.
- 1161 Tree Physiology 25(7): 781–792.
- Landsberg, J.J., Waring, R.H., 1997. A generalised model of forest productivity using simplified concepts of
- radiation-use efficiency, carbon balance and partitioning. Forest Ecology and Management 95(3): 209–
- **1164** 228.
- Lozano-Parra, J., Maneta, M..P., Schnabel, S., 2014. Climate and topographic controls on simulated pasture
- 1166 production in a semiarid Mediterranean watershed with scattered tree cover. Hydrology and Earth
- 1167 Systems Science 18: 1439.
- Maneta, M.P., Silverman, N.L., 2013. A spatially distributed model to simulate water, energy, and vegetation
- dynamics using information from regional climate models. Earth Interactions 17(11): 1–44.
- Moors, E., Stricker, J. van der Abeele, G. (1998) Evapotranspiration of cut over bog covered by Molinia Caerulea.
- 1171 Wagenigen.
- Oleksyn, J., Reich, P.B., Chalupka, W., Tjoelker, M.G., 1999. Differential Above- and Below-ground Biomass
- 1173 Accumulation of European Pinus sylvestris Populations in a 12-year-old Provenance Experiment.
- Scandinavian Journal of Forest Research 14(1): 7-17.
- 1175 Poorter, H., De Jong, R.O.B., 1999. A comparison of specific leaf area, chemical composition and leaf
- construction costs of field plants from 15 habitats differing in productivity. New Phytologist 143(1): 163–
- **1177** 176.

Wallén, B., 1980. Structure and Dynamics of Calluna Vulgaris on Sand Dunes in South Sweden. Oikos 35(1): 20.