Energetics and thermal adaptation in semifossorial pine-voles Microtus lusitanicus

and Microtus duodecimcostatus

Rita I. Monarca<sup>1\*</sup>, John R. Speakman<sup>2,3</sup> and Maria da Luz Mathias<sup>1</sup>

<sup>1</sup>CESAM – Center for Environmental and Marine Studies, Departamento de Biologia

Animal, Faculdade de Ciências da Universidade de Lisboa, Lisbon, Portugal

<sup>2</sup> Institute of Biological and Environmental Sciences, University of Aberdeen, Tillydrone

Ave, Aberdeen, Scotland, UK,

<sup>3</sup> Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, 1

West Beichen Road, Chaoyang, Beijing, China

\*Corresponding author:

Corresponding author: Rita I. Monarca

CESAM – Center for Environmental and Marine Studies, Departamento de Biologia

Animal, Faculdade de Ciências da Universidade de Lisboa, Lisboa, Portugal

Tel: +351 217500000 ext 22303

Fax: +351 217500028

Email: rimonarca@fc.ul.pt

Keywords:

Doubly labelled water; Resting metabolic rate; Water turnover, digging energetics

1

## Abstract

Rodents colonizing subterranean environments have developed several morphological, physiological and behaviour traits that promote the success of individuals in such demanding conditions.

Resting metabolic rate, thermoregulation capacity and daily energy expenditure were two fossorial pine-vole species Microtus lusitanicus M. duodecimcostatus inhabiting distinct areas of the Iberian Peninsula. Individuals were captured in locations with different habitat and soil features, allowing the comparison of energetic parameters with ecological characteristics, that can help explain the use of the subterranean environment and dependence of the burrow system. Results showed that M. duodecimcostatus has lower mass independent resting metabolic rate when compared with M. lusitanicus, which may be a response to environmental features of their habitat, such as dryer soils and lower water availability. Thermal conductance increased with body mass and was dependent on the ambient temperature. No significant differences were observed in the DEE, but water economy data demonstrated the influence of the water available on the habitat on the energetics of voles. These may rely on behavioural adaptations and seasonal use of burrows to cope which thermal challenges of subterranean activity and soil constraints. We found strong evidence that M.lusitanicus is able to use torpor as response to low ambient temperatures which is a new observation among Arvicolines.

#### Introduction

Energy metabolism has been extensively studied as a method to approach physiological adaptations to environmental variations (Mueller and Diamond 2001; Lovegrove 2003; Rezende et al 2004; Bozinovic et al 2007), as it represents the summed result of all functions simultaneously occurring in the whole animal, is ecologically relevant to define food requirements (Speakman 2000) and potential for heat stress (Speakman and Król 2010). The resting metabolic rate (RMR) represents a large portion of an animals' energetic requirement comprising 20 to 60% of the daily energetic demands (Ricklefs et al 1996; Hammond and Diamond 1997; Speakman 2000; Westerterp and Speakman

2008), therefore can be highly informative about the adaptive strategies of different species (Morgan and Price 1992; Bennett et al 1994) and populations (Bozinovic et al 2007; Castellanos-Frías et al 2015).

In this study we compared the energetics of two sister species the Lusitanian pine vole (*Microtus lusitanicus* Gerbe,1879) and the Mediterranean pine vole (*Microtus duodecimcostatus* de Selys-Longchamps, 1839).

The M.lusitanicus occurs in the northern most side of the Iberian peninsula, while M. duodecimcostatus inhabits the south eastern part of the Iberian Peninsula and southern France, the species share a small portion of their distribution area across the central area of the Iberian peninsula (Aulagnier et al 2008; Aulagnier and Palomo 2008). Although genetically closely related (Jaarola et al 2004), overall M. lusitanicus has smaller size than M. duodecimcostatus (Microtus Iusitanicus, body mass 14-19g; M. duodecimcostatus, body mass 19-32g). Broadly, pine voles can be found in meadows, pastures, agricultural areas and orchards where they can often cause severe damage (Vinhas 1993; Cotilla and Palomo 2007; Mira and Mathias 2007). However, at the micro scale species exhibit different ecological preferences (Borghi et al 1994; Santos et al 2011) and clear differences in their burrowing behaviour (Giannoni et al 1993), M.lusitanicus pushes earth from burrows mostly using the hindlegs and M.duodecimcostatus uses the incisors to break the soil and pushes it outwards using the head. Soil properties have been reported as a key factor in shaping the morphology of subterranean rodents (Stein 2000) and influencing the energetic demands of burrowing (Luna and Antinuchi 2006). Therefore, we analysed soil properties and compare them with energetic parameters to analyse the response of each species to their environmental conditions. We hypothesised that M.duodecimcostatus exhibits reduced RMR compared with M.lusitanicus given that inhabits warmer and dryer areas which influences the properties of the inhabited soil. Thus, soils with higher water content facilitate heat dissipation allowing animals to sustain elevated metabolic rates. Moreover, we measured the energy expenditure of free-living voles and compare it with captive voles to evaluate the costs of inhabit a subterranean environment.

#### Material and methods

# Field sampling and maintenance of voles

Field work was carried out across the species ranges from the north to the south of Portugal (N  $37^{\circ}00 - 41^{\circ}50$ ; W  $6^{\circ}30 - 9^{\circ}20$ ). The selection of trapping sites was dependent of the presence of fresh mounds, holes or burrows, mainly in agricultural areas, road verges and meadows. Voles were captured using modified tube traps to include a nesting box, partially filled with local grass and bated with apple. The number of traps per site varied in accordance with the extension of the colony, estimated by the number of signs such as mounds and tunnel entrances. Trapping was successful in 18 sites within the range of *M. lusitanicus* and in 30 sites within the range of *M. duodecimcostatus*. Soil samples were collected in the 48 sites where voles were captured. Samples were collected in a radius of 1 meter from the trapping point. Approximately 1 dm³ of soil was removed and kept in a closed plastic bag until being processed.

