

1 **Opinion paper for “Agriculture, Ecosystems & Environment”**

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3 **Water sources for root water uptake: using stable isotopes of hydrogen and oxygen as a research**
4 **tool in agricultural and agroforestry systems**

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15

16 **Abstract**

17 Understanding water sources for crop water uptake in agricultural and agroforestry systems is an
18 essential step to develop more efficient and sustainable water management strategies, which is
19 increasingly important in the light of current world population growth, changing climatic conditions
20 and consequent growing pressures on agricultural- and agroforestry production. Stable isotopes of
21 hydrogen and oxygen in the water molecule are powerful and, nowadays, affordable tracers that can
22 help to define the proportion of water sources accessed by plants. Yet, contrary to natural
23 environments, their application is still relatively limited in agricultural and agroforestry research. In
24 this work, we synthesize the advantages and the current knowledge deriving from the use of the stable
25 isotopes of hydrogen and oxygen, in support of more traditional techniques, to understand root water
26 uptake dynamics in agricultural and agroforestry systems. We also underline the practical implications

27 related to the application of this technique for management purposes, and provide a vision for new
28 challenges and future research opportunities in exploring crop and plant water use based on isotopic
29 data.

30

31 **Keywords:** stable water isotopes, root water uptake, water sources, water resources management,
32 irrigation, agriculture and agroforestry systems

33

34 **1. Introduction**

35 Water is considered as the most critical resource for sustainable agricultural development worldwide.
36 Today, 70 % of total water withdrawals from surface water and groundwater resources are used in
37 the agricultural sector for irrigation (World Bank, 2017). Around 20 % of the total agricultural land
38 worldwide is irrigated, with the majority of existing irrigation systems still having low efficiency (FAO,
39 2019). As such, there is an urgent need for the management of water resources in agricultural and
40 agroforestry systems to deliver freshwater more efficiently to plants. The global population is
41 projected to grow to almost 10 billion by 2050 (UN, 2019). Worldwide there is an enormous challenge
42 to double food production between today and the middle of the century (World Resources Institute,
43 2019). This will, in all likelihood, have to be achieved with less water, as a growing population and
44 increasing urbanisation and industrialisation, especially in developing countries, compete for water
45 (WWAP, 2019). These pressures on global freshwater resources are exacerbated by the effects of
46 climate change. While water availability is becoming more challenging, irrigation is regarded as one of
47 the main means to increase crop yields. Therefore, it is imperative to improve water governance and
48 irrigation systems and, just as importantly, to expand the knowledge about demand and uptake
49 patterns of crops so that irrigation can either be avoided or can be applied more efficiently, allowing
50 a sustainable intensification of agriculture.

51

52 Key water sources for crops and agroforestry plants include soil moisture, groundwater and, where
53 needed, water applied by irrigation. Crop water requirement has been studied extensively in the past
54 decades (e.g., Shalhevet and Bielorai, 1978; Beyazgül et al., 2000; Kahlown et al., 2005; Senay and
55 Verdin, 2014; please also refer to the review by Pereira and Alves, 2013). Whereas total crop water
56 requirement may be known for individual plants, the respective proportions of the different water
57 sources that plants use over the growing period are known only for relatively few species in
58 agricultural and agroforestry systems (see examples and references in Section 2). Practical questions
59 on root water uptake arise here in the context of water resources management: Which sources do
60 crops use? How much water from each source do they take up? And how do water proportions change
61 over the growing season, as a function of developmental stage, weather or environmental conditions?
62 And also, how much of any irrigation water is actually being taken up by crops? These might look like
63 trivial questions for some, as plants might eventually take up any water that is accessible, no matter
64 what the source is. However, answering these questions would greatly help to predict adaptability to
65 changes in crops or water availability, or improve and adjust the timing and amount of irrigation water
66 being applied, and thus conserve freshwater resources and lower financial expenses.

67

68 With this opinion paper, we aim to demonstrate that the use of stable isotopes of water ($^2\text{H}/^1\text{H}$,
69 $^{18}\text{O}/^{16}\text{O}$) can be a very effective tool in agriculture and agroforestry systems to better understand the
70 temporal and spatial patterns of root water uptake (RWU) and, thus, to tackle water management-
71 related questions as stated above. We synthesize the current status of the application of stable
72 isotopes in agricultural and agroforestry research and provide a perspective on opportunities and
73 challenges for isotope-related research.