Although a larger number of voles have been trapped, only 46 non-reproductive adults (include split by species) were used in the present study. Experimental animals were housed in small individual cages (255 x 220 mm) partially filled with soil collected in capture sites. The majority of voles (40 out of 46) were taken to the laboratory where they were kept under controlled light (12L:12D) and room temperature ( $\approx 25$ °C), with free access to water and fed with carrots, apples and grass *ad libitum* The remaining 6 individuals were used in field measurements of energy expenditure (see below).

# Respirometry

A total of 24 voles (M. lusitanicus: 5 females, body mass  $17.1g \pm 0.2g$  and 6 males, body mass  $16.4g \pm 0.4g$ ; M. duodecimcostatus: 7 females, body mass  $24.5g \pm 0.3g$  and 6 males, body mass  $25.5g \pm 0.4g$ ), were used to assess energy expenditure and thermoregulatory abilities. Energetic measurements were carried out after a 4 week period for laboratory acclimation. Oxygen consumption (VO2) was measured using an open-circuit respirometry system (Servomex, series 1100), as previously described by Duarte (2010). Animals were measured in a cylindrical chamber (approximately 1000 ml capacity), and dried atmospheric air was pumped into the chamber at a flow rate of 500ml/min.

Carbon dioxide was not removed to minimise error in the conversion of oxygen consumption to energy expenditure (Koteja 1996). Each vole was monitored twice, over two consecutive days, for 2 hours each day, in different periods, to avoid potential effects of daily metabolic cycle (Halle and Stenseth 1994). The average of the two measurements was use in the data analysis. Voles were not fasted prior to entering the chamber, but no food or water were available during the 2h-experiments. Consequently at the end of the runs voles had not been fed for at least 2 hours.

Measurements of oxygen consumption were recorded at 15s intervals, using the *Labtech* data acquisition and process control software. This procedure was repeated at different ambient temperatures: 5°C, 15°C, 20°C, 25°C, 30°C, 32.5°C, 35°C, 37.5°C and 40°C. A total of 358 measurement runs were performed, but 10 of these were excluded from analysis because the voles did not settle in the chamber. At higher ambient temperature, if animals showed signs of distress, they were removed from the chamber and the temperature registered as the upper survival limit. To correct for machine drift baseline, values of atmospheric oxygen were measured for 15 minutes before and after each trial.

At each ambient temperature, resting metabolic rate was estimated as the average value of the ten lowest consecutive readings (equivalent to 2min30s in the chamber) (Hayes et al 1992). At the beginning and end of each run, animals were weighed and body temperature measured rectally (at depth around 2.5 cm), with a thermocouple K probe (MI-K-Miniz-1.0-100) connected to a *Digitron* thermometer (2088T, Sifam Instruments Limited).

Oxygen consumption was calculated after Depocas and Hart (1957) as  $VO_2=V_2$  ( $F_1O_2-F_2O_2$ )/(1- $F_1O_2$ ), where  $V_2$  is the flow rate measured after the metabolic chamber, and  $F_1O_2$  and  $F_2O_2$  are the oxygen concentrations before and after the metabolic chamber. All  $VO_2$  measurements were corrected to standard temperature and pressure (STPD).

Thermal conductance was calculated as  $C=VO_2/(T_b-T_a)$ , where C is conductance,  $T_b$  is the body temperature of the animal and  $T_a$  the ambient temperature (McNab 1970).

## **Doubly Labelled Water**

The Doubly Labelled Water technique (DLW) (Butler et al 2004) was used to determine the Daily Energy Expenditure (DEE) and the Water Turnover (WTO), estimating how hard animals are working through the calculation of sustained metabolic scope (SusMs = DEE/RMR). WTO is considered as a balance between internal fluids and the input and output of water from the external environment (Speakman and Racey 1988).

A total of 22 animals (13 females and 9 males) were used in the DLW experiments. Three groups of voles were injected with doubly labelled water (30% <sup>18</sup>O, <sup>2</sup>H). The first group included six *M. lusitanicus* (4 females and 2 males), captured during the summer period in an apple orchard and released in their natural environment in the field. We failed to successfully recapture any individuals of *M. duodecimcostatus* and hence have no field metabolic rate measurements for this species. Accordingly we compared the DEE of two further groups measured in captive conditions. The second group included eight *M. lusitanicus* (5 females and 3 males) captured in the same site where the field measurements had been made, but taken to the laboratory, housed in individual cages, kept under natural light conditions and fed *ad libitum* with a mixture of fresh grass and carrots. The third group included eight *M. duodecimcostatus* (4 females and 4 males), captured in an orange orchard and kept under the same laboratory conditions.

One hour after the injection of the DLW, a blood sample was taken by retro orbital bleeding, to estimate the initial isotope enrichment of <sup>2</sup>H and <sup>18</sup>O. Blood samples were flame sealed in glass capillaries immediately after being collected. After the blood collection, the individuals from the first group were released in the same place of capture. The individuals from the second and third groups were kept in the laboratory, in individual cages, as described above. After 24h, a second blood sample was taken, to evaluate the final isotope enrichment (Speakman and Racey 1988). In the field, the blood samples were collected from 6 recaptured animals (from a total of 8 previously captured and injected). To assess the correct amount of injected isotope, the syringes were weighed before and after the administration of the water (0.0001g, Sartorius 4-figure balance).

The capillaries were vacuum distilled (Nagy 1983) and water from the resulting distillate was used to produce CO<sub>2</sub> and H<sub>2</sub> (methods in Speakman et al. (1990)and Speakman and

Król (2005). The isotope ratios <sup>2</sup>H: <sup>1</sup>H and <sup>18</sup>O: <sup>16</sup>O were then analysed using gas source isotope ratio mass spectrometry. The elimination constants ( $K_0$  and  $K_d$ ) and the dilution spaces ( $N_0$  and  $N_d$ ) of the ratios <sup>2</sup>H and <sup>18</sup>O injected were determined. Initial and final pools were calculated by the plateau method (Speakman et al 1993). This method assumes that the initial blood sample was taken when the administered isotope has equilibrated with water pool of the animal's body, and reached a maximal value before the isotope has been washed out from the body, according with the following equation (Król and Speakman 1999):  $N_i = M_{inj} (E_{peak} - E_{inj})/(E_{bg} - E_{peak})$ , where  $N_i$  (mol) is the dilution space for deuterium, oxygen-18 or tritium; Minj is the amount of injectate (mol) injected; Epeak is the initial isotope enrichment (ppm) of body water; Einj is the enrichment (ppm) of the deuterium/oxygen-18 or tritium injectate; Ebg is the background isotope enrichment (ppm) of body water. CO2 production was estimated according with equation rCO2=(N/2.078) x ( $K_0$  -  $K_d$ ) – 0.0062  $K_d$ N (Speakman, 1997, eq. 7.17), where rCO2 is the CO2 production, N (mol) is the size of body water pool. The rate of CO2 production was converted to DEE assuming a respiratory quotient of 0.85 and oxygen equivalent to 20.1kJ.L<sup>-1</sup>.

The WTO (ml/day) values were calculated using the deuterium elimination rates ( $K_d$  per day), and deuterium dilution spaces ( $N_d$ , ml), WTO =  $K_d$ .  $N_d$ .F, where F is the fractionation factor of the isotope (0.941; Speakman 1997). The Water Economy Index (WEI) was calculated was WTO/DEE. Sustained metabolic scope was calculated as SusMs=DEE/RMR (Peterson et al 1990).

All experimental procedures were conducted in the University of Lisbon facilities by an expert in laboratory animal science accredited by the Portuguese Veterinary Authority (1005/92, DGV-Portugal, following FELASA category C recommendations), according to the European guidelines (86/609/EEC).

# Data analysis

Resting metabolic rates and thermal conductance were analysed using a mixed model procedure, setting species and ambient temperature as factors, body mass as covariate

and individual id as random factor to account for repeated measures (methods in Tschöp et al (2011)). Thermal conductance (C) calculated from the individual measurements of oxygen consumption using the equation C=VO<sub>2</sub>/(Tb-Ta) (Mcnab 1980). Limits of thermoneutrality were determined for each species using a segmented regression model through SegReg software (www.waterlog.info). A General linear model was used to analyse DEE, WTO, WEI and SusMs data, setting species and environment (field vs lab) as fixed factors and body mass as covariate. All values were expressed as mean ± S.E. and p< 0.05 was taken as statistically significant, data were analysed using SPSS v19 for windows.

Soil samples were analysed for soil texture, percentage of organic matter, and water availability. The mineral contents of the soil were separated according with their sizes, and classified into three major classes: clay, silt and sand. Texture designates the relative proportion of each class in a sample; USDA soil textural classes were adopted (Soil Survey Division Staff 1993). The samples from the three main texture categories, were processed to the determine water availability and percentage of organic matter in the soil. Water availability was assessed by the difference between field capacity and permanent willing point (assessed by pF 2.14 and pF 4.2) (Richards 1947). Differences between sites were then compared using One-way ANOVA, setting species as fixed factor.

# Results

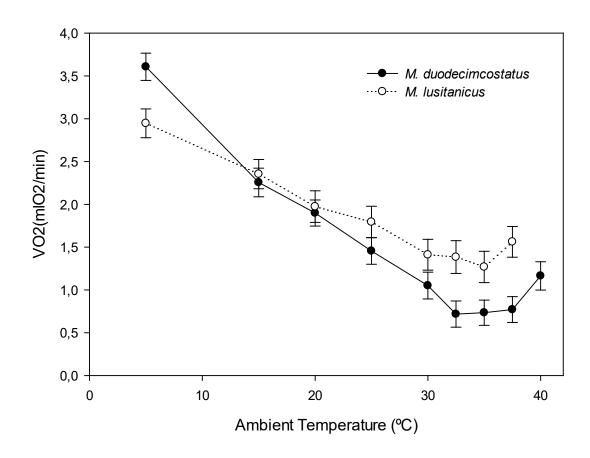
## Resting Metabolic Rate and Thermoregulation

Mean body temperature of *M.lusitanicus* ranged from 35.8±0.39°C at Ta=5°C to 40.1±0.51°C at Ta=37.5°C. At lower ambient temperatures some animals reduced their body temperature: six individuals had body temperatures between 30 and 33 °C and one individual had a body temperature of 26 °C (Figure 2A). The mean body temperature for *M. duodecimcostatus* ranged from 38.1±0.36°C at Ta=5°C to 42.3±0.55°C at Ta=40°C (Figure 2B). Upper survival limit was registered at 37.5°C for *M. lusitanicus* and at 40°C for *M. duodecimcostatus*.