74

75 **2. Isotopes in agricultural and agroforestry systems**

76 Plants predominantly absorb water from the soil through their root system (Fig. 1). Water held by the
77 soil is mostly derived from precipitation and, in many agroecosystems, irrigation, but also infiltration

78 from nearby surface water and the capillary rise of groundwater. Moreover, water contained in
79 deeper soil layers can be absorbed and redistributed to shallower soil by deep roots. The various water
80 sources may have different isotopic composition according to their past history (Sprenger et al., 2019)
81 which makes isotopes excellent tracers to investigate root water uptake (Dawson et al., 2002; Barbour,
82 2007). In addition to the robust and well tested traditional isotope mass spectrometer, recent
83 technical advances have led to the commercialization of new, relatively cheap and transportable
84 instruments, which can measure the stable isotope composition of different waters, including
85 transpired and leaf water (Cernusak et al., 2016). As a result, analyzing stable isotopes in water has
86 become quick, relatively easy, and inexpensive, while results are reproducible and highly informative.
87 For these reasons, analyzing stable isotopes has become the method of choice for many
88 environmental researchers, such as, e.g., hydrologists, forest scientists, and ecophysicists; yet, in
89 comparison to more natural environments, this technique is still relatively sparsely applied in
90 agricultural and agroforestry research to address RWU.

91

92 The rationale is that the water in tree vessels is derived, assuming without fractionation (Dawson et
93 al, 2002), from roots after uptake and that the contributing water sources completely mix inside the
94 plant. This implies that it should be possible to estimate RWU by comparing the isotope values of the
95 potential water sources with that of the water extracted from the plant xylem (Fig. 1). Several
96 methods, recently summarized by Rothfuss and Javaux (2017) and Wang et al. (2019), have been
97 developed to this aim. The simplest method consists of estimating the mean RWU depth by directly
98 comparing the isotope value of xylem with the isotope value of water extracted from different soil
99 depths. It is also possible to estimate the proportion of potential water sources to xylem, e.g., by
100 applying end-member mixing analysis (EMMA, Christophersen and Hooper, 1992). One limiting factor
101 to successfully applying an EMMA is that the difference in isotope composition of potential water
102 sources must be considerably larger than the precision of the isotopic measurements. When there are
103 only two potential water sources, their relative contribution can be estimated by applying a linear

104 model based on classical statistics. For more complex situations with several potential water sources,
105 models based on Bayesian statistics have been developed (Parnell et al., 2013).

106

107 Stable isotopes of oxygen and hydrogen in the water molecule have been used in managed ecosystems
108 for different purposes. Typical applications include the identification of flow pathways and the
109 quantification of runoff components in agricultural catchments (Merot et al., 1995; Tekleab et al.,
110 2014; Tweed et al., 2016; Zajíček et al., 2016), the assessment of the seasonal role of precipitation and
111 irrigation water in recharging groundwater (Kaown et al., 2009; Brown et al., 2011), the computation
112 of residence time (Tekleab et al., 2014), the partitioning of evapotranspiration fluxes in soil
113 evaporation and plant transpiration components (Yepez et al., 2003; Williams et al., 2004; Wen et al.,
114 2016; Lu et al., 2017; Wu et al., 2017), the determination and characterization of contaminant sources
115 to rivers (Briand et al., 2017), the movement of fertilizers from soil to streams and groundwater (Parra
116 Suarez et al., 2019), and to a limited extent, the determination of RWU from different water sources.