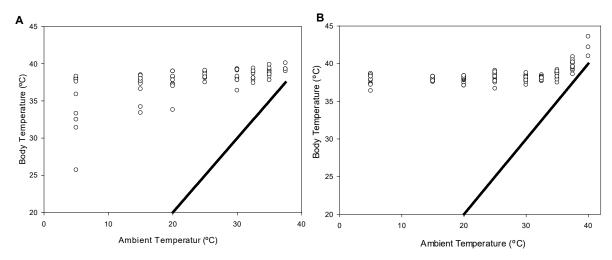
Resting metabolic rate variation was explained by the body mass ( $F_{1,21}$  =29.944; p<0.001), ambient temperature ( $F_{8,21}$  =49.860; p<0.001) and the interaction between ambient temperature and species ( $F_{8,21}$  =4.172; p<0.001). Overall, *M.duodecimcostatus* had lower metabolic rates than *M. lusitanicus*, except when ambient temperature was 5°C. At 5°C some *M.lusitanicus* reduced their metabolic rate by 68% when comparing all the individuals at 5°C and body temperature fell to between 26 and 33 °C (see above), suggesting the induction of a shallow torpor.

The thermoneutral zone was calculated for *M.duodecimcostatus* to be between 31.6°C and 37.5°C, for *M. lusitanicus* lower critical point was calculated as 30.7°C, and due to reduced data upper critical temperature was estimated 35 – 37.5°C.

Below thermoneutrality thermal conductance was fairly stable at 0.101 mlO<sub>2</sub>.min<sup>-1</sup>. $^{\circ}$ C<sup>-1</sup> for *M.lusitanicus* and 0.131 mlO<sub>2</sub>.min<sup>-1</sup>. $^{\circ}$ C<sup>-1</sup> *M.duodecimcostatus*. Above lower critical point thermal conductance increased about 186% to 0.287 mlO<sub>2</sub>.min<sup>-1</sup>. $^{\circ}$ C<sup>-1</sup> in *M.lusitanicus* and to 0.430 mlO<sub>2</sub>.min<sup>-1</sup>. $^{\circ}$ C<sup>-1</sup> (227%) in *M.duodecimcostatus*. Thermal conductance was mostly influenced by the ambient temperature (F<sub>8,20</sub> =33.102; p<0.001) and the body mass (F<sub>1,20</sub>=12.396; p=0.001).



**Figure 1** – Oxygen consumption of *Microtus duodecimcostatus* and *M. lusitanicus* across ambient temperature. (Adjusted values for BM= 21,53g).



**Figure 2** - Individual values of body temperature  $(T_b)$  in relation to ambient temperature  $(T_a)$  ( $-T_b = T_a$ ). A – Microtus lusitanicus; B – Microtus duodecimcostatus.

# **Soil characteristics**

The analysis of soil samples revealed that M.lusitanicus was mostly found on sandy-loam (41%), silty-loam (23%), loam (18%) and clay loam soils (14%), whereas M.duodecimcostatus occupied mainly loam (35%), sandy-loam (28%), silty-loam soils (17%), and other residual classes (20%). Figure 3 summarises data on water availability and % of organic matter on the three texture classes more abundant on both species. The soil from locations inhabited by M.lusitanicus have significant higher organic matter content (ANOVA  $F_{2,39}$ =35.283; p<0.001) and water available (ANOVA  $F_{2,38}$ =45.390; p<0.001) then those inhabited by M.duodecimcostatus.

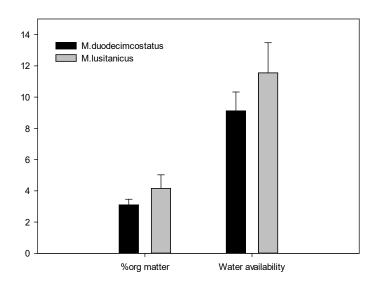


Figure 3 - % of organic matter and water availability on the sandy-loam, silty loam and loam soils, in sites inhabited by *M. duodecimcostatus* (dark) and *M.lusitanicus* (light).

# **Daily Energy Expenditure and Water Flux**

All the variables examined had no significant effects on the DEE and SusMs. The data are summarised in Table I.

The WTO (ml. day<sup>-1</sup>) was significantly higher in the field animals ( $F_{1,17}$ =18.256; p=0.001). The WEI was also significantly different between animals from the field and from the lab ( $F_{1,17}$ =22.264; p<0.001).

**Table I** – Mean ± standard error of the mean for Water turnover (WTO), daily energy expenditure (DEE), Sustained Metabolic Scope (SusMS) and Water economy index (WEI) (all estimated for body mass = 20.34g).

	M.lusitanicus		M.duodecimcostatus
	Field	Laboratory	Laboratory
N	6	7	8
WTO (ml.day <sup>-1</sup> )	3.04 ± 0.31	1.57 ± 0.25	1.52 ± 0.32
DEE (kJ.day <sup>-1</sup> )	57.0 ± 6.47	55.2 ± 5.37	52.9 ± 6.82
WEI (ml. kJ <sup>-1</sup> )	0.97 ± 0.07	0.58 ± 0.06	0.55 ± 0.07
SusMS	2.10 ± 0.23	1.87 ± 0.19	2.28 ± 0.24

#### Discussion

The extreme demands of subterranean life have resulted in unique specialisations that allow them to cope with the burrow environment. Reduced metabolic rates has been described as one of the features that allows subterranean species to avoid overheating due to the elevated costs of burrowing activity (Vleck 1979; Bozinovic et al 2005).

The measurements of oxygen consumption showed clear differences between the metabolic rates of the two studied species, *M.duodecimcostatus* had reduced resting metabolic rates, comparing with *M. lusitanicus*. According with hypothesised such variation can be interpreted as response to environmental features, such us soil dryness and availability of water.

Our data show evidence for the occurrence of torpor periods in the *M. lusitanicus*. Torpor is a common strategy in several rodent species as the house mouse *Mus musculus* (Overton and Williams 2004), deermice *Peromyscus sp.* (Tannenbaum and Pivorun 1987) and the Djungarian hamster *Phodopus sungorus* (Ruf et al 1991). However, torpor have never been reported in Arvicoline rodents (McNab 1992; Nieminen et al 2013). Voles rely on constant food availability, and are not adapted to prolonged periods of fasting, perishing after 6 to 26 hours without food (Mustonen et al 2008). Thus is unlikely that torpor in *M. lusitanicus* is induced by the scarcity of

resources. However, the expression of torpor occurred only in individuals with reduced body mass, observations on mice (Rikke et al 2003; Mitchell et al 2015) suggested that body composition may be involved in the mechanisms regulating torpor induction and body temperature.

On this study, torpor occurs as response to the exposure to reduced temperatures, hence possibly voles developed such strategy to save energy and cope with winter temperatures. Our study did not consider the circadian variation of body temperature (Refinetti and Menaker 1992), however the measured range of body temperature variation (30%) gives us confidence to suggest the occurrence of torpor bouts. Other Microtine species also reduced their body temperature in response to cold acclimation, but the variation range is reduced, about 5% on the *M. cabrerae* (Mathias et al 2003) and *M. arvalis* (Ishii et al 1996), not entering in a torpid state

Several studies (Mcnab 1979; Lovegrove 1986) have suggested that the adaptations towards a life more dependent on the subterranean environment includes a reduction of resting metabolic rates due to overheating risk and elevated costs of subterranean foraging. Moreover, previous studies comparing digging behaviour (Giannoni et al 1993) and morphology (Mathias 1990; Mathias 1996) of *M.duodecimcostatus* and *M.lusitanicus* suggested that *M.duodecimcostatus* developed features that are in line with an adaptation to subterranean environment whilst *M.lusitanicus* showed adaptive traits towards surface dwelling.

Following Ebensperger and Bozinovic (2000) and Lovegrove (1989) average metabolic rates of species digging in dry, harder soils are expected to be greater than those of species digging in moist, softer soils due to the higher cost of burrowing. In fact, the soil texture, water content, organic matter and tillage are parameters that also highly constrain the thermal conductivity of the soil (Abu-Hamdeh and Reeder 2000; Abu-Hamdeh 2003). Soil thermal conductivity increases with the soil water content, thus soils with higher water content also increase the capacity of heat dispersion.

This supports the hypothesis that high metabolic rates are more sustainable in environments that facilitate heat dispersion, potentially explaining the higher RMR of *M.lusitanicus* which inhabits moister soils than *M.duodecimcostatus*. Moreover, the percentage of organic matter also facilitates the water retention on soil (Farley et al.)

2004). This may partly explain why the abundance of pine voles is usually higher in agricultural areas (Vinhas 1993; Mira and Mathias 1994; Miñarro et al 2012). In fact, the permanent watering of soil, through dripping, is a common agronomic practice that facilitates the expansion of vole populations (Bertolino et al 2015), and in some cases it has been suggested to be a key factor explaining rodent outbreaks (Jareno et al 2015) and continuous reproduction throughout the year (Ventura et al 2010).

The importance of water as the driver of vole's behaviour was also made evident by the measured WTO rates. Some studies suggested that reduced water turnover is a strategy to save energy (Rubal et al 1995; Scantlebury et al 2003) in desert environments, moreover WTO is linked to the energy expenditure when water is not available for drinking.

Voles under field conditions had higher values of WTO and WEI, which are indicative of the higher water intake than captive voles. Generally, subterranean species, as many other species, do not drink free water (Buffenstein 2000), thus our data suggests that free-living voles may have access to water sources, such as food. Water economy data also suggests some phenotypic plasticity that should be investigated in further studies.

The absence of differences in DEE between the field group and the housed group were unexpected, it may be explained by the abundance of resources in the field area (agricultural area with dripping watering and vegetative cover), and an absence of intensive burrow digging during the measurement period.

Considering the present results, our data supports previous observations proposing seasonal fluctuation of burrowing activity, during the summer animals are less active (Guedon et al 1992) as during winter grass cover is abundant, animals move mainly on the surface, where food is available and can be stored for scarcity periods (Mira and Mathias 1994). The climate in the burrows is highly buffered, however the atmosphere inside the tunnels can be influenced by the environment on the surface above (Burda et al 2007). Thus underground activity (excluding active burrowing) may be preferred during the summer, because helps avoiding elevated temperatures of the dry season.

Deeper soil layers maintain constant levels of moisture which may contribute to heat dispersal (Kinlaw 1999; Burda et al 2007). Seasonal and daily variations in burrowing

activity were already reported in other species, *e.g* semifossorial *Octodon degus* (Ebensperger and Bozinovic 2000) and in *Spalacopus cyanus* (Rezende et al 2003; Urrejola et al 2005). Even though subterranean rodents can modify their digging behaviour according with soil texture, using only the forelimbs to dig in loose soils, and including the use of the incisors when help in required to break some rocks (Lessa and Thaeler Jr. 1989). Shifting the activity patterns according to the ambient temperatures, creating a seasonal routine, may allow voles to cope with the thermal constrains of burrowing during the dry season.

## Conclusion

The data obtained sustained our initial hypothesis that mass-independent RMR is reduced in the *M.duodecimcostatus* (considering ambient temperatures above 15°C) which is consistent with the occurrence of the species in drier and warmer environments, typical of the south of the Iberian Peninsula. On the other hand, *M.lusitanicus* may have developed strategies to cope with low temperatures through torpor in line with the colonisation of the north of the Iberian Peninsula.