117

118 *2.1. Water sources for plant water uptake*

119 A large variability of RWU patterns and water sources can exist, even within a single species. This
120 variability is typically a function of growing stage, seasonality, soil moisture, phenological stage, tree
121 size, transpiration rate, and irrigation and fertilization regimes (Schwendenmann et al., 2015; Ma and
122 Song, 2016; Bargaés Tobella et al., 2017; Hardanto et al., 2017; Liu et al., 2019). Specifically, most of
123 the research so far has focused on identifying the depth from which soil water is absorbed by plants
124 and its variability during the growing periods. For example, maize (*Zea mays*), across different
125 countries and various climatic and agronomic conditions, was typically found to access shallow soil
126 water, mainly in the first 20 cm of the soil and particularly during the seedling and jointing stage
127 (Asbjornsen et al., 2007 and 2008; Wang et al., 2010; Ma and Song, 2016; Zhao et al., 2016; Liao et al.,
128 2018). However, the primary soil water sources for maize normally vary with the growth stage, as
129 more water from deeper layers is used as the plants grow, and soil depths of 50 cm (Wang et al., 2010;

130 Ma and Song, 2016), 60 cm (Peng et al., 2017; Liu et al., 2018;), 80 cm (Zhang et al., 2011b), and, as
131 recently found, even 120 cm (Zhao et al., 2018a; Zhang et al., 2019) are reached.

132

133 Variability in RWU patterns as a function of the growing stage has also been reported for other crops,
134 both herbaceous and trees. For instance, the primary soil depths of RWU for winter wheat (*Triticum*
135 *aestivum*) were found to be between 0 and 50 cm for the majority of the growing period (Zhang et al.,
136 2011a; Wang et al., 2016; Yang et al., 2018), and especially in the 0-20 cm soil layers during the
137 wintering and seedling phases (Zhao et al., 2018a), and increasing depths were observed during the
138 end of the growing period (Cheng and Liu, 2017; Ma and Song, 2018) and up to 220 cm in the filling
139 stage (Guo et al., 2016). Similar trends were found for cotton (Li et al., 2017). Among trees, walnut
140 tree (*Juglans regia*) was reported to source water mainly in the 0-20 cm soil layer at the sprouting and
141 leaf expansion stages, and the 20-40 cm soil layer at blossoming and fruit-bearing, fruit expansion,
142 and fruit maturity stages (Liu et al., 2019), whereas cherry trees (*Prunus avium*) was found to take up
143 soil water from 20-50 cm at the fruit growth stage, then mainly 0-20 cm and 50-100 cm at the end of
144 the of growth period (Cao et al., 2018).

145

146 Switches between different soil water sources, especially for trees, can also occur as a function of
147 seasonality or different meteorological forcing, mostly in terms of dry vs. wet periods and high vs. low
148 soil moisture. Indeed, isotope-based observations in agro- and agroforestry ecosystems showed that
149 most trees are characterized by a plastic behaviour in water uptake depths, extracting water from
150 both shallow and deep soil layers but with a general tendency to access water in the first 50 or 60 cm
151 of the soil during wet conditions and switching to deeper soil water sources during dry periods when
152 the water content of shallow soil layers is low. This pattern was observed for walnut trees (Sun et al.,
153 2011), jujube trees (*Ziziphus jujube*) (Gao et al., 2018), rubber trees (*Hevea brasiliensis*) (Liu et al.,
154 2014; Wu et al., 2016; Hardanto et al., 2017; Wu et al., 2017), and indigenous West-African trees
155 (Smith et al., 1998; Bargués Tobella et al., 2017).

156

157 Variability in soil RWU depths and dynamics was also observed for crops of different age and,
158 especially for trees, variable size. Isotope studies reported different soil depths of RWU for 5-, 10-, 15-
159 , and 22-year-old apple trees (*Malus sp.*) in China, with soil depths contributions varying between 10
160 and 300 cm, and 22-year old apple trees mainly using water from the whole soil profile down to 160
161 cm during all growing stages (Wang et al., 2018; Zheng et al., 2018; 2019). Similarly, another study
162 carried out in China revealed that 4-year-old jujube trees mainly used soil water from 0-20 cm and 20-
163 60 cm layers, 8-year-old trees shifted flexibly their water sources between 0-20 cm, 20-60 cm, and 60-
164 200 cm layers, whereas 15- and 22-year-old trees often ignored moisture in shallow layers and
165 primarily used water from the deeper soils, especially when precipitation was low (Huo et al., 2018).
166 As for tree size, a clear relationship between stem diameter and soil water uptake depth was found
167 for rubber trees in cultivations in Sumatra, with bigger trees that tended to take up soil water closer
168 to the soil surface (Hardanto et al., 2017), although contrasting findings were obtained for native trees
169 (*Vitellaria paradoxa*) in West-Africa where the switch to deeper water sources was especially
170 pronounced for younger, smaller trees (Bargués Tobella et al., 2017).