These are new insights on the species their energetic demands which provides interesting information about their biology and the complex speciation process between the two species.

## **Acknowledgements**

Acknowledgments are due to FCT/MEC for the financial support to CESAM RU (UID/AMB/50017) through national funds, when applicable co-financed by the FEDER, within the PT2020 Partnership Agreement. RIM was supported by fellowship BPD/UI88/7346/2016 and JRS was supported by the 1000 talents program of the Chinese government.

#### References

- Abu-Hamdeh NH (2003) Thermal Properties pf Soils as affected by Density and Water Content. Biosyst Eng 86:97–102. doi: 10.1016/S1537-5110(03)00112-0
- Abu-Hamdeh NH, Reeder RC (2000) Soil Thermal Conductivity: Effects of Density, Moisture, Salt Concentration, and Organic Matter. Soil Sci Soc Am J 64:1285–1290.
- Aulagnier S, Amori G, Hutterer R, et al (2008) Microtus lusitanicus. IUCN Red List Threat Species 2008 e.T13494A4039347. doi: http://dx.doi.org/10.2305/IUCN.UK.2008.RLTS.T13494A4039347.en
- Aulagnier S, Palomo L. (2008) Microtus duodecimcostatus. IUCN Red List Threat Species 2008 e.T13493A4037759. doi: http://dx.doi.org/10.2305/IUCN.UK.2008.RLTS.T13493A4037759.en
- Bennett NC, Aguilar GH, Jarvis JUM, Faulkes CG (1994) Thermoregulation in three species of Afrotropical subterranean mole-rats (Rodentia: Bathyergidae) from Zambia and Angola and scaling within the genus Cryptomys. Oecologia 97:222–227.
- Bertolino S, Asteggiano L, Saladini MA, et al (2015) Environmental factors and agronomic practices associated with Savi's pine vole abundance in Italian apple orchards. J Pest Sci (2004) 88:135–142. doi: 10.1007/s10340-014-0581-7
- Borghi CE, Giannoni SM, Martínez-Rica JP (1994) Habitat segregation of three sympatric fossorial rodents in the Spanish Pyrenees. Zeitschrift fur Saugetierkd 59:52–57.
- Bozinovic F, Carter MJ, Ebensperger LA (2005) A test of the thermal-stress and the cost-of-burrowing hypotheses among populations of the subterranean rodent Spalacopus cyanus. Comp Biochem Physiol 140:329–336. doi: 10.1016/j.cbpb.2005.01.015
- Bozinovic F, Muñoz JLP, Cruz-Neto AP (2007) Intraspecific Variability in the Basal Metabolic Rate: Testing the Food Habits Hypothesis. Physiol Biochem Zool 80:452–460.
- Buffenstein R (2000) Ecophysiological responses of subterranean rodents to underground habitats. In: Lacey EA, Patton JL, Cameron GN (eds) Life Undergr. Biol. Subterr. rodents. University of Chicago Press, Chicago, pp 62–110
- Burda H, Sumbera R, Begall S (2007) Microclimate in burrows of subterranean Rodents Revisited. In: Begall S, Burda H, Schleich CE (eds) Subterr. Rodents News from Undergr. Springer-Verlag, Berlin, pp 21–33
- Butler PJ, Green J a., Boyd IL, Speakman JR (2004) Measuring metabolic rate in the field: the pros and cons of the doubly labelled water and heart rate methods. Funct Ecol 18:168–183. doi: 10.1111/j.0269-8463.2004.00821.x
- Castellanos-Frías E, García-Perea R, Gisbert J, et al (2015) Intraspecific variation in the energetics of the Cabrera vole. Comp Biochem Physiol Physiol Part A Mol Integr 190:32–38. doi: 10.1016/j.cbpa.2015.08.011
- Cotilla I, Palomo LJ (2007) Microtus duodecimcostatus (de Sélys-Longchamps, 1839). In: Palomo LJ, Gisbert J, Blanco JC (eds) Atlas y Libr. rojo los Mamíferos Terr.