171

172 Other factors leading to different soil sources for RWU are related to phenological stage and
173 transpiration rate, as showed by a study conducted in Panama that revealed a higher proportion of
174 deep water uptake being associated with a higher percentage foliage cover in the dry season, higher
175 sap flux densities, and water use rates (Schwendenmann et al., 2015). In case of rice, different RWU
176 patterns are typically related to the field management, e.g., continuous flooding versus alternate
177 flooding and drying (Shen et al., 2015; Mahindawansha et al., 2018).

178

179 Although precipitation-fed soil water is by far the primary source for plant transpiration, there are
180 cases in agricultural and agroforestry systems where isotopes helped to identify and quantify other
181 water sources, such as groundwater and irrigation. Groundwater is typically directly accessed only by

182 trees (Smith et al., 1997) and in some cases represents the dominant water source (e.g., the Australian
183 *Casuarina glauca*, Cramer et al., 1999). Most often, trees use soil water during the wet periods and
184 switch to groundwater only during the dry season (e.g., tagasaste tree, *Chamaecytisus proliferus*,
185 Lefroy et al., 2001, and the already mentioned *Vitellaria paradoxa*, Bargués Tobella et al., 2017).
186 Irrigation was identified and quantified using stable isotopes as water source for plant transpiration
187 in two cherry tree orchards in China with contributions declining below 11 % from the postharvest
188 stage in one case (Chao et al., 2018) and, conversely, being the highest (up to 22 %) during the drought
189 period in spring in another case (Li et al., 2019).

190

191 2.2. RWU under different irrigation and fertilization conditions

192 Irrigation and fertilization can largely affect RWU by cultivated plants, leading crops to access distinct
193 proportions of water sources or different proportions of water from various soil depths, especially
194 during dry periods (Goebel et al., 2015; Du et al., 2018). For instance, for maize, different depths of
195 RWU were observed under the application of distinct irrigation treatments, such as furrow irrigation,
196 border irrigation, and alternate furrow irrigation (Wu et al 2018a). In the study by Ma and Song (2016)
197 maize was fertilized with different nitrogen application rates from zero to 368 kg N ha⁻¹, resulting in
198 clear differences in the proportional contribution of soil water at different depths. Similarly,
199 fertilization with 105 kg N ha⁻¹ or irrigation of 20 mm during the greening-jointing stage significantly
200 promoted soil water uptake of winter wheat at 70-150 cm and 150-200 cm whereas other treatments
201 led to the much shallower RWU depths of 0-20 cm (Ma and Song, 2018).

202

203 Isotope-based analysis of distinct RWU patterns under different irrigation regimes and fertilization
204 treatments can thus be useful to provide insight and develop recommendations for fertilization and
205 irrigation management in agricultural and agroforestry systems. Observations of the seasonal and
206 growth stage variability of soil water sources used by maize and winter wheat in a rotation system in
207 Northern China suggested that irrigation events should be applied at a specific stage of the winter

208 wheat growth (Zhao et al., 2018a). Similarly, considerations of the RWU patterns of cherry trees in an
209 irrigated Chinese field led to the identification of the optimal water regulation treatment, resulting in
210 different irrigation quota at the end of the fruit growth stage, at the postharvest stage and during the
211 final part of the growth period (Cao et al., 2018). Later isotope work in the same area showed that the
212 drip irrigation mode demonstrated higher irrigation water use efficiency for cherry trees compared to
213 the surface irrigation mode (Li et al., 2019). In this context, isotopes can help to clarify the relationship
214 between root growth and irrigation method, and give practical, scientific-based indications to apply
215 ad-hoc irrigation strategies (e.g, alternate furrow irrigation, deficit irrigation) or irrigation methods
216 (e.g., pit irrigation systems) that contribute to plant water use efficiency and water saving mechanisms
217 (Wang et al., 2010; Wu et al., 2018a,b; Zheng et al., 2019).

218

219 2.3. Effects of co-cultivated or coexisting plants

220 Stable isotopes of hydrogen and oxygen are also useful tools to assess the possible competitive or
221 complementary RWU patterns of co-cultivated or coexisting plants. Evaluating the water use
222 strategies of co-cultivated or coexisting crops or trees is indeed useful to plan water management
223 interventions and agricultural practices. Various studies reported isotope-based evidence of similar
224 (and therefore competitive) or alternative (and therefore complementary) water use strategies in
225 different cultivates species. For example, complementary use of soil water resources through vertical
226 partitioning of water uptake was found in an agroforest stand with cocoa trees (*Theobroma cacao*)
227 and gliricidia trees (*Gliricidia sepium*) in Sulawesi, Indonesia (Schwendenmann et al., 2010). This
228 behaviour, in combination with acclimation, may help cocoa trees to cope with drought conditions.
229 Walnut tree (*Juglans regia* and *Juglans nigra*) was also reported to extract water from different soil
230 layers compared to other species: from deeper soils compared to the Italian alder (*Alnus cordata*) in a
231 mixed plantation in central Italy (Lauteri et al., 2006) but from shallower soils compared to a hybrid
232 poplar (*Populus deltoids* X *Populus nigra* clone) in an agroforestry system in Ontario, Canada (Link et
233 al., 2015). Poplar was also shown to be plastic in the depth of RWU and to have complementary water

234 use strategies compared to maize in an agroecosystem in Northern China (Liu et al., 2018). Other
235 complementary relations were found for the jujube tree with respect to annual and perennial crops
236 (Gao et al., 2018), for native trees in mixed species plots in Panama (Schwendenmann et al., 2015),
237 and for the rubber tree with tea, coffee and cocoa plants in Chinese agroforestry systems (Wu et al.,
238 2016; 2017). However, rubber tree was reported to have competitive displacement of RWU in a
239 plantation in Sumatra (Hartanto et al., 2017), indicating that water use strategies are not only species-
240 specific and might be also driven by local conditions. Furthermore, pronounced competitive RWU
241 patterns were observed in acacias (*Acacia nilotica* and *Acacia holosericea*) as they extracted large
242 quantities of water through lateral roots, and the competition was particularly severe at locations where
243 trees could not access groundwater (Smith et al., 1998). Competition was found for herbaceous crops
244 as well: an example from Northern Namibia in a pearl millet (*Pennisetum glaucum*) plantation
245 intercropped with cowpea (*Vigna unguiculata*) showed that the greater ability of cowpea to acquire
246 existing soil water forced pearl millet to develop deeper roots and shift to recently supplied water
247 (Zegada-Lizarazu et al., 2006). Other studies have reported low degrees of competition for water use
248 between herbaceous species and shrub and tree species due to the plasticity in RWU depths of the
249 latter (Asbjornsen et al., 2008). Some studies even reported contrasting findings, such as an alteration
250 of fine root distribution patterns of cocoa trees due to interspecific competition but, at the same time,
251 a not-significant limitation in their productivity (Rajab et al., 2018).

252

253 **3. Opportunities and challenges**

254 Despite the potential of using water isotopes for RWU analysis, some remaining methodological
255 questions still need to be considered. For example, this includes the validity of the assumption that no
256 fractionation occurs either during root water uptake (Barbeta et al., 2019; Poca et al., 2019) or
257 elsewhere in the subsurface, due, for example, to the interactions with soil clay content (Gaj et al.,
258 2017a; Meißner et al., 2014), to the effect of root exudates in the rhizosphere (Schwartz et al., 2015),
259 or to the activity of arbuscular mycorrhizal fungi (Coomans de Brachène, 2018). Furthermore, much

260 recent water isotope-based work has shown that there is an apparent distinction between ‘tightly-
261 bound’ and more mobile soil water (e.g. Oerter et al., 2019a; Sprenger et al., 2016), and that plants
262 might favour one over the other (McDonnell, 2014) or that differences reflect variations in spatio-
263 temporal patterns of soil water processes (Dubbert et al., 2019; Oerter et al., 2019b). This implies that
264 it is important to sample potential soil source water that is held across the variability of soil water
265 tensions and at multiple times. There are further challenges associated with the need for complete
266 extraction to avoid artificial effects of fractionation when soil and xylem water are extracted in the
267 laboratory for isotope analyses (Gaj et al., 2017b; Orłowski et al., 2018).