- España. Dirección General para la Biodiversidad-SECEM-SECEM, Madrid, pp 423–425
- Depocas F, Hart JS (1957) Use of the paulling oxygen analyzer for mesurement of oxygen consumption of animals in open-circuit systems and in a short-lag, closed circuit apparatus. J Appl Physiol 10:388–392.
- Duarte LC, Vaanholt LM, Sinclair RE, et al (2010) Limits to sustained energy intake XII: is the poor relation between resting metabolic rate and reproductive performance because resting metabolism is not a repeatable trait? J Exp Biol 213:278–87. doi: 10.1242/jeb.037069
- Ebensperger LA, Bozinovic F (2000) Energetics and burrowing behaviour in the semifossorial degu Octodon degus (Rodentia: Octodontidae). J Zool 252:179–186. doi: 10.1017/S0952836900009912
- Farley KA, Kelly EF, Hofstede RGM (2004) Soil organic carbon and water retention after convertion of grasslands to pine plantations in Ecuadorian Andes. Ecosystems 7:729–739.
- Giannoni SM, Borghi CE, Martínez-Rica JP (1993) Comparing the burrowing behaviour of the Iberian mole voles (Microtus (Terricola) lusitanicus, M. (T.) pyrenaicus and M. (T.) duodecimcostatus). Mammalia 57:483–490.
- Guedon G, Paradis E, Croset H (1992) Capture-recapture study of a population of the Mediterranean Pine vole (Microtus duodecimcostatus) in Southern France. Zeitschrift fur Saugetierkd 57:364–372.
- Halle S, Stenseth NCHR (1994) Microtine ultradian rhythm of activity: an evaluation of different hypotheses on the triggering mechanism. Mamm Rev 24:17–39.
- Hammond KA, Diamond J (1997) Maximal sustained energy budgets in humans and animals. Nature 386:457–462.
- Hayes JP, Speakman JR, Racey PA (1992) SamplingBias in Respirometry. Physiol Zool 65:604–619.
- Ishii K, Kuwahara M, Tsubone H, Sugano S (1996) The telemetric monitoring of heart rate, locomotor activity, and body temperature in mice and voles (Microtus arvalis) during ambient temperature changes. Lab Anim 30:7–12. doi: 10.1258/002367796780744992
- Jaarola M, Martínková N, Gündüz I, et al (2004) Molecular phylogeny of the speciose vole genus Microtus (Arvicolinae, Rodentia) inferred from mitochondrial DNA sequences. Mol Phylogenet Evol 33:647–63. doi: 10.1016/j.ympev.2004.07.015
- Jareno D, Vinuela J, Luque-Larena JJ, et al (2015) Factors associated with the colonization of agricultural areas by common voles Microtus arvalis in NW Spain. Biol Invasions 17:2315–2327. doi: 10.1007/s10530-015-0877-4
- Kinlaw A (1999) A review of burrowing by semi-fossorial vertebrates in arid environments. J Arid Environ 41:127–145.
- Koteja P (1996) Measuring energy metabolism with open-flow metric systems: which design to choose? Funct Ecol 10:675–677.

- Król E, Speakman JR (1999) Isotope dilution spaces of mice injected simultaneously with deuterium, tritium and oxygen-18. J Exp Biol 202:2839–2849.
- Lessa EP, Thaeler Jr. CS (1989) A reassessment of morphological specializations for digging in pocket gophers. J Mammal 70:689–700. doi: 10.2307/1381704
- Lovegrove BG (2003) The influence of climate on the basal metabolic rate of small mammals: a slow-fast metabolic continuum. J Comp Physiol B 173:87–112. doi: 10.1007/s00360-002-0309-5
- Lovegrove BG (1986) The metabolism of social subterranean rodents: adaptation to aridity. Oecologia 69:551–555.
- Lovegrove BG (1989) The Cost of Burrowing by the Social Mole Rats (Bathyergidae) Cryptomyys damarensis and Heterocephalus glaber: The role of Soil Moisture. Physiol Zool 62:449–469.
- Luna F, Antinuchi CD (2006) Cost of foraging in the subterranean rodent Ctenomys talarum: effect of soil hardness. Can J Zool Can Zool 84:661–667. doi: 10.1139/Z06-040
- Mathias M da L (1990) Morphology of the incisors and the burrowing activity of Mediterranean and Lusitanian pine voles (Mammalia, Rodentia). Mammalia 54:302–306.
- Mathias M da L (1996) Skull size variability in adaptation and speciation of the semifossorial pine voles Microtus duodecimcostatus and M. luistanicus (Arvicolidae, Rodentia). Proc. I Eur. Congr. Mammal. Lisboa, pp 271–286
- Mathias ML, Klunder M, Santos SM (2003) Metabolism and thermoregulation in the Cabrera vole (Rodentia: Microtus cabrerae). Comp Biochem Physiol Part A Mol Integr Physiol 136:441–446.
- Mcnab BK (1980) On Estimating Thermal Conductance in Endotherms. Physiol Zool 53:145–156.
- Mcnab BK (1979) The Influence of Body Size on the Energetics and Distribution of Fossorial and Burrowing mammals. Ecology 60:1010–1021.
- McNab BK (1970) Body weight and the energetics of temperature regulation. J Exp Biol 53:329–348.
- McNab BK (1992) The comparative energetics of rigid endothermy the Arvicolidae. J Zool 227:585–606.
- Miñarro M, Montiel C, Dapena E (2012) Vole pests in apple orchards: use of presence signs to estimate the abundance of Arvicola terrestris cantabriae and Microtus lusitanicus. J Pest Sci (2004) 85:477–488. doi: 10.1007/s10340-012-0438-x
- Mira A, Mathias MDL (2007) Microtus lusitanicus (Gerbe, 1879). In: Palomo LJ, Gisbert J, Blanco JC (eds) Atlas y Libr. rojo los mamífers Terr. España. Dirección General para la Biodiversidad-SECEM-SECEM, Madrid, pp 418–421
- Mira A, Mathias ML (1994) Conditions controlling the colonization of an orange orchard by Microtus duodecimcostatus (Rodentia, Arvicolidae). Pol Ecol Stud 20:249–255.

- Mitchell SE, Delville C, Konstantopedos P, et al (2015) The effects of graded levels of calorie restriction: III. Impact of short term calorie and protein restriction on mean daily body temperature and torpor use in the C57BL/6 mouse. Oncotarget 6:22–28.
- Morgan KR, Price M V (1992) Foraging in Heteromyid Rodents: The Energy Costs of Scratch-Digging. Ecology 73:2260–2272.
- Mueller P, Diamond J (2001) Metabolic rate and environmental productivity: well-provisioned animals evolved to run and idle fast. Proc Natl Acad Sci U S A 98:12550–4. doi: 10.1073/pnas.221456698
- Mustonen A-M, Saarela S, Nieminen P (2008) Food deprivation in the common vole (Microtus arvalis) and the tundra vole (Microtus oeconomus). J Comp Physiol B 178:199–208.
- Nagy KA (1983) The Doubly Labeled Water (3HH18O) Method: A guide to its use., UCLA Publi. University of California, Los Angeles, California, USA
- Nieminen P, Hohtola E, Mustonen A-M (2013) Body temperature rhythms in Microtus voles during feeding, food deprivation, and winter acclimatization. J Mammal 94:591–600. doi: 10.1644/12-MAMM-A-219.1
- Overton JM, Williams TD (2004) Behavioral and physiologic responses to caloric restriction in mice. Physiol Behav 81:749–54. doi: 10.1016/j.physbeh.2004.04.025
- Peterson CC, Nagy KA, Diamond J (1990) Sustained metabolic scope. Proc Natl Acad Sci USA 87:2324–2328.
- Refinetti R, Menaker M (1992) The circadian rhythm of body temperature. Physiol Behav 51:613–637. doi: 10.2741/3634
- Rezende EL, Bozinovic F, Garland TJ (2004) Climatic adaptation and the evolution of basal and maximum rates of metabolism in rodents. Evolution (N Y) 58:1361–1374.
- Rezende EL, Cortés A, Bacigalupe LD, et al (2003) Ambient temperature limits above-ground activity of the subterranean rodent Spalacopus cyanus. J Arid Environ 55:63–74. doi: 10.1016/S0140-1963(02)00259-8
- Richards LA (1947) Pressure-membrane apparatus, construction and use. Agron Eng 28:451–454.
- Ricklefs RE, Konarzewski M, Daan S (1996) The Relationship between basal metabolic rate and daily energy expenditure in birds and mammals. Am Nat 147:1047–1071.
- Rikke BA, Yerg JE, III, et al (2003) Strain variation in the response of body temperature to dietary restriction. Mecha 124:663–678. doi: 10.1016/S0047-6374(03)00003-4
- Rubal A, Haim A, Choshniak I (1995) Resting metabolic rates and daily energy intake in desert and non-desert murid rodents. Comp Biochem Physiol Part A Physiol 112:511–515. doi: 10.1016/0300-9629(95)02020-9
- Ruf T, Klingenspor M, Preis H, Heldmaier G (1991) Daily torpor in the Djungarian hamster (Phodopus sungorus): interactions with food intake, activity, and social behaviour. J Comp Physiol B 160:609–615. doi: 10.1007/BF00571257

- Santos SM, Mathias MDL, Mira AP (2011) The influence of local, landscape and spatial factors on the distribution of the Lusitanian and the Mediterranean pine voles in a Mediterranean landscape. Mamm Biol 76:133–142. doi: 10.1016/j.mambio.2010.03.007
- Scantlebury M, Shanas U, Speakman JR, et al (2003) Energetics and water economy of common spiny mice Acomys cahirinus from north- and south-facing slopes of a Mediterranean valley. Funct Ecol 17:178–185. doi: 10.1046/j.1365-2435.2003.00717.x
- Speakman J, Nagy K, Masman D, et al (1990) Interlaboratory comparison of different analytical techniques for the determination of oxygen-18 abundance. Anal Chem 62:703–708. doi: 10.1021/ac00206a011
- Speakman JR (2000) The Cost of Living: Field Metabolic Rates of Small Mammals. Adv Ecol Res 30:177–297.
- Speakman JR (1997) Doubly Labelled Water: Theory and Practice. Kluwer Academic Publishers, New York
- Speakman JR, Król E (2010) Maximal heat dissipation capacity and hyperthermia risk: neglected key factors in the ecology of endotherms. J Anim Ecol 79:726–46. doi: 10.1111/j.1365-2656.2010.01689.x
- Speakman JR, Król E (2005) Comparison of different approaches for the calculation of energy expenditure using doubly labeled water in a small mammal. Physiol Biochem Zool PBZ 78:650–667.
- Speakman JR, Nair KS, Goran MI (1993) Revised equations for calculating CO2 production from doubly labeled water in humans. Am J Physiol 264:E912–E917.
- Speakman JR, Racey PA (1988) The doubly-labelled water technique for measurement of energy expenditure in free-living animals. pp 227–237
- Stein BR (2000) Morphology of subterranean rodents. In: Lacey EA, Patton JL, Cameron GN (eds) Life Undergr. Biol. Subterr. rodents. The University of Chicago Press, Chicago, pp 19–61
- Tannenbaum MG, Pivorun EB (1987) Differential Effect of Food Restriction on the Induction of Daily Torpor in Peromyscus-Maniculatus and Peromyscus-Leucopus. J Therm Biol 12:159–162.
- Tschöp MH, Speakman JR, Arch JRS, et al (2012) A guide to analysis of mouse energy metabolism. Nat Methods 9:57–63. doi: 10.1038/nmeth.1806
- Urrejola D, Lacey EA, Wieczorek JR, Ebensperger LA (2005) Daily activity patterns of free-living cururos (Spalacopus cyanus). J Mammal 86:302–308.
- Ventura J, Jiménez L, Gisbert J (2010) Breeding characteristics of the Lusitanian pine vole, Microtus lusitanicus. Anim Biol 60:1–14. doi: 10.1163/157075610X12610595764011
- Vinhas A (1993) Microtus lusitanicus (rato cego) e Microtus duodecimcostatus (rato toupeira) roedores pragas das culturas. Rev Ciências Agrárias XVI:375–382.
- Vleck D (1979) The energy cost of burrowing by the pocket gopher Thomomys bottae.

Physiol Zool 52:122–136.

Westerterp KR, Speakman JR (2008) Physical activity energy expenditure has not declined since the 1980s and matches energy expenditures of wild mammals. Int J Obes 32:1256–1263.