268

269 Another aspect that requires consideration is that isotope signatures in soil and xylem water may be
270 the product of any combination of source variation, mixing, and fractionation effects so that isotopic
271 values may not be exclusive to specific origins and processes (Benettin et al., 2018; Penna et al., 2018).
272 Such non-uniqueness can be largely addressed, for example via the use of Bayesian mixing models
273 that explore the uncertainties in mixing of various plant water sources (Rothfuss and Javaux, 2017;
274 Wang et al., 2019). Limitations can be further overcome by also considering nutrient availability to
275 further understand plant water and nutrient resource uptake and how this varies in time (Dubbert et
276 al., 2019). In this context, a critical point is the quantification of the uncertainty in the estimates of
277 sources for RWU that is missing from most of the studies so far conducted in agro- and agroforestry
278 ecosystems but that is of high importance in order to provide more realistic scenarios of RWU
279 dynamics and more reliable implementation of irrigation and fertilization treatments.

280

281 The key contribution that the analyses of stable isotopes of hydrogen and oxygen in agricultural and
282 agroforestry systems could make is to provide insights into the origin and the proportional water use
283 of different sources. Hence, the use of stable water isotopes to address questions on RWU is
284 complementary to and cannot fully replace the more traditional techniques that explore quantitative
285 water fluxes, nor can they fully complete our ecophysiological understanding (including, for example,

286 water use efficiency and other aspects of plant production). For a more complete picture of water-
287 plant-environment interactions and utilization, a combination with other available tools may be
288 employed. These include other stable isotopes, particularly ^{13}C for understanding utilisation of water
289 resources (e.g. Cernusak et al., 2013; Rumman et al., 2018) and ^{15}N for nutrient uptake (e.g. Kulmatiski
290 et al., 2017). Nevertheless, addressing RWU questions with knowledge on proportional source water
291 use can have important practical implications for the management of agricultural and agroforestry
292 systems. These could include the improvement of local to regional farming practices such as irrigation
293 or fertilization treatment schemes (e.g. Du et al., 2018; Ma and Song, 2018). Strategic planning at
294 longer timescales would also benefit from insights into the ability of plants to access different sources,
295 especially under changing hydro-climatological conditions such as extreme weather events, droughts
296 or altered flow regimes (e.g. Sun et al., 2011; Gao et al., 2018), or whether co-existent species have
297 competitive or complementary water use strategies (e.g. Muñoz-Villers et al., 2019). Especially for
298 agricultural and agroforestry regions which are sustained at the natural limits of water availability,
299 understanding the implications of climate or management change could be essential, for example in
300 terms of evaluating the need for irrigation, which in turn is vital for designing and implementing
301 efficient and equitable farming programs.

302

303 Most of the current isotope-based studies in agricultural and agroforestry regions involve RWU in
304 maize. Moreover, a high proportion of these studies were carried out in the semi-arid regions of North
305 China. These involve applications where RWU questions are clearly pressing and where irrigation
306 forms an important water source. However, many opportunities lay in extending these isotope-based
307 RWU studies to other geographic and climatic regions and a wider range of crops, especially as climate
308 change may affect the ratio between rain-fed and irrigation-supported agriculture. RWU patterns and
309 responses are known to vary in space and time (e.g., Geris et al., 2017) and between vegetation types
310 (e.g., Zegada-Lizarazu and Iijima, 2004; Wu et al., 2018). Hence, beyond practical local applications,
311 increased understanding of RWU sources across a broader range of conditions will also enable new

312 scientific insights into the wider functioning of plants (including plasticity or adaptability to change)
313 which are typical for agricultural and agroforestry environments, especially as previous research has
314 focussed mainly on natural systems. As such, extending our knowledge on RWU across different
315 ecosystems will provide new opportunities for addressing fundamental ecohydrological questions
316 with implications of global relevance.

317

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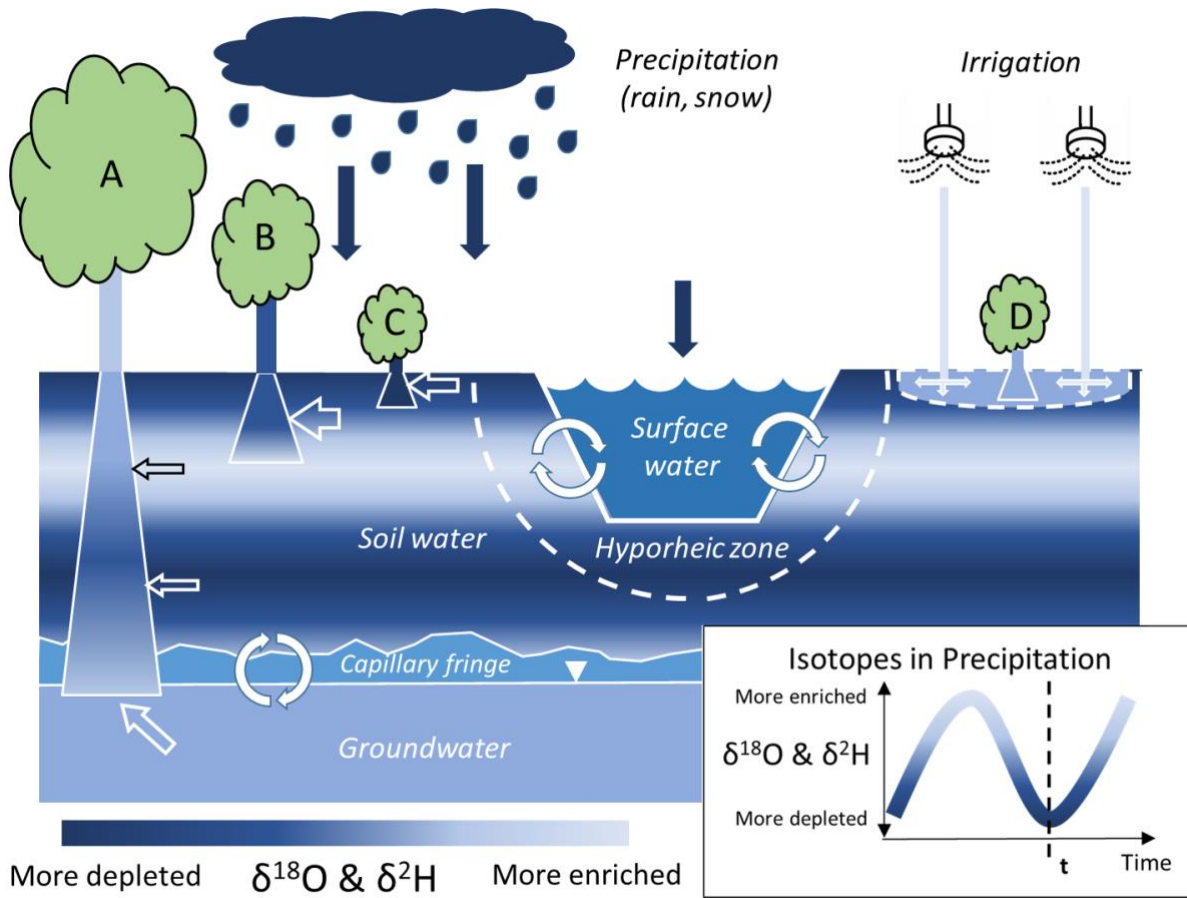
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730 Fig. 1. Conceptual diagram of stable water isotope signatures in different root water sources
 731 (precipitation, soil water at different depths, surface- and groundwater, and irrigation water) at time
 732 t. The lower right inset shows that precipitation isotopic signatures change with time, e.g. seasonally
 733 (after Allen et al., 2019). The isotopic composition of water sources and crop water are indicated by
 734 colours, with more enriched values in light and more depleted values in dark blue. Subsurface arrows
 735 indicate from where crops might take up water, with arrow thickness reflecting the proportion of each
 736 water source; white circular arrows refer to active hydrological mixing zones. Depending on different
 737 factors (see text at Section 2.1), the crop roots access different water sources, and the isotopic value
 738 of various crops (e.g., herbaceous, shrubs, and trees, A-D) can be different at one specific moment in
 739 time.